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The Director

of the United States Patent and Trademark Office has received an application for a patent for a new and useful invention. The title and description of the invention are enclosed. The requirements of law have been complied with, and it has been determined that a patent on the invention shall be granted under the law.

Therefore, this United States

Patent

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Katherine Kelly Vidal



DIRECTOR OF THE UNITED STATES PATENT AND TRADEMARK OFFICE

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Patent Term Notice

If the application for this patent was filed on or after June 8, 1995, the term of this patent begins on the date on which this patent issues and ends twenty years from the filing date of the application or, if the application contains a specific reference to an earlier filed application or applications under 35 U.S.C. 120, 121, 365(c), or 386(c), twenty years from the filing date of the earliest such application (“the twenty-year term”), subject to the payment of maintenance fees as provided by 35 U.S.C. 41(b), and any extension as provided by 35 U.S.C. 154(b) or 156 or any disclaimer under 35 U.S.C. 253.

If this application was filed prior to June 8, 1995, the term of this patent begins on the date on which this patent issues and ends on the later of seventeen years from the date of the grant of this patent or the twenty-year term set forth above for patents resulting from applications filed on or after June 8, 1995, subject to the payment of maintenance fees as provided by 35 U.S.C. 41(b) and any extension as provided by 35 U.S.C. 156 or any disclaimer under 35 U.S.C. 253.



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- (*) Notice: Subject to any disclaimer, the term of this patent is extended or adjusted under 35 U.S.C. 154(b) by 0 days.

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(57) **ABSTRACT**

The invention relates to the production and use of Cas-encoding sequences and vectors comprising these. Aspects of the invention provide products, vectors, delivery vehicles, uses and methods for producing Cas-encoding sequences in bacterial or archaeal cells.

26 Claims, 6 Drawing Sheets

Specification includes a Sequence Listing.

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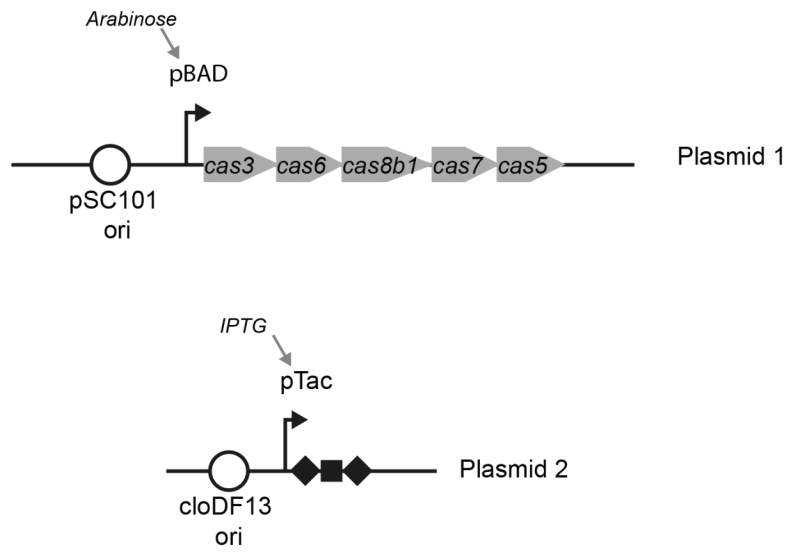


FIG. 1A

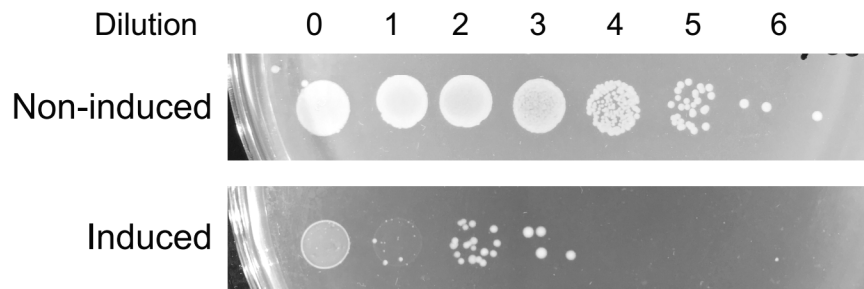


FIG. 1B

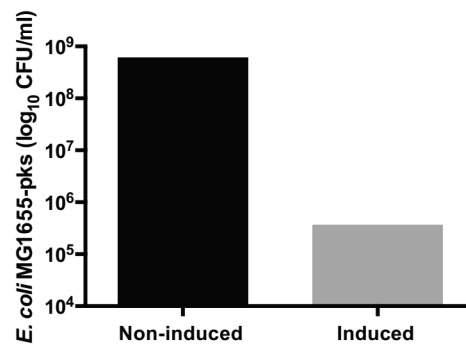


FIG. 1C

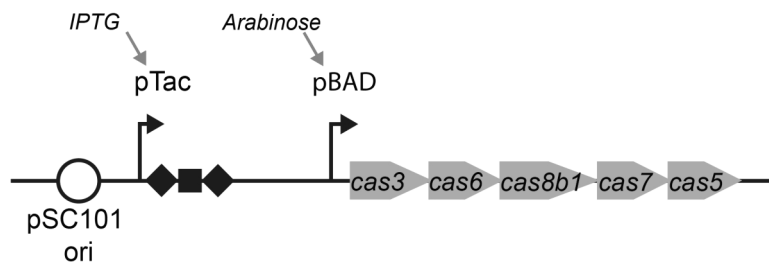


FIG. 2A

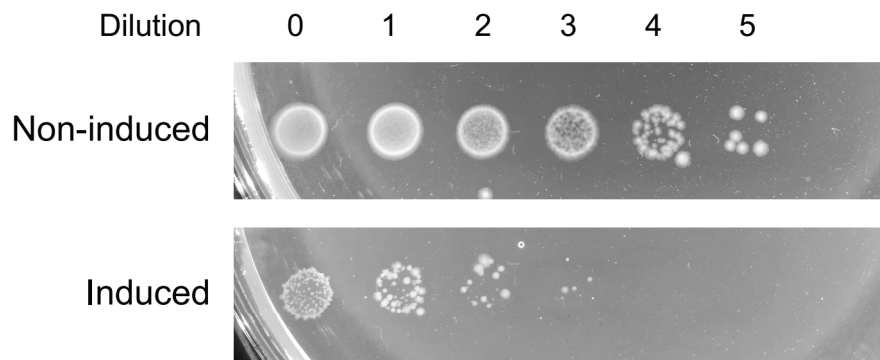


FIG. 2B

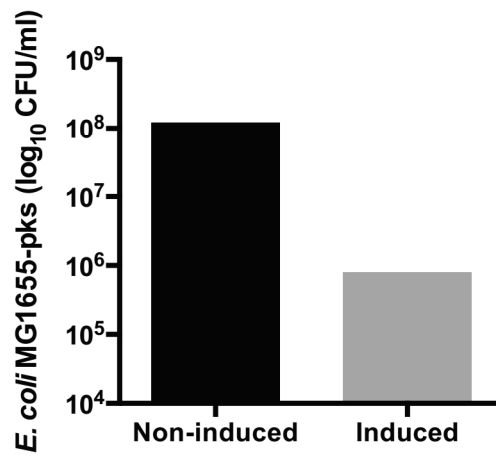


FIG. 2C

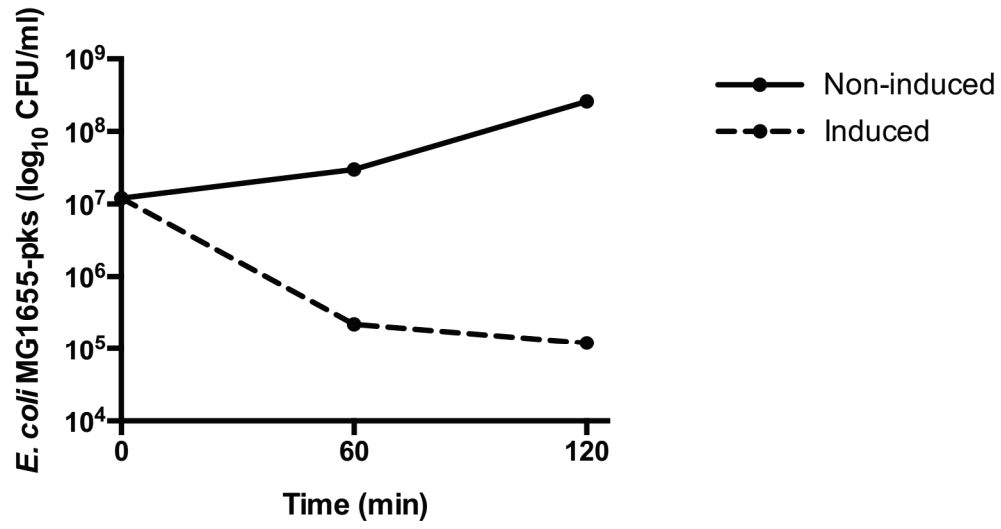


FIG. 3A

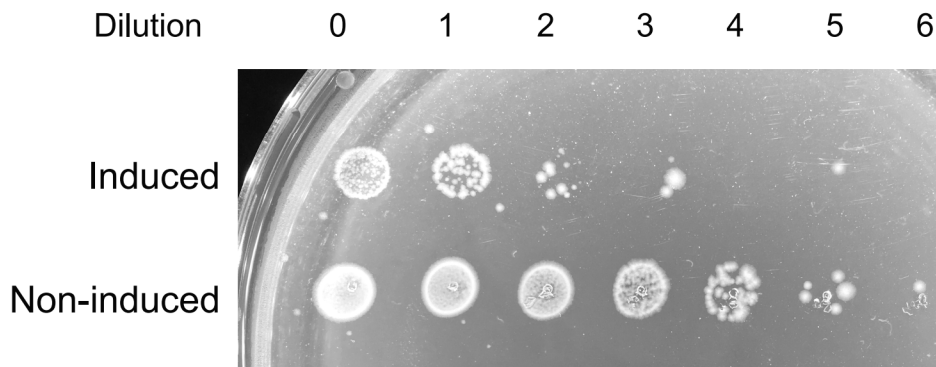


FIG. 3B

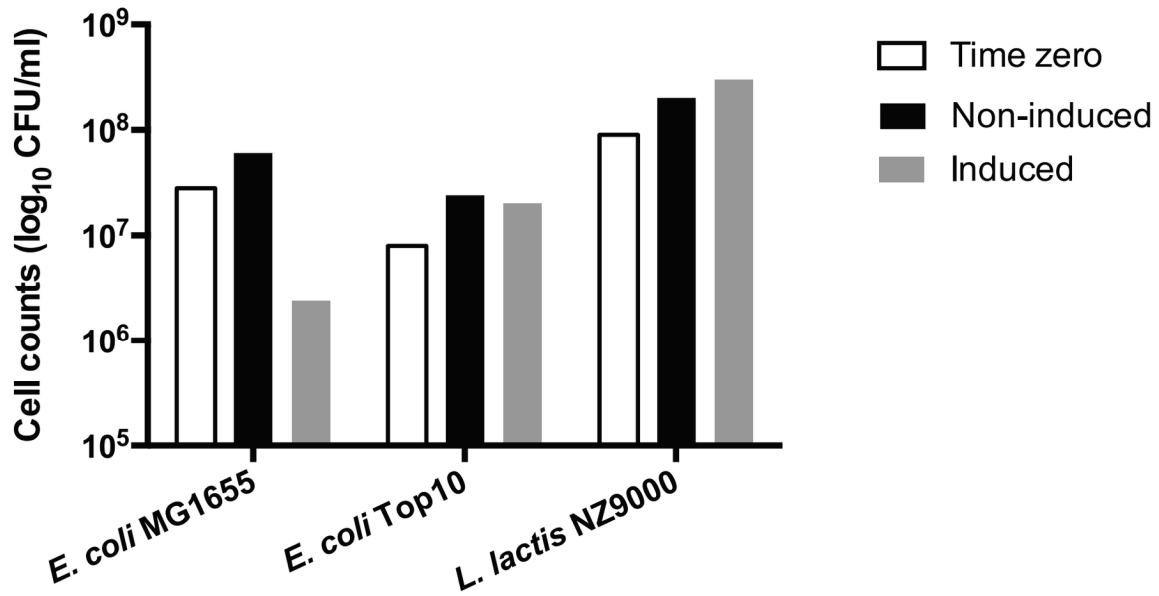


FIG. 4A

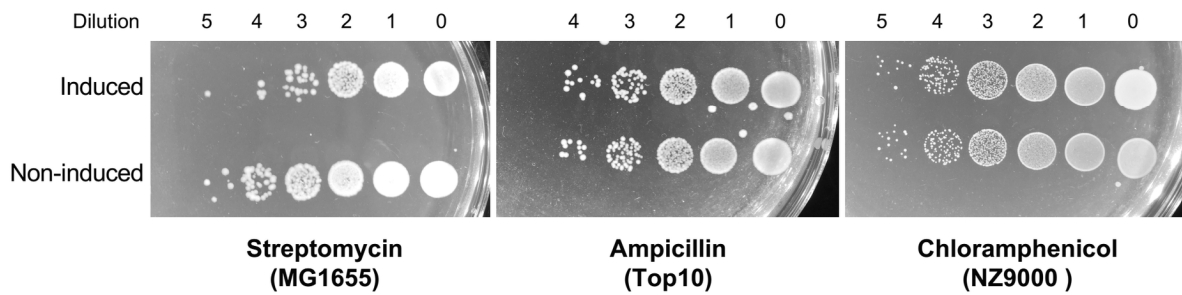


FIG. 4B

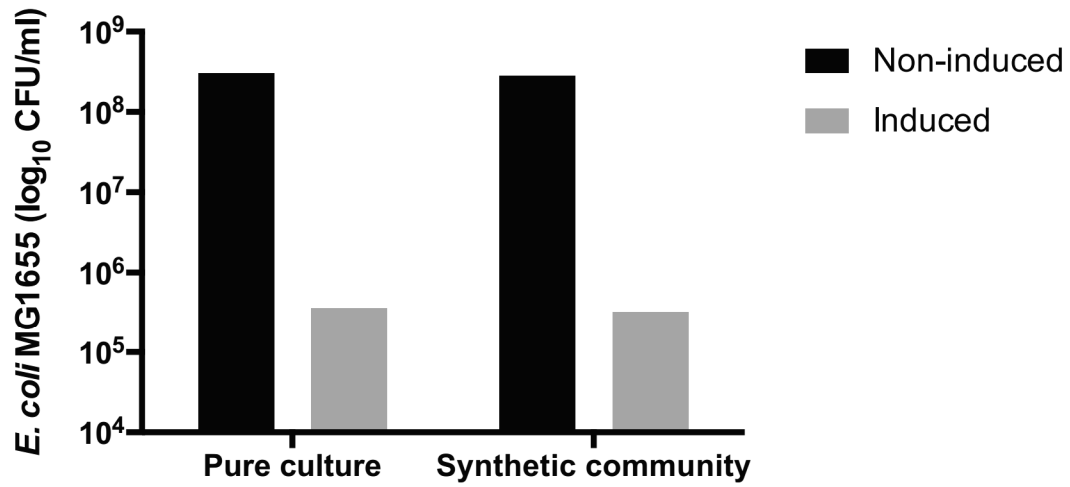


FIG. 5A

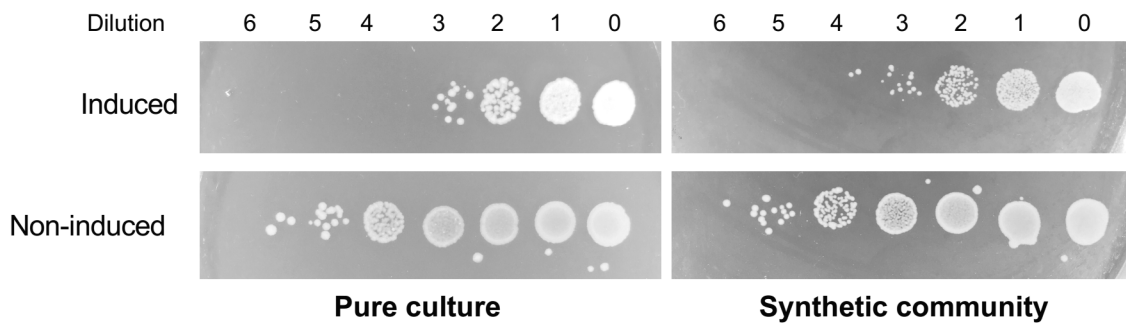


FIG. 5B

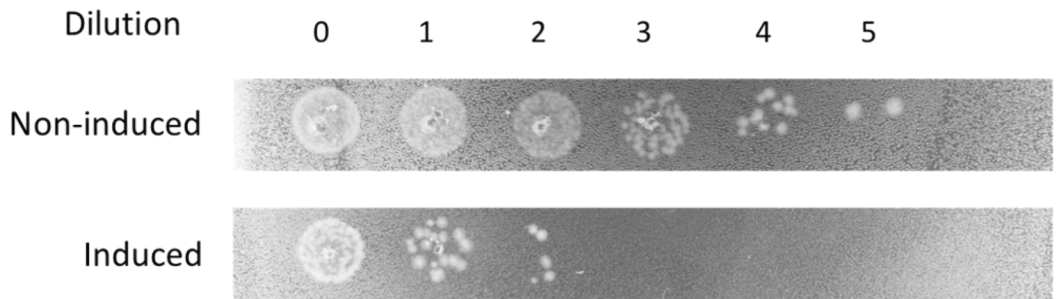


FIG. 6A

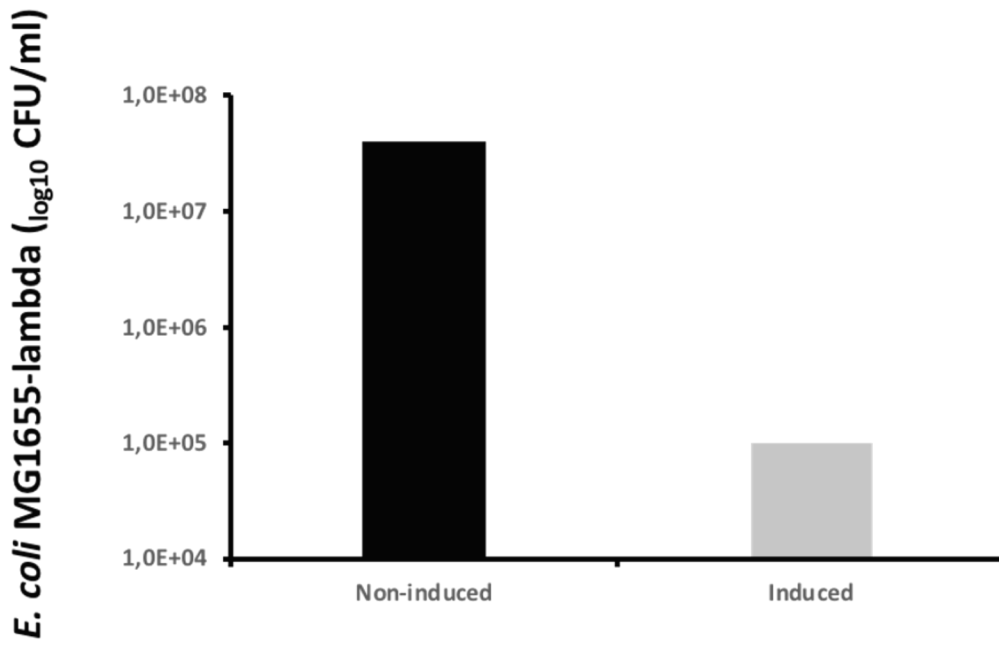


FIG. 6B

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SINGLE-VECTOR TYPE I VECTORS**CROSS-REFERENCE TO RELATED APPLICATIONS**

This application claims priority to Great Britain Patent Application No. 1816700.7, filed Oct. 14, 2018, and Great Britain Patent Application No. 1817509.1, filed Oct. 27, 2018, the contents of each of which are hereby incorporated herein by reference in their entirety.

SUBMISSION OF SEQUENCE LISTING ON ASCII TEXT FILE

The content of the following submission on ASCII text file is incorporated herein by reference in its entirety: a computer readable form (CRF) of the Sequence Listing (file name: 786212000600SEQLIST.TXT, date recorded: Nov. 26, 2018, size: 6,008 bytes).

TECHNICAL FIELD

The invention relates to the production and use of Cas-encoding sequences and vectors comprising these. Aspects of the invention provide products, vectors, delivery vehicles, uses and methods for producing Cas-encoding sequences in bacterial or archaeal cells.

BACKGROUND

The state of the art describes vectors and uses of these that employ CRISPR/Cas systems. For example, reference is made to WO2017/118598, US20180140698, US20170246221, US20180273940, US20160115488, US20180179547, US20170175142, US20160024510, US20150064138, US20170022499, US20160345578, US20180155729, US20180200342, WO2017112620, WO2018081502, PCT/EP2018/066954, PCT/EP2018/066980, PCT/EP2018/071454 and U.S. Ser. No. 15/985,658 and equivalent publications by the US Patent and Trademark Office (USPTO) or WIPO, the disclosures of which are incorporated herein by reference.

SUMMARY OF THE INVENTION

The invention provides the following configurations.

In a First Configuration

A nucleic acid vector for introduction into a host cell, the vector comprising a first nucleotide sequence encoding a Type I Cas3 and a second nucleotide sequence encoding one or more Cascade proteins, wherein the first and second sequences are under the control of one or more promoters comprised by the vector for expression of the proteins in the cell.

In an example, the vector comprises an operon for expression in the cell of the Cas3 and Cascade proteins from a Cas module, the module comprising the nucleotide sequences encoding the Cas3 and Cascade proteins, and the operon comprising the Cas module under the control of a promoter for controlling the expression of both the Cas3 and Cascade proteins.

The invention also provides a delivery vehicle comprising the vector, as well as a pharmaceutical composition comprising the vector or vehicle and a pharmaceutically acceptable diluent, excipient or carrier.

The invention also provides a method of treating or reducing the risk of a disease or condition in a human or

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animal subject, the method comprising administering the vector, vehicle or composition to the subject, and introducing the vector into target host bacterial or archaeal cells in the subject (eg, in a gut microbiota, lung, eye or blood of the subject), wherein the Cas cuts (or otherwise modifies) one or more target sequences in the target cells and the cells are killed or growth or proliferation of the cells is reduced.

In a Second Configuration

A method of amplifying copies of a DNA encoding a functional Cas protein (optionally a Cas nuclease) in a bacterial or archaeal production strain of cells, the method comprising

(a) Providing production strain cells, each cell comprising a copy of said DNA, wherein each DNA comprises a nucleotide sequence encoding said Cas, wherein the nucleotide sequence is under the control of a promoter for controlling the expression of the Cas in the production strain cell, the DNA comprising an origin of replication that is operable in the cell for replication of the DNA;

(b) Culturing the cells to allow replication of the DNA, whereby the DNA is amplified; and

(c) Optionally isolating copies of the DNA,

Optionally wherein the promoter is an attenuated constitutive promoter.

In a Third Configuration

Use of an attenuated promoter in a DNA construct comprising a nucleotide sequence encoding a functional Cas protein (optionally a Cas nuclease) that is under the control of the promoter, in a method of amplifying copies of the DNA in a population of bacterial or archaeal production strain cells, the method comprising culturing the cells to allow replication of the DNA thereby amplifying the DNA in the cells, for enhancing the yield of amplified DNA produced by the production host cells.

In a Fourth Configuration

Use of an attenuated promoter in a DNA construct comprising a nucleotide sequence encoding a functional Cas protein (optionally a Cas nuclease) that is under the control of the promoter, in a method of amplifying copies of the DNA in a population of bacterial or archaeal production strain cells, the method comprising culturing the cells to allow replication of the DNA thereby amplifying the DNA in the cells, for reducing toxicity of the Cas in the production strain.

In a Fifth Configuration

Use of an attenuated promoter in a DNA construct comprising a nucleotide sequence encoding a functional Cas protein (optionally a Cas nuclease) that is under the control of the promoter, in a method of amplifying copies of the DNA in a population of bacterial or archaeal production strain cells, the method comprising culturing the cells to allow replication of the DNA thereby amplifying the DNA in the cells, for reducing mutation of the DNA (optionally the Cas-encoding sequence) in the production strain.

In a Sixth Configuration

Use of an attenuated promoter in a DNA construct comprising a nucleotide sequence encoding a functional Cas protein (optionally a Cas nuclease) that is under the control of the promoter, in a method of amplifying copies of the DNA in a population of bacterial or archaeal production strain cells, the method comprising culturing the cells to allow replication of the DNA thereby amplifying the DNA in the cells, for promoting production cell viability during the amplification of the DNA.

In a Seventh Configuration

Use of an attenuated promoter in a DNA construct comprising a nucleotide sequence encoding a functional Cas protein (optionally a Cas nuclease) that is under the control of the promoter, in a method of amplifying copies of the DNA in a population of bacterial or archaeal production strain cells, the method comprising culturing the cells to allow replication of the DNA thereby amplifying the DNA in the cells, for reducing the occurrence of Cas cutting of DNA.

In an Eighth Configuration

A method for enhancing the yield of amplified copies of a DNA construct in a population of bacterial or archaeal production strain cells, wherein the construct comprises a nucleotide sequence encoding a functional Cas protein (optionally a Cas nuclease) that is under the control of a promoter, the method comprising culturing the cells to allow replication of the DNA thereby amplifying the DNA in the cells, wherein the promoter is an attenuated promoter.

In a Ninth Configuration

A method for reducing toxicity of a functional Cas protein (optionally a Cas nuclease) in a population of bacterial or archaeal production strain cells in a process of amplifying copies of a DNA construct, wherein the construct comprises a nucleotide sequence encoding the Cas and the sequence is under the control of a promoter, the method comprising culturing the cells to allow replication of the DNA thereby amplifying the DNA in the cells, wherein the promoter is an attenuated promoter.

In a Tenth Configuration

A method for reducing mutation of a DNA construct encoding a functional Cas protein (optionally a Cas nuclease) in a population of bacterial or archaeal production strain cells in a process of amplifying copies of the construct, wherein the construct comprises a nucleotide sequence encoding the Cas and the sequence is under the control of a promoter, the method comprising culturing the cells to allow replication of the DNA thereby amplifying the DNA in the cells, wherein the promoter is an attenuated promoter.

In an Eleventh Configuration

A method for promoting production cell viability of a population of bacterial or archaeal production strain cells in a process of amplifying copies of a DNA construct comprised by the cells, wherein the construct comprises a nucleotide sequence encoding a functional Cas protein (optionally a Cas nuclease) and the sequence is under the control of a promoter, the method comprising culturing the cells to allow replication of the DNA thereby amplifying the DNA in the cells, wherein the promoter is an attenuated promoter.

In a Twelfth Configuration

A method for reducing the occurrence of Cas nuclease cutting of a DNA construct in a population of bacterial or archaeal production strain cells in a process of amplifying copies of the construct, wherein the construct comprises a nucleotide sequence encoding the Cas and the sequence is under the control of a promoter, the method comprising culturing the cells to allow replication of the DNA thereby amplifying the DNA in the cells, wherein the promoter is an attenuated promoter.

BRIEF DESCRIPTION OF THE DRAWINGS

FIGS. 1A-1C. Type I CRISPR-Cas system of *C. difficile* targeting *E. coli* MG1655. (FIG. 1A) Layout of the CRISPR Guided Vector™, CGV™. Plasmid 1: pSC101 ori, pBAD promoter (induced by arabinose), cas3 and cascade genes.

Plasmid 2: pCloDF13 ori, pTac promoter (induced by IPTG), CRISPR array. (FIG. 1B) Dilution series (10^1 - 10^6) of drop spots (5 μ l) of *E. coli* MG1655 harboring the CGV on LB agar plates with and without inducers. (FIG. 1C) CRISPR induction killed 99.9% of the population (grey bar). Growth in absence of induction is shown in black. CGV™ refers to a CRISPR Guided Vector™, which is a nucleic acid vector comprising nucleotide sequences encoding CRISPR/Cas components.

FIGS. 2A-2C. Type I CRISPR-Cas system of *C. difficile* targeting *E. coli* MG1655. (FIG. 2A) Layout of the CRISPR Guided Vector™, CGV™. pSC101 ori, pTac promoter (induced by IPTG), CRISPR array, pBAD promoter (induced by arabinose), cas3 and cascade genes. (FIG. 2B) Dilution series (10^1 - 10^6) of drop spots (5 μ l) of *E. coli* MG1655 harboring the CGV on SM agar plates with and without inducers. (FIG. 2C) CRISPR induction killed 99% of the population (grey bar). Growth in absence of induction is shown in black. CGV™ refers to a CRISPR Guided Vector™, which is a nucleic acid vector comprising nucleotide sequences encoding CRISPR/Cas components.

FIGS. 3A-3B. Time-kill curves for *E. coli* MG1655 harboring the CGV. (FIG. 3A) CRISPR induction killed 99% of the population in 60 minutes (dashed line). Growth in absence of induction is shown in black lines. CRISPR/Cas was induced at time-point 0 and monitored until 120 minutes. (FIG. 3B) Dilution series (10^1 - 10^6) of drop spots (5 μ l) on SM agar plates of *E. coli* MG1655 after 60 minutes of induction.

FIGS. 4A-4B. Specific killing of *E. coli* MG1655 with type I-B CRISPR-Cas system of *C. difficile* in a synthetic microbial consortium. (FIG. 4A) Bacteria count of a synthetic population composed of three different strains. CRISPR was induced at time-point 0 and monitored for 60 minutes. Growth in absence of induction is shown in black. CRISPR induction prompted 1- \log_{10} reduction in viable cells of target strain *E. coli* MG1655, while leaving *E. coli* Top10 and *L. lactis* NZ9000 populations intact (dark grey bars). (FIG. 4B) Dilution series (10^1 - 10^6) of drop spots (5 μ l) of the bacterial community mixture after 60 minutes of induction. *E. coli* MG1655 grows selectively on BHI+ streptomycin, *E. coli* Top10 on ampicillin, and *L. lactis* NZ9000 on chloramphenicol.

FIGS. 5A-5B. Killing of *E. coli* MG1655 with type I-B CRISPR-Cas system of *C. difficile* in a synthetic microbial consortium compared to a pure culture of *E. coli* MG1655. (FIG. 5A) CRISPR induction generated 4- \log_{10} reductions in viable cells of target strain *E. coli* MG1655, both in the pure culture and in the community mixture (grey bars). Growth in absence of induction is shown in black. (FIG. 5B) Dilution series of a pure culture of *E. coli* MG1655 and the bacterial community mixture on streptomycin plates with and without inducers.

FIGS. 6A-6B. Type I CRISPR-Cas system of *E. coli* targeting *E. coli* MG1655. (FIG. 6A) Dilution series (10^1 - 10^6) of drop spots (5 μ l) of *E. coli* MG1655 harboring the CGV on SM agar plates with and without inducers. (FIG. 6B) CRISPR induction killed 99% of the population (grey bar). Growth in absence of induction is shown in black. CGV™ refers to a CRISPR Guided Vector™, which is a nucleic acid vector comprising nucleotide sequences encoding CRISPR/Cas components.

DETAILED DESCRIPTION

The invention relates to the production and use of Cas-encoding sequences and vectors comprising these. Aspects

of the invention provide products, vectors, delivery vehicles, uses and methods for producing Cas-encoding sequences in bacterial or archaeal cells.

An aspect of the invention provides for the control of expression of Cas and optionally also Cascade proteins from single vectors, such as by regulated use of Cas modules in an operon and/or using attenuated promoters.

Concepts:

An aspect of the invention provides nucleic acid vectors that are useful for introducing into target host cells of any eukaryotic or prokaryotic species (eg, ex vivo or in vitro) for expressing Type I Cas and optionally other components of a Type I CRISPR/Cas system. Usefully, the vector may in some examples therefore provide a single-vector means for introducing a complete exogenous Type I CRISPR/Cas system into a target cell for modification (eg, cutting by Cas3) of DNA in the target cell. In an example, a chromosomal target sequence (ie, protospacer that is cognate with the Cas3) is modified. In another example, an episomal DNA sequence is modified, for example a plasmid sequence or a DNA that has been introduced into the cell. The latter may be useful in a recombinering method of the invention wherein exogenous DNA in the target cell is cut by the Cas3 and optionally this produces one or more recombinogenic ends for recombination of the cut DNA with a further DNA of interest, thereby producing a recombination product in the cell. For example, in such a recombinering method, the target cell is a recombinogenic *E. coli* cell, eg, comprising a red/ET system. In another example, the target cell is an undesired cell (eg, a cell of a species or strain that is pathogenic to humans or animals, such as a bacterial disease-causing species or strain) and the cutting by Cas3 kills the cell. This may be useful for treating or preventing an infection in a human or animal harbouring target cells. The provision of single-vector means that express minimally a Cas endonuclease (eg, Cas3), cognate accessory proteins (eg, Cascade proteins) and at least one CRISPR array (or nucleotide sequence encoding a guide RNA (eg, a single guide RNA)), wherein the Cas, accessory proteins and array (or nucleotide sequence) comprise a functional CRISPR/Cas system is more convenient and the inventors believe more efficient for introducing into a target cell and effecting CRISPR/Cas modification of a target sequence therein than the use of 2 or 3 or more separate vectors (eg, a vector encoding the Cas nuclease and a different vector encoding the accessory proteins, and possibly a further vector comprising the array (or gRNA-encoding nucleotide sequence) which all need to transform the target cell for the system to function). This may provide one or more benefits, therefore, such as simplifying delivery (and thus the design of delivery vehicles), simplifying construction of the vector and vehicle and/or providing for better cutting or killing efficiencies. Conveniently, an example of the invention therefore uses an operon for the coordinated expression in the target cells of the Cas and accessory proteins (and optionally also the array or gRNA-encoding sequence(s)). Whilst not wishing to be bound by any particular theory, the introduction of a single vector (eg, using an operon) as per the invention may advantageously coordinate the expression of the Cas and accessory proteins (and optionally production of cRNAs or gRNAs) so that these are available to operate together without undue delay in the target cell. This may be important to tip the balance between, on the one hand the target cell using its endogenous anti-restriction, endogenous Cas or other endogenous mechanisms that seek out and degrade invading phage and DNA, and on the other hand efficient cell killing or deactivation of such mechanisms by the

invading CRISPR components of the vector of the invention. In such an arms race, concerted and early operation of the CRISPR components in the cell are likely to be important to gain the upper hand and effect cell killing. The invention provides means to assist this.

By way of example, the invention thus provides the following Concepts:

1. A nucleic acid vector for introduction into a host cell, the vector comprising a first nucleotide sequence encoding a Type I Cas3 and a second nucleotide sequence encoding one or more Cascade proteins, wherein the first and second sequences are under the control of one or more promoters comprised by the vector for expression of the proteins in the cell.
2. The vector of concept 1, wherein the vector comprises an operon for expression in the cell of the Cas3 and Cascade proteins from a Cas module, the module comprising the nucleotide sequences encoding the Cas3 and Cascade proteins, and the operon comprising the Cas module under the control of a promoter for controlling the expression of both the Cas3 and Cascade proteins.
3. The vector of concept 2, wherein
 - (a) the first sequence is between the promoter and the second sequence in the operon;
 - (b) the operon comprises no Cas-encoding nucleotide sequences between the promoter and the first nucleotide sequence; and/or
 - (c) the operon comprises (in 5' to 3' direction) the promoter, the first sequence and the second sequence.
4. The vector of any preceding concept, wherein each promoter is a constitutive promoter.
5. The vector of any one of concepts 1 to 3, wherein the promoter is repressible (optionally repressible by a tetracycline repressor or lac repressor).
6. The vector of any one of concepts 1 to 3, wherein the promoter is inducible.
7. The vector of any preceding concept, wherein the first sequence is under the control of a medium strength promoter.
8. The vector of any preceding concept, wherein the first sequence is under the control of a promoter that has an Anderson Score (AS) of $0.5 > AS > 0.1$.
9. The vector of any preceding concept, wherein the first sequence (and optionally the second sequence) is under the control of a promoter and translation initiation site (TIS) combination that is capable of producing expression of green fluorescent protein (GFP) from a first expression operating unit (EOU) in *E. coli* strain BW25113 cells with a fluorescence of from 0.5 to 4 times the fluorescence produced in *E. coli* strain BW25113 cells using a second EOU comprising a P10 promoter (SEQ ID NO: 1) combined with a BCD14 TIS (SEQ ID NO: 2), wherein the EOUs differ only in their promoter and TIS combinations, wherein each EOU comprises (in 5' to 3' direction) an upstream initiator, the respective promoter, the respective TIS, a nucleotide sequence encoding GFP, a 3' UTR, a transcription terminator and a downstream insulator.
10. The vector of concept 9, wherein fluorescence using the first EOU is 0.5 to 2 times the fluorescence using the second EOU.
11. The vector of any preceding concept, wherein the vector comprises an origin of replication that is operable in the host cell.

12. The vector of any preceding concept, wherein the vector comprises an origin of replication that is operable in a bacterial cell of a vector production strain, wherein the Cas3 is not operable in the production strain cell to target and cut a chromosomal sequence thereof. 5
13. The vector of concept 12, wherein the first sequence is under the control of a promoter that is capable of controlling expression of the Cas3 at a level that is not toxic to the production strain cell. 10
14. The vector of any preceding concept, wherein the vector is a high copy number vector.
15. The vector of any preceding concept, wherein the first nucleotide sequence or operon is comprised by a mobile genetic element. 15
16. The vector of any preceding concept, wherein the vector is devoid of a Cas adaption module.
17. The vector of any preceding concept, wherein the vector is devoid of nucleotide sequence encoding one, more or all of a Cas1, Cas2, Cas4, Cas6, Cas7 and Cas8. 20
18. The vector of any preceding concept, wherein the vector comprises (optionally in 5' to 3' direction) nucleotide sequence encoding one, more or all of Cas11, Cas7 and Cas8a1. 25
19. The vector of concept 18, wherein the vector comprises nucleotide sequence encoding Cas3' and/or Cas3".
20. The vector or concept 19, wherein the nucleotide sequences encoding the Cas3' and/or Cas3" are between the promoter and the sequence(s) recited in concept 18. 30
21. The vector of any one of concepts 18 to 20, wherein the host cell comprises a Type IA CRISPR array that is cognate with the Cas3. 35
22. The vector of any one of concepts 18 to 20, wherein the host cell comprises an endogenous Type IB, C, U, D, E or F CRISPR/Cas system.
23. The vector of any one of concepts 1 to 17, wherein the vector comprises (optionally in 5' to 3' direction) nucleotide sequence encoding one, more or all of Cas8b1, Cas7 and Cas5. 40
24. The vector of concept 23, wherein the vector comprises a nucleotide sequence encoding Cas3 between the promoter and the sequence(s) recited in concept 23. 45
25. The vector of concept 23 or 24, wherein the host cell comprises a Type IB CRISPR array that is cognate with the Cas3.
26. The vector of concept 23 or 24, wherein the host cell comprises an endogenous Type IA, C, U, D, E or F CRISPR/Cas system. 50
27. The vector of any one of concepts 1 to 17, wherein the vector comprises (optionally in 5' to 3' direction) nucleotide sequence encoding one, more or all of Cas5, Cas8c and Cas7. 55
28. The vector of concept 27, wherein the vector comprises a nucleotide sequence encoding Cas3 between the promoter and the sequence(s) recited in concept 27.
29. The vector of concept 27 or 28, wherein the host cell comprises a Type IC CRISPR array that is cognate with the Cas3. 60
30. The vector of concept 27 or 28, wherein the host cell comprises an endogenous Type IA, B, U, D, E or F CRISPR/Cas system. 65
31. The vector of any one of concepts 1 to 17, wherein the vector comprises (optionally in 5' to 3' direction)

- nucleotide sequence encoding one, more or all of Cas8U2, Cas7, Cas5 and Cas6.
32. The vector of concept 31, wherein the vector comprises a nucleotide sequence encoding Cas3 between the promoter and the sequence(s) recited in concept 31.
33. The vector of concept 31 or 32, wherein the host cell comprises a Type IU CRISPR array that is cognate with the Cas3.
34. The vector of concept 31 or 32, wherein the host cell comprises an endogenous Type IA, B, C, D, E or F CRISPR/Cas system.
35. The vector of any one of concepts 1 to 17, wherein the vector comprises (optionally in 5' to 3' direction) nucleotide sequence encoding one, more or all of Cas10d, Cas7 and Cas5.
36. The vector of concept 35, wherein the vector comprises a nucleotide sequence encoding Cas3' and/or Cas3".
37. The vector of concept 36, wherein the nucleotide sequences encoding the Cas3' and/or Cas3" are between the promoter and the sequence(s) recited in concept 35.
38. The vector of any one of concepts 35 to 37, wherein the host cell comprises a Type ID CRISPR array that is cognate with the Cas3.
39. The vector of any one of concepts 35 to 37, wherein the host cell comprises an endogenous Type IA, B, C, U, E or F CRISPR/Cas system.
40. The vector of any one of concepts 1 to 17, wherein the vector comprises (optionally in 5' to 3' direction) nucleotide sequence encoding one, more or all of Cas8e, Cas11, Cas7, Cas5 and Cas6.
41. The vector of concept 40, wherein the vector comprises a nucleotide sequence encoding Cas3 between the promoter and the sequence(s) recited in concept 40.
42. The vector of concept 40 or 41, wherein the host cell comprises a Type IE CRISPR array that is cognate with the Cas3.
43. The vector of concept 40 or 41, wherein the host cell comprises an endogenous Type IA, B, C, D, U or F CRISPR/Cas system.
44. The vector of any one of concepts 1 to 17, wherein the vector comprises (optionally in 5' to 3' direction) nucleotide sequence encoding one, more or all of Cas8f, Cas5, Cas7 and Cas6f.
45. The vector of concept 44, wherein the vector comprises a nucleotide sequence encoding Cas3 between the promoter and the sequence(s) recited in concept 44, wherein the vector is devoid of nucleotide sequence encoding further Cas between the promoter and the sequence encoding the Cas3.
46. The vector of concept 44 or 45, wherein the host cell comprises a Type IF CRISPR array that is cognate with the Cas3.
47. The vector of concept 44 or 45, wherein the host cell comprises an endogenous Type IA, B, C, D, U or E CRISPR/Cas system.
48. The vector of any one of concepts 1 to 17, wherein the Cas and Cascade are
 - (a) Type IA Cas and Cascade proteins;
 - (b) Type IB Cas and Cascade proteins;
 - (c) Type IC Cas and Cascade proteins;
 - (d) Type ID Cas and Cascade proteins;
 - (e) Type IE Cas and Cascade proteins;
 - (f) Type IF Cas and Cascade proteins; or
 - (g) Type IU Cas and Cascade proteins.

49. The vector of any preceding concept, wherein the Cas and Cascade are *E coli* (optionally Type IE or IF) Cas and Cascade proteins.
50. The vector of concept 49, wherein the *E coli* is ESBL-producing *E. coli* or *E. coli* ST131-O25b:H4.
51. The vector of any preceding concept, wherein the Cas and Cascade are
- (a) *Clostridium* (eg, *C difficile*) Cas and Cascade proteins, optionally *C difficile* resistant to one or more antibiotics selected from aminoglycosides, lincomycin, tetracyclines, erythromycin, clindamycin, penicillins, cephalosporins and fluoroquinolones;
 - (b) *Pseudomonas aeruginosa* Cas and Cascade proteins, optionally *P aeruginosa* resistant to one or more antibiotics selected from carbapenems, aminoglycosides, cefepime, ceftazidime, fluoroquinolones, piperacillin and tazobactam; or
 - (c) *Klebsiella pneumoniae* (eg, carbapenem-resistant *Klebsiella pneumoniae* or Extended-Spectrum Beta-Lactamase (ESBL)-producing *K pneumoniae*) Cas and Cascade proteins.
52. The vector of any preceding concept, wherein the Cas and Cascade are *E coli*, *C difficile*, *P aeruginosa*, *K pneumoniae*, *P furiosus* or *B halodurans* Cas and Cascade proteins.
53. The vector of any preceding concept, wherein the Cas3 is a Cas3 of a CRISPR/Cas locus of a first bacterial or archaeal species, wherein the distance between the Cas3-encoding sequence of the locus and its cognate promoter is further than the distance between the Cas3-encoding sequence and the respective promoter comprised by the vector.
54. The vector of any preceding concept, wherein the distance between the promoter and the Cas3-encoding sequence and/or Cascade protein-encoding sequence(s) is shorter than in a corresponding wild-type Type I locus.
55. The vector of any preceding concept, wherein the vector comprises (i) a CRISPR array for producing crRNAs in the host cell and/or (ii) one or more nucleotide sequences encoding one or more guide RNAs (gRNAs or single gRNAs), wherein the crRNAs or gRNAs are cognate to the Cas3 (and optionally cognate to the Cascade proteins).
56. The vector of concept 55 when dependent from concept 2, wherein the array or gRNA-encoding sequence(s) are comprised by the operon and under the control of the promoter.
57. The vector of concept 56, wherein the array or gRNA-encoding sequence(s) are under the control of a promoter that is different from the promoter that controls the expression of the Cas3.
58. The vector of concept 56 or 57, wherein one or more of the crRNAs or gRNAs comprises a spacer sequence that is capable of hybridising to a target nucleotide sequence of the host cell, wherein the target sequence is adjacent a PAM, the PAM being cognate to the Cas3.
59. The vector of concept 58, wherein the target sequence is a chromosomal sequence of the host cell.
60. The vector of concept 58 or 59, wherein the Cas3 is operable to cut the target sequence.
61. The vector of any preceding concept, wherein the vector is a plasmid or phagemid.
62. A delivery vehicle comprising the vector of any preceding concept, wherein the delivery vehicle is capable of delivering the vector into the host cell.

63. The vehicle of concept 62, wherein the delivery vehicle is a phage, non-replicative transduction particle, nanoparticle carrier, bacterium or liposome.
64. The vector or vehicle of any preceding concept, wherein the host cell is a bacterial or archaeal cell, optionally, the host cell is a *C difficile*, *P aeruginosa*, *K pneumoniae* (eg, carbapenem-resistant *Klebsiella pneumoniae* or Extended-Spectrum Beta-Lactamase (ESBL)-producing *K pneumoniae*), *E coli* (eg, ESBL-producing *E. coli*, or *E. coli* ST131-O25b:H4), *H pylori*, *S pneumoniae* or *S aureus* cell.
65. The vector or vehicle of any preceding concept for administration to a human or animal subject for treating or reducing the risk of a disease or condition in the subject.
66. The vector or vehicle of concept 65, wherein the disease or condition is an infection of the subject with host cells (eg, bacterial cells), or wherein the disease or condition is mediated by host cells (eg, bacterial cells).
67. A pharmaceutical composition comprising the vector or vehicle of any preceding concept and a pharmaceutically acceptable diluent, excipient or carrier.
68. A method of treating or reducing the risk of a disease or condition in a human or animal subject, the method comprising administering the vector, vehicle or composition of any preceding concept to the subject, and introducing the vector into target host bacterial or archaeal cells in the subject (eg, in a gut microbiota, lung, eye or blood of the subject), wherein the Cas cuts (or otherwise modifies) one or more target sequences in the target cells and the cells are killed or growth or proliferation of the cells is reduced.
69. The method of concept 68, wherein the target cells are cells of a disease pathogen species.
70. The method of concept 68 or 69, wherein the target cells are *C difficile*, *P aeruginosa*, *K pneumoniae* (eg, carbapenem-resistant *Klebsiella pneumoniae* or Extended-Spectrum Beta-Lactamase (ESBL)-producing *K pneumoniae*), *E coli* (eg, ESBL-producing *E. coli*, or *E. coli* ST131-O25b:H4), *H pylori*, *S pneumoniae* or *S aureus* cells.

EMBODIMENTS

An aspect of the invention provides improved ways of amplifying DNA constructs in bacterial and archaeal production strain cells. For example, the DNA may be a high copy number plasmid or phagemid comprising a constitutive promoter for controlling the expression of one or more Cas proteins when the DNA has been introduced into a target host bacterial or host cell. It is desirable, according to an aspect of the invention, to consider attenuating the promoter activity during amplification of the DNA in the production strain. This is useful, since the inventors have found that Cas expression in production strains may be toxic to production strain cells, thereby reducing the yield of amplified DNA. Toxicity may be due, for example, to off-target cutting of the production strain chromosomal DNA when the Cas is a nuclease (such as Cas9 or Cas3) and/or due to relatively high levels of expression of the Cas in the cells. Additionally or alternatively, undesirably the Cas expression or activity may impose a selective pressure that favours mutation and propagation of mutated DNA constructs (such as mutation in one more or all of a CRISPR/Cas operon, Cas-encoding gene, Cascade-encoding gene, CRISPR array and gRNA-encoding sequence of the DNA construct) in production cells, thereby reducing the yield of desired amplified constructs and

imposing an undesired step of separating desired from mutated DNA constructs for further formulation into useful compositions. Such compositions may be pharmaceutical compositions, herbicides, pesticides, environmental remediation compositions etc. In one example, the promoter attenuation in production strains is achieved by using a medium strength (not high or low) promoter to control the Cas-encoding nucleotide sequence of the DNA constructs. A medium level of Cas expression may be tolerable in the production strains, and yet once the DNA is subsequently introduced into target host cells the Cas is expressed at sufficiently high levels to produce desired activity to modify (eg, cut) target sequences in target cells. In an alternative, the invention uses a repressible promoter, wherein the promoter is repressed in production strain, but not repressed in target host cells. For example, aspects of the invention use a tetracycline repressor (tetR) expressed in production strain cells that represses the promoter.

Thus, the yield can be enhanced by one or more of

- (a) reducing toxicity of the Cas in the production strain;
- (b) reducing mutation of the DNA (optionally the Cas-encoding sequence) in the production strain;
- (c) promoting production cell viability during the amplification of the DNA; and
- (d) reducing the occurrence of Cas cutting of DNA (optionally cutting of production host cell chromosomal DNA or said DNA construct).

To this end, the invention provides Embodiments as follows:

1. A method of amplifying copies of a DNA encoding a functional Cas protein (optionally a Cas nuclease) in a bacterial or archaeal production strain of cells, the method comprising
 - (a) Providing production strain cells, each cell comprising a copy of said DNA, wherein each DNA comprises a nucleotide sequence encoding said Cas, wherein the nucleotide sequence is under the control of a promoter for controlling the expression of the Cas in the production strain cell, the DNA comprising an origin of replication that is operable in the cell for replication of the DNA;
 - (b) Culturing the cells to allow replication of the DNA, whereby the DNA is amplified; and
 - (c) Optionally isolating copies of the DNA, wherein the promoter is an attenuated constitutive promoter.

In an example, promoter is a medium strength promoter. In another example, the promoter is repressed in the production strain cell. Hence, the promoter is an attenuated promoter in these examples.
2. Use of an attenuated promoter in a DNA construct comprising a nucleotide sequence encoding a functional Cas protein (optionally a Cas nuclease) that is under the control of the promoter, in a method of amplifying copies of the DNA in a population of bacterial or archaeal production strain cells, the method comprising culturing the cells to allow replication of the DNA thereby amplifying the DNA in the cells, for enhancing the yield of amplified DNA produced by the production host cells.
3. The use of paragraph 2, wherein the use is for enhancing said yield by
 - (a) reducing toxicity of the Cas in the production strain;
 - (b) reducing mutation of the DNA (optionally the Cas-encoding sequence) in the production strain;
 - (c) promoting production cell viability during the amplification of the DNA; and/or

(d) reducing the occurrence of Cas cutting of DNA (optionally cutting of production host cell chromosomal DNA or said DNA construct).

4. Use of an attenuated promoter in a DNA construct comprising a nucleotide sequence encoding a functional Cas protein (optionally a Cas nuclease) that is under the control of the promoter, in a method of amplifying copies of the DNA in a population of bacterial or archaeal production strain cells, the method comprising culturing the cells to allow replication of the DNA thereby amplifying the DNA in the cells, for reducing toxicity of the Cas in the production strain.
5. Use of an attenuated promoter in a DNA construct comprising a nucleotide sequence encoding a functional Cas protein (optionally a Cas nuclease) that is under the control of the promoter, in a method of amplifying copies of the DNA in a population of bacterial or archaeal production strain cells, the method comprising culturing the cells to allow replication of the DNA thereby amplifying the DNA in the cells, for reducing mutation of the DNA (optionally the Cas-encoding sequence) in the production strain.
6. Use of an attenuated promoter in a DNA construct comprising a nucleotide sequence encoding a functional Cas protein (optionally a Cas nuclease) that is under the control of the promoter, in a method of amplifying copies of the DNA in a population of bacterial or archaeal production strain cells, the method comprising culturing the cells to allow replication of the DNA thereby amplifying the DNA in the cells, for promoting production cell viability during the amplification of the DNA.
7. Use of an attenuated promoter in a DNA construct comprising a nucleotide sequence encoding a functional Cas protein (optionally a Cas nuclease) that is under the control of the promoter, in a method of amplifying copies of the DNA in a population of bacterial or archaeal production strain cells, the method comprising culturing the cells to allow replication of the DNA thereby amplifying the DNA in the cells, for reducing the occurrence of Cas cutting of DNA.
8. A method for enhancing the yield of amplified copies of a DNA construct in a population of bacterial or archaeal production strain cells, wherein the construct comprises a nucleotide sequence encoding a functional Cas protein (optionally a Cas nuclease) that is under the control of a promoter, the method comprising culturing the cells to allow replication of the DNA thereby amplifying the DNA in the cells, wherein the promoter is an attenuated promoter.
9. A method for reducing toxicity of a functional Cas protein (optionally a Cas nuclease) in a population of bacterial or archaeal production strain cells in a process of amplifying copies of a DNA construct, wherein the construct comprises a nucleotide sequence encoding the Cas and the sequence is under the control of a promoter, the method comprising culturing the cells to allow replication of the DNA thereby amplifying the DNA in the cells, wherein the promoter is an attenuated promoter.
10. A method for reducing mutation of a DNA construct encoding a functional Cas protein (optionally a Cas nuclease) in a population of bacterial or archaeal production strain cells in a process of amplifying copies of the construct, wherein the construct comprises a nucleotide sequence encoding the Cas and the sequence is under the control of a promoter, the method com-

prising culturing the cells to allow replication of the DNA thereby amplifying the DNA in the cells, wherein the promoter is an attenuated promoter.

11. A method for promoting production cell viability of a population of bacterial or archaeal production strain cells in a process of amplifying copies of a DNA construct comprised by the cells, wherein the construct comprises a nucleotide sequence encoding a functional Cas protein (optionally a Cas nuclease) and the sequence is under the control of a promoter, the method comprising culturing the cells to allow replication of the DNA thereby amplifying the DNA in the cells, wherein the promoter is an attenuated promoter.
12. A method for reducing the occurrence of Cas nuclease cutting of a DNA construct in a population of bacterial or archaeal production strain cells in a process of amplifying copies of the construct, wherein the construct comprises a nucleotide sequence encoding the Cas and the sequence is under the control of a promoter, the method comprising culturing the cells to allow replication of the DNA thereby amplifying the DNA in the cells, wherein the promoter is an attenuated promoter.
13. The use of paragraph 5 or 7, or the method of paragraph 10 or 12, wherein the mutation or cutting is mutation or cutting of host cell chromosomal DNA or the construct DNA.
14. The method or use of any one of paragraphs 2 to 13, wherein the promoter is a constitutive promoter.
15. The method or use of any preceding paragraph, wherein the promoter is repressed in the production strain cells (optionally repressed by a tetracycline repressor or a lac repressor).
16. The method or use of paragraph 15, wherein the promoter is P_{tetO-1} , $P_{LlacO-1}$ or a repressible homologue thereof.

Other examples of suitable repressible promoters are Ptac (repressed by lacI) and the Leftward promoter (pL) of phage lambda (which repressed by the λ cl repressor). In an example, the promoter comprises a repressible operator (eg, tetO or lacO) fused to a promoter sequence. The corresponding repressor is encoded by a nucleic acid in the production strain (eg, a chromosomally-integrated sequence or a sequence comprised by an episome) and the repressor is expressed during the DNA or vector amplification method of the invention, whereby the promoter controlling Cas expression is repressed. In delivery vehicles that are subsequently produced from isolated amplified DNA/vector, the vehicle is devoid of an expressible nucleotide sequence encoding the repressor, whereby the promoter is functional when the DNA/vector is introduced into a target host cell. For example, in the absence of the repressor the promoter is constitutively ON for expression of the Cas. The system is therefore primed to work once the DNA/vector is introduced into the host cells, and this effect can be enhanced further by using a high copy number DNA/vector comprising an origin of replication that is operable in the host cell. A high copy number vector or DNA is also desirable in the production strain cells for enhancing yield of the DNA/vector, and by use of an attenuated promoter as described herein (eg, medium strength promoter and/or repressed promoter in the production strain cells) one can minimise Cas toxicity whilst culturing to maximise amplification and thus yield of the DNA/vector.

17. The method or use of any preceding paragraph, wherein the promoter is a medium strength promoter.
18. The method or use of any preceding paragraph, wherein the promoter has an Anderson Score (AS) of $0.5 > AS > 0.1$.
19. The method or use of any preceding paragraph, wherein the nucleotide sequence encoding said Cas is under the control of a promoter and translation initiation site (TIS) combination that is capable of producing expression of green fluorescent protein (GFP) from a first expression operating unit (EOU) in *E. coli* strain BW25113 cells with a fluorescence of from 0.5 to 4 times the fluorescence produced in *E. coli* strain BW25113 cells using a second EOU comprising a P10 promoter (SEQ ID NO: 1) combined with a BCD14 TIS (SEQ ID NO: 2), wherein the EOUs differ only in their promoter and TIS combinations, wherein each EOU comprises (in 5' to 3' direction) an upstream initiator, the respective promoter, the respective TIS, a nucleotide sequence encoding GFP, a 3' UTR, a transcription terminator and a downstream insulator.
20. The method or use of paragraph 19, wherein fluorescence using the first EOU is 0.5 to 2 times the fluorescence using the second EOU.
21. The method or use of any preceding paragraph, wherein the nuclease is Cas3 and optionally the DNA or cell encodes cognate Cascade proteins.
22. The method or use of any one of paragraphs 1 to 20, wherein the Cas is a Cas9.
23. The method or use of any preceding paragraph, wherein the production strain cells comprise a helper phage genome that is inducible to produce phage coat proteins in the cells, wherein the method further comprises inducing production of the phage proteins and causing packaging of the amplified DNA into phage particles or non-self-replicative transduction particles, and further isolating the phage or transduction particles and optionally formulating the particles into a pharmaceutical composition for administration to a human or animal subject for treating or reducing the risk of a disease or condition in the subject.
24. The method or use of paragraph 23, wherein the particles are capable of infecting target host cells in the subject and transducing the cells with the DNA, wherein the Cas and crRNAs (or guide RNAs, gRNAs) encoded by the DNA are expressed in the cells, the crRNAs or (gRNAs) being operable to guide the Cas to a target nucleotide sequence (optionally a chromosomal sequence) comprised by the cells, wherein the Cas cuts the target sequences in the cells, thereby killing host cells and treating or reducing the risk of the disease or condition.
25. The method or use of paragraph 24, wherein the host cells are bacterial or archaeal cells, optionally, the host cells are *C. difficile*, *P. aeruginosa*, *K. pneumoniae* (eg, carbapenem-resistant *Klebsiella pneumoniae* or Extended-Spectrum Beta-Lactamase (ESBL)-producing *K. pneumoniae*), *E. coli* (eg, ESBL-producing *E. coli*, or *E. coli* ST131-O25b:H4), *H. pylori*, *S. pneumoniae* or *S. aureus* cells.
26. The method or use of any preceding paragraph, wherein each DNA is comprised by a high copy number plasmid or phagemid.
27. The method or use of any preceding paragraph, wherein the DNA construct comprises one or more

nucleotide sequences for producing crRNAs or gRNAs that are operable for Cas nuclease targeting in target host cells.

Paragraphs & Generally Applicable Features

The invention provides the following Paragraphs, which are supported by the Examples below. Any features of the Concepts are combinable with any features of the Embodiments. Any features of the Concepts are combinable with any features of the Embodiments. Any features of the Paragraphs are combinable with any features of the Embodiments.

Any cell herein (eg, a production strain cell or target host cell) may be a bacterial cell, archaeal cell, algal cell, fungal cell, protozoan cell, invertebrate cell, vertebrate cell, fish cell, bird cell, mammal cell, companion animal cell, dog cell, cat cell, horse cell, mouse cell, rat cell, rabbit cell, eukaryotic cell, prokaryotic cell, human cell, animal cell, rodent cell, insect cell or plant cell. Preferably, the cell is a bacterial cell. Alternatively, the cell is a human cell. Optionally, the production strain cell(s) and target host cell(s) are of the same phylum, order, family, genus, species or strain.

1. A nucleic acid vector for introduction into a host cell, the vector comprising a first nucleotide sequence encoding a Type I Cas3, wherein the sequence is under the control of a promoter comprised by the vector for expression of the Cas3 in the cell.

In an example, the vector is a DNA vector, eg, ssDNA vector or dsDNA vector.

2. The vector of paragraph 1, wherein the vector comprises a second nucleotide sequence encoding one or more Cascade proteins, wherein the first and second sequences are under the control of one or more promoters comprised by the vector for expression of the proteins in the cell.

3. The vector of paragraph 2, wherein the Cascade protein(s) are cognate with the Cas3.

In an example, the Cas3 is cognate with Cascade proteins encoded by the host cell and/or encoded by a second operon. Optionally, the second operon is comprised by the vector. Optionally, the second operon is comprised by a second vector that is capable of introducing the second operon into the host cell, whereby the Cas3 and Cascade proteins are expressed from the operons in the host cell and are operable with crRNA or gRNA to target the Cas to a host cell target sequence, wherein the Cas3 is capable of modifying the target sequence.

4. The vector of paragraph 2 or 3, wherein the vector comprises an operon for expression in the cell of the Cas3 and Cascade proteins from a Cas module, the module comprising the nucleotide sequences encoding the Cas3 and Cascade proteins, and the operon comprising the Cas module under the control of a promoter for controlling the expression of both the Cas3 and Cascade proteins.

The term "operon" is known to the skilled person such as relating to a functioning unit of DNA containing at least expressible 2 nucleotide sequences respectively encoding for an expression product (eg, a respective translatable mRNA), wherein the sequences are under common promoter control.

5. The vector of paragraph 4, wherein the first sequence is between the promoter and the second sequence in the operon.

6. The vector of paragraph 4 or 5, wherein the operon comprises no Cas-encoding nucleotide sequences between the promoter and the first nucleotide sequence.

Optionally, the Cas3 is a Cas3 encoded by a CRISPR/Cas locus of a first bacterial or archaeal species, wherein in the locus the Cas3-encoding sequence is 3' of Cascade protein-encoding sequences (ie, the latter are between the Cas3 and the 5'-most promoter of the locus).

Optionally, the Cas3 is a ygcB protein (eg, wherein the production strain cell and/or host target cell is an *E. coli*).

Optionally, the Cascade proteins comprise or consist of cas5 (casD, csy2)

cas6 (cas6f, cse3, casE)

cas7 (csc2, csy3, cse4, casC)

cas8 (casA, cas8a1, cas8b1, cas8c, cas10d, cas8e, cse1, cas8f, csy1).

Optionally herein the promoter and the Cas3-encoding sequence are spaced no more than 150, 100, 50, 40, 30, 20 or 10 bp apart, eg, from 30-45, or 30-40, or 39 or around 39 bp apart.

Optionally herein a ribosome binding site and the Cas3-encoding sequence are spaced no more than 20, 15, 14, 13, 12, 11, 10, 9, 8, 7, 6, 4 or 3 bp apart, eg, from 10-5, 6 or around 6 bp apart.

7. The vector of any one of paragraphs 4 to 6, wherein the operon comprises (in 5' to 3' direction) the promoter, the first sequence and the second sequence.

8. The vector of any preceding paragraph, wherein each promoter is a constitutive promoter.

9. The vector of any one of paragraphs 1 to 7, wherein the promoter is repressible (optionally repressible by a tetracycline repressor or lac repressor).

10. The vector of any one of paragraphs 1 to 7, wherein the promoter is inducible.

11. The vector of any preceding paragraph, wherein the first sequence is under the control of a weak promoter.

12. The vector of any one of paragraphs 1 to 7, wherein the first sequence is under the control of a medium strength promoter.

13. The vector of any one of paragraphs 1 to 7, wherein the first sequence is under the control of a strong promoter.

In an example, the promoter is in combination with a Shine-Dalgarno sequence comprising the sequence 5'-aaagaggagaaa-3' (SEQ ID NO: 5) or a ribosome binding site homologue thereof.

14. The vector of any one of paragraphs 1 to 7, wherein the first sequence is under the control of a promoter that has an Anderson Score (AS) of $AS \geq 0.5$.

See Table 2 for more information on Anderson Scores in relation to promoters.

15. The vector of any one of paragraphs 1 to 7, wherein the first sequence is under the control of a promoter that has an Anderson Score (AS) of $0.5 > AS > 0.1$.

16. The vector of any one of paragraphs 1 to 7, wherein the first sequence is under the control of a promoter that has an Anderson Score (AS) of ≤ 0.1 .

17. The vector of any one of paragraphs 1 to 7, wherein the first sequence (and optionally the second sequence) is under the control of a promoter and translation initiation site (TIS) combination that is capable of producing expression of green fluorescent protein (GFP) from a first expression operating unit (EOU) in *E. coli* strain BW25113 cells with a fluorescence of from 0.5 to 4 times the fluorescence produced in *E. coli* strain BW25113 cells using a second EOU comprising

a P10 promoter (SEQ ID NO: 1) combined with a BCD14 TIS (SEQ ID NO: 2), wherein the EOUs differ only in their promoter and TIS combinations, wherein each EOU comprises (in 5' to 3' direction) an upstream initiator, the respective promoter, the respective TIS, a nucleotide sequence encoding GFP, a 3' UTR, a transcription terminator and a downstream insulator.

18. The vector of paragraph 17, wherein fluorescence using the first EOU is 0.5 to 2 times the fluorescence using the second EOU.

For example, fluorescence using the first EOU is 0.5 to X times the fluorescence using the second EOU, wherein X is from 3.0 to 1.0, eg, 3, 2.5, 2, 1.5 or 1, wherein fluorescence is determined using excitation at 481 nm and emission at 507 nm. Optionally, *E coli* cultures at OD600 of 0.3-0.5 in the exponential growth phase are used.

For example, the upstream insulator, the nucleotide sequence encoding GFP, 3' UTR, transcription terminator and downstream insulator of each EOU are as disclosed in Mutalik et al (2013). For example, the upstream insulator, the nucleotide sequence encoding GFP, 3' UTR, transcription terminator and downstream insulator of each EOU are corresponding sequences of SEQ ID NO: 4. For example, the *E coli* is *E. coli* BW25113 is grown in MOPS EZ Rich Medium (Teknova) supplemented with 50 µg/ml kanamycin (kan) at 37° C., shaken at 900 r.p.m. For example, each EOU is comprised by a medium copy plasmid, eg, a plasmid derived from pFAB217 comprising a p15A replication origin and a kan resistance gene.

19. The vector of any preceding paragraph, wherein the vector comprises an origin of replication that is operable in the host cell.

20. The vector of any preceding paragraph, wherein the vector comprises an origin of replication that is operable in a bacterial cell of a vector production strain, wherein the Cas3 is not operable in the production strain cell to target and cut a chromosomal sequence thereof.

An example of a production strain cell is an *E coli* cell. A production strain cell is a cell that is used to amplify DNA encoding Cas (and optionally other components of a CRISPR/Cas system). Usefully, the strain may package the amplified DNA into transduction particles that are may be isolated to produce a composition that can be contacted with a population of target host cells (eg, bacterial, archaeal, prokaryotic, eukaryotic, human, animal, mammal, rodent, mouse, rat, rabbit, *Xenopus*, fish, bird, amphibian, insect, plant, amoeba or algae cells) wherein the DNA is introduced into the cells for expression of the Cas (and optional other CRISPR/Cas system components), wherein the Cas is guided to a protospacer target sequence in the host cells and modifies (eg, cuts) the sequence. In another example, the amplified DNA isolated from a population of production strain cells and is combined with a delivery vehicle (eg, a carrier bacterium, nanoparticle or liposome), wherein the delivery vehicle can be contacted with a population of target host cells (eg, bacterial, archaeal, prokaryotic, eukaryotic, human, animal, mammal, rodent, mouse, rat, rabbit, *Xenopus*, fish, bird, amphibian, insect, plant, amoeba or algae cells) wherein the DNA is introduced into the cells for expression of the Cas (and optional other CRISPR/Cas system components), wherein the Cas is guided to a

protospacer target sequence in the host cells and modifies (eg, cuts) the sequence.

21. The vector of paragraph 20, wherein the first sequence is under the control of a promoter that is capable of controlling expression of the Cas3 at a level that is not toxic to the production strain cell.

In an example, substantially no production strain cells are killed when the Cas3-encoding sequence is amplified therein. In another example, no more than 40, 30, 20, 10, 5, 4, 3, 2, or 1% of production strain cells are killed when the Cas3-encoding sequence is amplified therein. For example this is in a 1, 2, 3, 4, 5, 6, 7, 8 9 10, 12 or 24 hour period of culturing the cells.

22. The vector of paragraph 20, wherein the first sequence is under the control of a promoter that controls expression of the Cas3 in the production strain cell such that the cell is capable of growth and propagation sufficient to produce at least 1000 copies of the vector.

For example this is in a 1, 2, 3, 4, 5, 6, 7, 8 9 10, 12 or 24 hour period of culturing the cells. For example, at least 10^4 , 10^5 , 10^6 , 10^7 , 10^8 , 10^9 , 10^{10} , 10^{11} , 10^{12} , 10^{13} , 10^{14} , 10^{15} , 10^{16} , 10^{17} or 10^{18} copies of the vector are produced per 10^3 , 10^4 , 10^5 , 10^6 , 10^7 , 10^8 , 10^9 , 10^{10} , 10^{11} , 10^{12} , 10^{13} , 10^{14} , 10^{15} , 10^{16} , 10^{17} production strain cells respectively.

23. The vector of any one of paragraphs 20 to 22, wherein the cell is capable of at least 2 or 3 logs of expansion when the vector is comprised therein.

For example, this is in a 1, 2, 3, 4, 5, 6, 7, 8 9 10, 12 or 24 hour period of culturing the cells.

24. The vector of any preceding paragraph, wherein the vector is a high copy number vector.

25. The vector of any preceding paragraph, wherein the first nucleotide sequence or operon is comprised by a mobile genetic element.

Suitable mobile genetic elements, eg, transposons, are disclosed in WO2016177682 and US20170246221, the disclosures of which are explicitly incorporated herein for possible use in the invention and for providing one or more features for the claims herein.

26. The vector of any preceding paragraph, wherein the vector is devoid of a Cas adaption module. For example, the vector is devoid of nucleotide sequences encoding a Cas1, Cas2 and/or Cas4.

27. The vector of any preceding paragraph, wherein the vector is devoid of nucleotide sequence encoding one, more or all of a Cas1, Cas2, Cas4, Cas6 (optionally Cas6f), Cas7 and Cas 8 (optionally Cas8f).

28. The vector of any preceding paragraph, wherein the vector is devoid of a sequence encoding a Cas6 (optionally a Cas6f).

29. The vector of any one of paragraphs 1 to 27, wherein the module encodes a Cas6 (optionally a Cas6f).

30. The vector of any preceding paragraph, wherein the vector comprises (optionally in 5' to 3' direction) nucleotide sequence encoding one, more or all of Cas 11, Cas7 and Cas8a1.

31. The vector of paragraph 30, wherein the vector comprises nucleotide sequence encoding Cas3' and/or Cas3" (optionally wherein the nucleotide sequences encoding the Cas3' and/or Cas3" are between the promoter and the sequence(s) recited in paragraph 30).

In one embodiment, the vector comprises nucleotide sequences (in 5' to 3' direction) that encode a Cas3 (eg, Cas3' and/or Cas3"), Cas11, Cas7 and Cas8a1. Optionally, a nucleotide sequence encoding Cas6 is between the Cas3 sequence(s) and the Cas11 sequence. Option-

ally, the vector comprises a Type IA CRISPR array or one or more nucleotide sequences encoding single guide RNA(s) (gRNA(s)), wherein the array and each gRNA comprises repeat sequence that is cognate with the Cas3. Thus, the array is operable in a host cell when the vector has been introduced into the cell for production of guide RNAs, wherein the guide RNAs are operable with the Cas and Cascade proteins to target and modify (eg, cut) a target nucleotide sequence in the host cell, optionally thereby killing the host cell. Similarly, the single guide RNAs encoded by the vector in one embodiment are operable with the Cas and Cascade proteins to target and modify (eg, cut) a target nucleotide sequence in the host cell, optionally thereby killing the host cell.

32. The vector of paragraph 30 or 31, wherein the host cell comprises a Type IA CRISPR array that is cognate with the Cas3.

33. The vector of paragraph 30 or 31, wherein the host cell comprises an endogenous Type IB, C, U, D, E or F CRISPR/Cas system.

34. The vector of any one of paragraphs 1 to 29, wherein the vector comprises (optionally in 5' to 3' direction) nucleotide sequence encoding one, more or all of Cas8b1, Cas7 and Cas5.

35. The vector of paragraph 34, wherein the vector comprises a nucleotide sequence encoding Cas3 between the promoter and the sequence(s) recited in paragraph 34.

In one embodiment, the vector comprises nucleotide sequences (in 5' to 3' direction) that encode a Cas3, Cas8b1, Cas7 and Cas5. Optionally, a nucleotide sequence encoding Cas6 is between the Cas3 sequence (s) and the Cas8b1 sequence. Optionally, the vector comprises a Type IB CRISPR array or one or more nucleotide sequences encoding single guide RNA(s) (gRNA(s)), wherein the array and each gRNA comprises repeat sequence that is cognate with the Cas3. Thus, the array is operable in a host cell when the vector has been introduced into the cell for production of guide RNAs, wherein the guide RNAs are operable with the Cas and Cascade proteins to target and modify (eg, cut) a target nucleotide sequence in the host cell, optionally thereby killing the host cell. Similarly, the single guide RNAs encoded by the vector in one embodiment are operable with the Cas and Cascade proteins to target and modify (eg, cut) a target nucleotide sequence in the host cell, optionally thereby killing the host cell.

36. The vector of paragraph 34 or 35, wherein the host cell comprises a Type IB CRISPR array that is cognate with the Cas3.

37. The vector of paragraph 34 or 35, wherein the host cell comprises an endogenous Type IA, C, U, D, E or F CRISPR/Cas system.

38. The vector of any one of paragraphs 1 to 29, wherein the vector comprises (optionally in 5' to 3' direction) nucleotide sequence encoding one, more or all of Cas5, Cas8c and Cas7.

39. The vector of paragraph 38, wherein the vector comprises a nucleotide sequence encoding Cas3 between the promoter and the sequence(s) recited in paragraph 38.

In one embodiment, the vector comprises nucleotide sequences (in 5' to 3' direction) that encode a Cas3, Cas5, Cas8c and Cas7. Optionally, a nucleotide sequence encoding Cas6 is between the Cas3

sequence(s) and the Cas5 sequence. Optionally, the vector comprises a Type IC CRISPR array or one or more nucleotide sequences encoding single guide RNA(s) (gRNA(s)), wherein the array and each gRNA comprises repeat sequence that is cognate with the Cas3. Thus, the array is operable in a host cell when the vector has been introduced into the cell for production of guide RNAs, wherein the guide RNAs are operable with the Cas and Cascade proteins to target and modify (eg, cut) a target nucleotide sequence in the host cell, optionally thereby killing the host cell. Similarly, the single guide RNAs encoded by the vector in one embodiment are operable with the Cas and Cascade proteins to target and modify (eg, cut) a target nucleotide sequence in the host cell, optionally thereby killing the host cell.

40. The vector of paragraph 38 or 39, wherein the host cell comprises a Type IC CRISPR array that is cognate with the Cas3.

41. The vector of paragraph 38 or 39, wherein the host cell comprises an endogenous Type IA, B, U, D, E or F CRISPR/Cas system.

42. The vector of any one of paragraphs 1 to 29, wherein the vector comprises (optionally in 5' to 3' direction) nucleotide sequence encoding one, more or all of Cas8U2, Cas7, Cas5 and Cas6.

43. The vector of paragraph 42, wherein the vector comprises a nucleotide sequence encoding Cas3 between the promoter and the sequence(s) recited in paragraph 42.

In one embodiment, the vector comprises nucleotide sequences (in 5' to 3' direction) that encode a Cas3, Cas8U2, Cas7, Cas5 and Cas6. Optionally, a nucleotide sequence encoding Cas6 is between the Cas3 sequence(s) and the Cas8U2 sequence. Optionally, the vector comprises a Type IU CRISPR array or one or more nucleotide sequences encoding single guide RNA(s) (gRNA(s)), wherein the array and each gRNA comprises repeat sequence that is cognate with the Cas3. Thus, the array is operable in a host cell when the vector has been introduced into the cell for production of guide RNAs, wherein the guide RNAs are operable with the Cas and Cascade proteins to target and modify (eg, cut) a target nucleotide sequence in the host cell, optionally thereby killing the host cell. Similarly, the single guide RNAs encoded by the vector in one embodiment are operable with the Cas and Cascade proteins to target and modify (eg, cut) a target nucleotide sequence in the host cell, optionally thereby killing the host cell.

44. The vector of paragraph 42 or 43, wherein the host cell comprises a Type IU CRISPR array that is cognate with the Cas3.

45. The vector of paragraph 42 or 43, wherein the host cell comprises an endogenous Type IA, B, C, D, E or F CRISPR/Cas system.

46. The vector of any one of paragraphs 1 to 29, wherein the vector comprises (optionally in 5' to 3' direction) nucleotide sequence encoding one, more or all of Cas10d, Cas7 and Cas5.

47. The vector of paragraph 46, wherein the vector comprises a nucleotide sequence encoding Cas3' and/or Cas3" (optionally wherein the nucleotide sequences encoding the Cas3' and/or Cas3" are between the promoter and the sequence(s) recited in paragraph 46).

In one embodiment, the vector comprises nucleotide sequences (in 5' to 3' direction) that encode a Cas3,

- Cas10d, Cas7 and Cas5. Optionally, a nucleotide sequence encoding Cas6 is between the Cas3 sequence(s) and the Cas10d sequence. Optionally, the vector comprises a Type ID CRISPR array or one or more nucleotide sequences encoding single guide RNA(s) (gRNA(s)), wherein the array and each gRNA comprises repeat sequence that is cognate with the Cas3. Thus, the array is operable in a host cell when the vector has been introduced into the cell for production of guide RNAs, wherein the guide RNAs are operable with the Cas and Cascade proteins to target and modify (eg, cut) a target nucleotide sequence in the host cell, optionally thereby killing the host cell. Similarly, the single guide RNAs encoded by the vector in one embodiment are operable with the Cas and Cascade proteins to target and modify (eg, cut) a target nucleotide sequence in the host cell, optionally thereby killing the host cell.
48. The vector of paragraph 46 or 47, wherein the host cell comprises a Type ID CRISPR array that is cognate with the Cas3.
49. The vector of paragraph 46 or 47, wherein the host cell comprises an endogenous Type IA, B, C, U, E or F CRISPR/Cas system.
50. The vector of any one of paragraphs 1 to 29, wherein the vector comprises (optionally in 5' to 3' direction) nucleotide sequence encoding one, more or all of Cas8e, Cas11, Cas7, Cas5 and Cas6.
51. The vector of paragraph 50, wherein the vector comprises a nucleotide sequence encoding Cas3 between the promoter and the sequence(s) recited in paragraph 50.
- In one embodiment, the vector comprises nucleotide sequences (in 5' to 3' direction) that encode a Cas3, Cas8e, Cas11, Cas7, Cas5 and Cas6. Optionally, a nucleotide sequence encoding Cas6 is between the Cas3 sequence(s) and the Cas11 sequence. Optionally, the vector comprises a Type IE CRISPR array or one or more nucleotide sequences encoding single guide RNA(s) (gRNA(s)), wherein the array and each gRNA comprises repeat sequence that is cognate with the Cas3. Thus, the array is operable in a host cell when the vector has been introduced into the cell for production of guide RNAs, wherein the guide RNAs are operable with the Cas and Cascade proteins to target and modify (eg, cut) a target nucleotide sequence in the host cell, optionally thereby killing the host cell. Similarly, the single guide RNAs encoded by the vector in one embodiment are operable with the Cas and Cascade proteins to target and modify (eg, cut) a target nucleotide sequence in the host cell, optionally thereby killing the host cell.
52. The vector of paragraph 50 or 51, wherein the host cell comprises a Type IE CRISPR array that is cognate with the Cas3.
53. The vector of paragraph 50 or 51, wherein the host cell comprises an endogenous Type IA, B, C, D, U or F CRISPR/Cas system.
54. The vector of any one of paragraphs 1 to 29, wherein the vector comprises (optionally in 5' to 3' direction) nucleotide sequence encoding one, more or all of Cas8f, Cas5, Cas7 and Cas6f.
55. The vector of paragraph 54, wherein the vector comprises a nucleotide sequence encoding Cas3 between the promoter and the sequence(s) recited in paragraph 54, wherein the vector is devoid of nucleotide

- sequence encoding further Cas between the promoter and the sequence encoding the Cas3.
- In one embodiment, the vector comprises nucleotide sequences (in 5' to 3' direction) that encode a Cas3, Cas8f, Cas5, Cas7 and Cas6f. Optionally, a nucleotide sequence encoding Cas6 is between the Cas3 sequence(s) and the Cas8f sequence. Optionally, the vector comprises a Type IF CRISPR array or one or more nucleotide sequences encoding single guide RNA(s) (gRNA(s)), wherein the array and each gRNA comprises repeat sequence that is cognate with the Cas3. Thus, the array is operable in a host cell when the vector has been introduced into the cell for production of guide RNAs, wherein the guide RNAs are operable with the Cas and Cascade proteins to target and modify (eg, cut) a target nucleotide sequence in the host cell, optionally thereby killing the host cell. Similarly, the single guide RNAs encoded by the vector in one embodiment are operable with the Cas and Cascade proteins to target and modify (eg, cut) a target nucleotide sequence in the host cell, optionally thereby killing the host cell.
56. The vector of paragraph 54 or 55, wherein the host cell comprises a Type IF CRISPR array that is cognate with the Cas3.
57. The vector of paragraph 54 or 55, wherein the host cell comprises an endogenous Type IA, B, C, D, U or E CRISPR/Cas system.
58. The vector of any one of paragraphs 1 to 29, wherein the Cas and Cascade are Type IA Cas and Cascade proteins.
59. The vector of any one of paragraphs 1 to 29, wherein the Cas and Cascade are Type IB Cas and Cascade proteins.
60. The vector of any one of paragraphs 1 to 29, wherein the Cas and Cascade are Type IC Cas and Cascade proteins.
61. The vector of any one of paragraphs 1 to 29, wherein the Cas and Cascade are Type ID Cas and Cascade proteins.
62. The vector of any one of paragraphs 1 to 29, wherein the Cas and Cascade are Type IE Cas and Cascade proteins.
63. The vector of any one of paragraphs 1 to 29, wherein the Cas and Cascade are Type IF Cas and Cascade proteins.
64. The vector of any one of paragraphs 1 to 29, wherein the Cas and Cascade are Type IU Cas and Cascade proteins.
65. The vector of any one of paragraphs 1 to 29, wherein the Cas and Cascade are *E. coli* (optionally Type IE or IF) Cas and Cascade proteins, optionally wherein the *E. coli* is ESBL-producing *E. coli* or *E. coli* ST131-O25b:H4.
66. The vector of any one of paragraphs 1 to 29, wherein the Cas and Cascade are *Clostridium* (eg, *C. difficile*) Cas and Cascade proteins, optionally *C. difficile* resistant to one or more antibiotics selected from aminoglycosides, lincomycin, tetracyclines, erythromycin, clindamycin, penicillins, cephalosporins and fluoroquinolones.
67. The vector of any one of paragraphs 1 to 29, wherein the Cas and Cascade are *Pseudomonas aeruginosa* Cas and Cascade proteins, optionally *P. aeruginosa* resistant to one or more antibiotics selected from carbapenems, aminoglycosides, cefepime, ceftazidime, fluoroquinolones, piperacillin and tazobactam.

68. The vector of any one of paragraphs 1 to 29, wherein the Cas and Cascade are *Klebsiella pneumoniae* (eg, carbapenem-resistant *Klebsiella pneumoniae* or Extended-Spectrum Beta-Lactamase (ESBL)-producing *K pneumoniae*) Cas and Cascade proteins. 5
69. The vector of any one of paragraphs 1 to 29, wherein the Cas and Cascade are *E coli*, *C difficile*, *P aeruginosa*, *K pneumoniae*, *P furiosus* or *B halodurans* Cas and Cascade proteins.
70. The vector of any preceding paragraph, wherein the Cas3 is a Cas3 of a CRISPR/Cas locus of a first bacterial or archaeal species, wherein the distance between the Cas3-encoding sequence of the locus and its cognate promoter is further than the distance between the Cas3-encoding sequence and the respective promoter comprised by the vector. 10
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The cognate promoter here is the one that controls expression of Cas3 in the wild-type locus.
71. The vector of any preceding paragraph, wherein the distance between the promoter and the Cas3-encoding sequence and/or Cascade protein-encoding sequence(s) is shorter than in a corresponding wild-type Type I locus. 20
- A corresponding locus is a wild-type locus of a bacterial or archaeal species or strain that comprises an endogenous CRISPR/Cas system encoding the Cas3 and/or Cascade proteins of the type that are also encoded by the vector. Thus, when the vector comprises an operon, the operon may comprise Cas3- and Cascade-encoding nucleotide sequences that are not in a natural configuration. 25
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72. The vector of any preceding paragraph, wherein the vector comprises (i) a CRISPR array for producing crRNAs in the host cell and/or (ii) one or more nucleotide sequences encoding one or more single guide RNAs (gRNAs), wherein the crRNAs or gRNAs are cognate to the Cas3 (and optionally cognate to the Cascade proteins). 35
73. The vector of paragraph 72 when dependent from paragraph 4, wherein the array or gRNA-encoding sequence(s) are comprised by the operon and under the control of the promoter. 40
74. The vector of paragraph 72, wherein the array or gRNA-encoding sequence(s) are under the control of a promoter that is different from the promoter that controls the expression of the Cas3. 45
75. The vector of any one of paragraphs 72 to 74, wherein one or more of the crRNAs or gRNAs comprises a spacer sequence that is capable of hybridising to a target nucleotide sequence of the host cell, wherein the target sequence is adjacent a PAM, the PAM being cognate to the Cas3. 50
- Thus, the spacer hybridises to the protospacer to guide the Cas3 to the protospacer. Optionally, the Cas3 cuts the protospacer, eg, using exo- and/or endonuclease activity of the Cas3. Optionally, the Cas3 removes a plurality (eg, at least 2, 3, 4, 5, 6, 7, 8, 9 or 10) nucleotides from the protospacer. 55
76. The vector of paragraph 75, wherein the target sequence is a chromosomal sequence of the host cell. 60
77. The vector of paragraph 75 or 76, wherein the Cas3 is operable to cut the target sequence.
78. The vector of any preceding paragraph, wherein the vector is a plasmid or phagemid.
79. A delivery vehicle comprising the vector of any preceding paragraph, wherein the delivery vehicle is capable of delivering the vector into the host cell. 65

80. The vehicle of paragraph 79, wherein the delivery vehicle is a phage, non-replicative transduction particle, nanoparticle carrier, bacterium or liposome.
- The phage or particles comprise phage coat proteins encapsidating DNA, wherein the DNA comprises the vector. Suitable examples of phage and particles are disclosed in U.S. Ser. No. 15/985,658 (and its equivalent publication by USPTO) the disclosures of which are incorporated herein by reference for possible use in the invention and for providing one or more features that may be included in the claims herein. Phage or particle is capable of infecting the cell, thereby introducing the vector into the cell.
81. The vector or vehicle of any preceding paragraph, wherein the host cell is a bacterial or archaeal cell, optionally, the host cell is a *C difficile*, *P aeruginosa*, *K pneumoniae* (eg, carbapenem-resistant *Klebsiella pneumoniae* or Extended-Spectrum Beta-Lactamase (ESBL)-producing *K pneumoniae*), *E coli* (eg, ESBL-producing *E. coli*, or *E. coli* ST131-O25b:H4), *H pylori*, *S pneumoniae* or *S aureus* cell.
82. The vector or vehicle of any preceding paragraph for administration to a human or animal subject for treating or reducing the risk of a disease or condition in the subject.
83. The vector or vehicle of paragraph 82, wherein the disease or condition is an infection of the subject with host cells (eg, bacterial cells), or wherein the disease or condition is mediated by host cells (eg, bacterial cells).
84. A pharmaceutical composition comprising the vector or vehicle of any preceding paragraph and a pharmaceutically acceptable diluent, excipient or carrier.
85. A method of amplifying copies of a DNA encoding a functional Cas protein (optionally a Cas nuclease) in a bacterial or archaeal production strain of cells, the method comprising
- Providing production strain cells, each cell comprising a copy of said DNA, wherein each DNA comprises a nucleotide sequence encoding said Cas, wherein the nucleotide sequence is under the control of a promoter for controlling the expression of the Cas in the production strain cell, the DNA comprising an origin of replication that is operable in the cell for replication of the DNA;
 - Culturing the cells to allow replication of the DNA, whereby the DNA is amplified; and
 - Optionally isolating copies of the DNA,
86. The method of paragraph 85, wherein the promoter is a constitutive promoter.
87. The method of paragraph 85, wherein the promoter is repressible (optionally repressible by a tetracycline repressor or a lac repressor).
88. The method of paragraph 85, wherein the promoter is inducible.
89. The method of any one of paragraphs 85 to 88, wherein the promoter is a medium strength promoter.
90. The method of any one of paragraphs 85 to 89, wherein the promoter has an Anderson Score (AS) of $0.5 > AS > 0.1$.
91. The method of any one of paragraphs 85 to 90, wherein the nucleotide sequence encoding said Cas is under the control of a promoter and translation initiation site (TIS) combination that is capable of producing expression of green fluorescent protein (GFP) from a first expression operating unit (EOU) in *E. coli* strain BW25113 cells with a fluorescence of from 0.5 to 4 times the fluorescence produced in *E. coli* strain

- BW25113 cells using a second EOU comprising a P10 promoter (SEQ ID NO: 1) combined with a BCD14 TIS (SEQ ID NO: 2), wherein the EOUs differ only in their promoter and TIS combinations, wherein each EOU comprises (in 5' to 3' direction) an upstream initiator, the respective promoter, the respective TIS, a nucleotide sequence encoding GFP, a 3' UTR, a transcription terminator and a downstream insulator.
92. The method of paragraph 91, wherein fluorescence using the first EOU is 0.5 to 2 times the fluorescence using the second EOU.
93. The method of any one of paragraphs 85 to 92, wherein the nuclease is Cas3 and optionally the DNA or cell encodes cognate Cascade proteins and/or one or more crRNAs that are operable for Cas nuclease targeting.
- For example, the targeting is targeting of the Cas to a protospacer sequence comprised by a host cell chromosome or an episome thereof. In another example the targeting is in a recombineering method and the Cas is targeted to a protospacer sequence of a DNA that has been introduced into or amplified in the host cell. In an example of such recombineering, the host cell is an *E. coli* cell.
94. The method of any one of paragraphs 85 to 92, wherein the Cas is a Cas9.
95. The method of any one of paragraphs 85 to 92, wherein the Cas is a Type IIIA csm protein or a Type IIIB cmr protein.
96. The method of any one of paragraphs 85 to 92, wherein the Cas is a Csf1.
97. The method of any one of paragraphs 85 to 92, wherein the Cas is a Cpf1.
98. The method of any one of paragraphs 85 to 92, wherein the Cas is a Cas13 (optionally Cas13a or Cas13b).
99. The method of any one of paragraphs 85 to 92, wherein the Cas is selected from a Cas3, Cas8a, Cas5, Cas8b, Cas8c, Cas10d, Cse1, Cse2, Csy1, Csy2, Csy3, GSU0054, Cas10, Csm2, Cmr5, Cas10, Csx11, Csx10, Csf1, Cas9, Csn2, Cas4, Cpf1, C2c1, C2c3, Cas13a, Cas13b and Cas13c.
100. The method of any one of paragraphs 85 to 99, wherein the production strain cells comprise a helper phage genome that is inducible to produce phage coat proteins in the cells, wherein the method further comprises inducing production of the phage proteins and causing packaging of the amplified DNA into phage particles or non-self-replicative transduction particles, and further isolating the phage or transduction particles and optionally formulating the particles into a pharmaceutical composition for administration to a human or animal subject for treating or reducing the risk of a disease or condition in the subject.
101. The method of paragraph 100, wherein the particles are capable of infecting target host cells in the subject and transducing the cells with the DNA, wherein the Cas and crRNAs (or gRNAs) encoded by the DNA are expressed in the cells, the crRNAs or (gRNAs) being operable to guide the Cas to a target nucleotide sequence (optionally a chromosomal sequence) comprised by the cells, wherein the Cas cuts the target sequences in the cells, thereby killing host cells and treating or reducing the risk of the disease or condition.
102. The method of paragraph 101, wherein the host cells are bacterial or archaeal cells, optionally, the host cells are *C. difficile*, *P. aeruginosa*, *K. pneumoniae* (eg, car-

- bapenem-resistant *Klebsiella pneumoniae* or Extended-Spectrum Beta-Lactamase (ESBL)-producing *K. pneumoniae*, *E. coli* (eg, ESBL-producing *E. coli*, or *E. coli* ST131-O25b:H4), *H. pylori*, *S. pneumoniae* or *S. aureus* cells.
103. The method of any one of paragraphs 85 to 102, wherein each DNA is comprised by a high copy number vector, optionally a high copy number plasmid (an optionally the promoter is a constitutive promoter).
104. The method of any one of paragraphs 85 to 103, wherein each DNA is comprised by a vector or vehicle according to any one of paragraphs 1 to 83.
105. Use of an attenuated strength promoter in a DNA construct comprising a nucleotide sequence encoding a functional Cas protein (optionally a Cas nuclease) that is under the control of the promoter, in a method of amplifying copies of the DNA in a population of bacterial or archaeal production strain cells, the method comprising culturing the cells to allow replication of the DNA thereby amplifying the DNA in the cells, for enhancing the yield of amplified DNA produced by the production host cells.
- Thus, said enhancing may be relative to the yield produced using a strong promoter, eg, a strong constitutive promoter (for example a promoter having an Anderson Score (AS) of $AS \geq 0.5$). In another example, the strong promoter is a promoter comprised by a promoter and translation initiation site (TIS) combination that is capable of producing expression of green fluorescent protein (GFP) from a first expression operating unit (EOU) in *E. coli* strain BW25113 cells with a fluorescence of >4 times the fluorescence produced in *E. coli* strain BW25113 cells using a second EOU comprising a P10 promoter (SEQ ID NO: 1) combined with a BCD14 TIS (SEQ ID NO: 2), wherein the EOUs differ only in their promoter and TIS combinations, wherein each EOU comprises (in 5' to 3' direction) an upstream initiator, the respective promoter, the respective TIS, a nucleotide sequence encoding GFP, a 3' UTR, a transcription terminator and a downstream insulator.
106. The use of paragraph 105, wherein the use is for enhancing said yield by
- (d) reducing toxicity of the Cas in the production strain;
 - (e) reducing mutation of the DNA (optionally the Cas-encoding sequence) in the production strain;
 - (f) promoting production cell viability during the amplification of the DNA; and/or
 - (g) reducing the occurrence of Cas cutting of DNA (optionally cutting of production host cell chromosomal DNA or said DNA construct).
107. Use of an attenuated strength promoter in a DNA construct comprising a nucleotide sequence encoding a functional Cas protein (optionally a Cas nuclease) that is under the control of the promoter, in a method of amplifying copies of the DNA in a population of bacterial or archaeal production strain cells, the method comprising culturing the cells to allow replication of the DNA thereby amplifying the DNA in the cells, for reducing toxicity of the Cas in the production strain.
108. Use of an attenuated strength promoter in a DNA construct comprising a nucleotide sequence encoding a functional Cas protein (optionally a Cas nuclease) that is under the control of the promoter, in a method of amplifying copies of the DNA in a population of bacterial or archaeal production strain cells, the method comprising culturing the cells to allow replication of the DNA thereby amplifying the DNA in the cells, for

- reducing mutation of the DNA (optionally the Cas-encoding sequence) in the production strain.
109. Use of an attenuated strength promoter in a DNA construct comprising a nucleotide sequence encoding a functional Cas protein (optionally a Cas nuclease) that is under the control of the promoter, in a method of amplifying copies of the DNA in a population of bacterial or archaeal production strain cells, the method comprising culturing the cells to allow replication of the DNA thereby amplifying the DNA in the cells, for promoting production cell viability during the amplification of the DNA.
110. Use of an attenuated strength promoter in a DNA construct comprising a nucleotide sequence encoding a functional Cas protein (optionally a Cas nuclease) that is under the control of the promoter, in a method of amplifying copies of the DNA in a population of bacterial or archaeal production strain cells, the method comprising culturing the cells to allow replication of the DNA thereby amplifying the DNA in the cells, for reducing the occurrence of Cas cutting of DNA.
111. A method for enhancing the yield of amplified copies of a DNA construct in a population of bacterial or archaeal production strain cells, wherein the construct comprises a nucleotide sequence encoding a functional Cas protein (optionally a Cas nuclease) that is under the control of a promoter, the method comprising culturing the cells to allow replication of the DNA thereby amplifying the DNA in the cells, wherein the promoter is an attenuated strength promoter.
112. A method for reducing toxicity of a functional Cas protein (optionally a Cas nuclease) in a population of bacterial or archaeal production strain cells in a process of amplifying copies of a DNA construct, wherein the construct comprises a nucleotide sequence encoding the Cas and the sequence is under the control of a promoter, the method comprising culturing the cells to allow replication of the DNA thereby amplifying the DNA in the cells, wherein the promoter is an attenuated strength promoter.
113. A method for reducing mutation of a DNA construct encoding a functional Cas protein (optionally a Cas nuclease) in a population of bacterial or archaeal production strain cells in a process of amplifying copies of the construct, wherein the construct comprises a nucleotide sequence encoding the Cas and the sequence is under the control of a promoter, the method comprising culturing the cells to allow replication of the DNA thereby amplifying the DNA in the cells, wherein the promoter is an attenuated strength promoter.
114. A method for promoting production cell viability of a population of bacterial or archaeal production strain cells in a process of amplifying copies of a DNA construct comprised by the cells, wherein the construct comprises a nucleotide sequence encoding a functional Cas protein (optionally a Cas nuclease) and the sequence is under the control of a promoter, the method comprising culturing the cells to allow replication of the DNA thereby amplifying the DNA in the cells, wherein the promoter is an attenuated strength promoter.
115. A method for reducing the occurrence of Cas nuclease cutting of a DNA construct in a population of bacterial or archaeal production strain cells in a process of amplifying copies of the construct, wherein the construct comprises a nucleotide sequence encoding the Cas and the sequence is under the control of a

- promoter, the method comprising culturing the cells to allow replication of the DNA thereby amplifying the DNA in the cells, wherein the promoter is an attenuated strength promoter.
116. The use of paragraph 108 or 110, or the method of paragraph 113 or 115, wherein the mutation or cutting is mutation or cutting of host cell chromosomal DNA or the construct DNA.
117. The use or method of any one of paragraphs 105 to 116, wherein the promoter is a constitutive promoter.
118. The use or method of any one of paragraphs 105 to 117, wherein the promoter is repressible (optionally repressible by a tetracycline repressor or a lac repressor).
- In an example, the promoter is a constitutive promoter and optionally the DNA is comprised by a high copy number plasmid or phagemid.
119. The use or method of any one of paragraphs 105 to 118, wherein the promoter is $P_{LtetO-1}$, $P_{LlacO-1}$ or a repressible homologue thereof.
- $P_{LlacO-1}$ is repressed by lac repressor (LacR). $P_{LtetO-1}$ is repressed by tet repressor (TetR).
120. The use or method of any one of paragraphs 105 to 119, wherein the promoter is a medium strength promoter.
121. The use or method of any one of paragraphs 105 to 120, wherein the promoter has an Anderson Score (AS) of $0.5 > AS > 0.1$.
122. The use or method of any one of paragraphs 105 to 121, wherein the nucleotide sequence encoding said Cas is under the control of a promoter and translation initiation site (TIS) combination that is capable of producing expression of green fluorescent protein (GFP) from a first expression operating unit (EOU) in *E. coli* strain BW25113 cells with a fluorescence of from 0.5 to 4 times the fluorescence produced in *E. coli* strain BW25113 cells using a second EOU comprising a P10 promoter (SEQ ID NO: 1) combined with a BCD14 TIS (SEQ ID NO: 2), wherein the EOUs differ only in their promoter and TIS combinations, wherein each EOU comprises (in 5' to 3' direction) an upstream initiator, the respective promoter, the respective TIS, a nucleotide sequence encoding GFP, a 3' UTR, a transcription terminator and a downstream insulator.
123. The use or method of paragraph 122, wherein fluorescence using the first EOU is 0.5 to 2 times the fluorescence using the second EOU.
124. The use or method of any one of paragraphs 105 to 123, wherein the nuclease is Cas3 and optionally the DNA construct encodes cognate Cascade proteins.
125. The use or method of any one of paragraphs 105 to 123, wherein the Cas is a Cas9.
126. The use or method of any one of paragraphs 105 to 123, wherein the Cas is a Type IIIA csm protein or a Type IIIB cmr protein.
127. The use or method of any one of paragraphs 105 to 123, wherein the Cas is a Csf1.
128. The use or method of any one of paragraphs 105 to 123, wherein the Cas is a Cpf1.
129. The use or method of any one of paragraphs 105 to 123, wherein the Cas is a Cas13 (optionally Cas13a or Cas13b).
130. The use or method of any one of paragraphs 105 to 123, wherein the Cas is selected from a Cas3, Cas8a, Cas5, Cas8b, Cas8c, Cas10d, Cse1, Cse2, Csy1, Csy2, Csy3, GSU0054, Cas10, Csm2, Cmr5, Cas10, Csx11,

- Csx10, Csf1, Cas9, Csn2, Cas4, Cpf1, C2c1, C2c3, Cas13a, Cas13b and Cas13c.
131. The use or method of any one of paragraphs 105 to 130, wherein the DNA construct comprises one or more nucleotide sequences for producing crRNAs or gRNAs that are operable for Cas nuclease targeting.
132. The use or method of any one of paragraphs 105 to 131, wherein the production strain cells comprise a helper phage genome that is inducible to produce phage coat proteins in the cells, wherein the method further comprises inducing production of the phage proteins and causing packaging of the amplified DNA into phage particles or non-self-replicative transduction particles, and further isolating the phage or transduction particles and optionally formulating the particles into a pharmaceutical composition for administration to a human or animal subject for treating or reducing the risk of a disease or condition in the subject.
133. The method of paragraph 132, wherein the particles are capable of infecting target host cells in the subject and transducing the cells with the DNA, wherein the Cas and crRNAs (or gRNAs) encoded by the DNA are expressed in the cells, the crRNAs or (gRNAs) being operable to guide the Cas to a target nucleotide sequence (optionally a chromosomal sequence) comprised by the cells, wherein the Cas cuts the target sequences in the cells, thereby killing host cells and treating or reducing the risk of the disease or condition.
134. The method of paragraph 133, wherein the host cells are bacterial or archaeal cells, optionally, the host cells are *C difficile*, *P aeruginosa*, *K pneumoniae* (eg. carbapenem-resistant *Klebsiella pneumoniae* or Extended-Spectrum Beta-Lactamase (ESBL)-producing *K pneumoniae*), *E coli* (eg, ESBL-producing *E. coli*, or *E. coli* ST131-O25b:H4), *H pylori*, *S pneumoniae* or *S aureus* cells.
135. The use or method of any one of paragraphs 105 to 134, wherein each DNA is comprised by a high copy number vector, optionally a high copy number plasmid (an optionally the promoter is a constitutive promoter).
136. The use or method of any one of paragraphs 105 to 135, wherein each DNA is comprised by a vector according to any one of paragraphs 1 to 78 and 81 to 83.

Clauses

The invention provides, by way of example, the following Clauses; the features of these are combinable with any other disclosure herein.

1. A nucleic acid vector for introduction into a host cell, the vector comprising a first nucleotide sequence encoding a Type I Cas3 and a second nucleotide sequence encoding one or more Cascade proteins, wherein the first and second sequences are under the control of one or more promoters comprised by the vector for expression of the proteins in the cell.
2. The vector of Clause 1, wherein the vector comprises an operon for expression in the cell of the Cas3 and Cascade proteins from a Cas module, the module comprising the nucleotide sequences encoding the Cas3 and Cascade proteins, and the operon comprising the Cas module under the control of a promoter for controlling the expression of both the Cas3 and Cascade proteins.
3. The vector of Clause 2, wherein
 - (a) the first sequence is between the promoter and the second sequence in the operon;

- (b) the operon comprises no Cas-encoding nucleotide sequences between the promoter and the first nucleotide sequence; and/or
- (c) the operon comprises (in 5' to 3' direction) the promoter, the first sequence and the second sequence.
4. The vector of any preceding Clause, wherein each promoter is a constitutive promoter.
5. The vector of any one of Clauses 1 to 3, wherein the promoter is repressible (optionally repressible by a tetracycline repressor or lac repressor).
6. The vector of any one of Clauses 1 to 3, wherein the promoter is inducible.
7. The vector of any preceding Clause, wherein the first sequence is under the control of a medium strength promoter.
8. The vector of any preceding Clause, wherein the first sequence is under the control of a promoter that has an Anderson Score (AS) of $0.5 > AS > 0.1$.
9. The vector of any preceding Clause, wherein the first sequence (and optionally the second sequence) is under the control of a promoter and translation initiation site (TIS) combination that is capable of producing expression of green fluorescent protein (GFP) from a first expression operating unit (EOU) in *E. coli* strain BW25113 cells with a fluorescence of from 0.5 to 4 times the fluorescence produced in *E. coli* strain BW25113 cells using a second EOU comprising a P10 promoter (SEQ ID NO: 1) combined with a BCD14 TIS (SEQ ID NO: 2), wherein the EOUs differ only in their promoter and TIS combinations, wherein each EOU comprises (in 5' to 3' direction) an upstream initiator, the respective promoter, the respective TIS, a nucleotide sequence encoding GFP, a 3' UTR, a transcription terminator and a downstream insulator.
10. The vector of Clause 9, wherein fluorescence using the first EOU is 0.5 to 2 times the fluorescence using the second EOU.
11. The vector of any preceding Clause, wherein the vector comprises an origin of replication that is operable in the host cell.
12. The vector of any preceding Clause, wherein the vector comprises an origin of replication that is operable in a bacterial cell of a vector production strain, wherein the Cas3 is not operable in the production strain cell to target and cut a chromosomal sequence thereof.
13. The vector of Clause 12, wherein the first sequence is under the control of a promoter that is capable of controlling expression of the Cas3 at a level that is not toxic to the production strain cell.
14. The vector of any preceding Clause, wherein the vector is a high copy number vector.
15. The vector of any preceding Clause, wherein the first nucleotide sequence or operon is comprised by a mobile genetic element.
16. The vector of any preceding Clause, wherein the vector is devoid of a Cas adaption module.
17. The vector of any preceding Clause, wherein the vector is devoid of nucleotide sequence encoding one, more or all of a Cas1, Cas2, Cas4, Cas6, Cas7 and Cas8.
18. The vector of any preceding Clause, wherein the vector comprises (optionally in 5' to 3' direction) nucleotide sequence encoding one, more or all of Cas11, Cas7 and Cas8a1.

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19. The vector of Clause 18, wherein the vector comprises nucleotide sequence encoding Cas3' and/or Cas3".
20. The vector or Clause 19, wherein the nucleotide sequences encoding the Cas3' and/or Cas3" are between the promoter and the sequence(s) recited in Clause 18. 5
21. The vector of any one of Clauses 18 to 20, wherein the host cell comprises a Type IA CRISPR array that is cognate with the Cas3.
22. The vector of any one of Clauses 18 to 20, wherein the host cell comprises an endogenous Type IB, C, U, D, E or F CRISPR/Cas system. 10
23. The vector of any one of Clauses 1 to 17, wherein the vector comprises (optionally in 5' to 3' direction) nucleotide sequence encoding one, more or all of Cas8b1, Cas7 and Cas5. 15
24. The vector of Clause 23, wherein the vector comprises a nucleotide sequence encoding Cas3 between the promoter and the sequence(s) recited in Clause 23.
25. The vector of Clause 23 or 24, wherein the host cell comprises a Type IB CRISPR array that is cognate with the Cas3. 20
26. The vector of Clause 23 or 24, wherein the host cell comprises an endogenous Type IA, C, U, D, E or F CRISPR/Cas system. 25
27. The vector of any one of Clauses 1 to 17, wherein the vector comprises (optionally in 5' to 3' direction) nucleotide sequence encoding one, more or all of Cas5, Cas8c and Cas7.
28. The vector of Clause 27, wherein the vector comprises a nucleotide sequence encoding Cas3 between the promoter and the sequence(s) recited in Clause 27. 30
29. The vector of Clause 27 or 28, wherein the host cell comprises a Type IC CRISPR array that is cognate with the Cas3. 35
30. The vector of Clause 27 or 28, wherein the host cell comprises an endogenous Type IA, B, U, D, E or F CRISPR/Cas system.
31. The vector of any one of Clauses 1 to 17, wherein the vector comprises (optionally in 5' to 3' direction) nucleotide sequence encoding one, more or all of Cas8U2, Cas7, Cas5 and Cas6. 40
32. The vector of Clause 31, wherein the vector comprises a nucleotide sequence encoding Cas3 between the promoter and the sequence(s) recited in Clause 31. 45
33. The vector of Clause 31 or 32, wherein the host cell comprises a Type IU CRISPR array that is cognate with the Cas3.
34. The vector of Clause 31 or 32, wherein the host cell comprises an endogenous Type IA, B, C, D, E or F CRISPR/Cas system. 50
35. The vector of any one of Clauses 1 to 17, wherein the vector comprises (optionally in 5' to 3' direction) nucleotide sequence encoding one, more or all of Cas10d, Cas7 and Cas5. 55
36. The vector of Clause 35, wherein the vector comprises a nucleotide sequence encoding Cas3' and/or Cas3".
37. The vector of Clause 36, wherein the nucleotide sequences encoding the Cas3' and/or Cas3" are between the promoter and the sequence(s) recited in Clause 35. 60
38. The vector of any one of Clauses 35 to 37, wherein the host cell comprises a Type ID CRISPR array that is cognate with the Cas3.
39. The vector of any one of Clauses 35 to 37, wherein the host cell comprises an endogenous Type IA, B, C, U, E or F CRISPR/Cas system. 65

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40. The vector of any one of Clauses 1 to 17, wherein the vector comprises (optionally in 5' to 3' direction) nucleotide sequence encoding one, more or all of Cas8e, Cas11, Cas7, Cas5 and Cas6.
41. The vector of Clause 40, wherein the vector comprises a nucleotide sequence encoding Cas3 between the promoter and the sequence(s) recited in Clause 40.
42. The vector of Clause 40 or 41, wherein the host cell comprises a Type IE CRISPR array that is cognate with the Cas3.
43. The vector of Clause 40 or 41, wherein the host cell comprises an endogenous Type IA, B, C, D, U or F CRISPR/Cas system.
44. The vector of any one of Clauses 1 to 17, wherein the vector comprises (optionally in 5' to 3' direction) nucleotide sequence encoding one, more or all of Cas8f, Cas5, Cas7 and Cas6f.
45. The vector of Clause 44, wherein the vector comprises a nucleotide sequence encoding Cas3 between the promoter and the sequence(s) recited in Clause 44, wherein the vector is devoid of nucleotide sequence encoding further Cas between the promoter and the sequence encoding the Cas3.
46. The vector of Clause 44 or 45, wherein the host cell comprises a Type IF CRISPR array that is cognate with the Cas3.
47. The vector of Clause 44 or 45, wherein the host cell comprises an endogenous Type IA, B, C, D, U or E CRISPR/Cas system.
48. The vector of any one of Clauses 1 to 17, wherein the Cas and Cascade are
 - (a) Type IA Cas and Cascade proteins;
 - (b) Type IB Cas and Cascade proteins;
 - (c) Type IC Cas and Cascade proteins;
 - (d) Type ID Cas and Cascade proteins;
 - (e) Type IE Cas and Cascade proteins;
 - (f) Type IF Cas and Cascade proteins; or
 - (g) Type IU Cas and Cascade proteins.
49. The vector of any preceding Clause, wherein the Cas and Cascade are *E coli* (optionally Type IE or IF) Cas and Cascade proteins.
50. The vector of Clause 49, wherein the *E coli* is ESBL-producing *E. coli* or *E. coli* ST131-O25b:H4.
51. The vector of any preceding Clause, wherein the Cas and Cascade are
 - (a) *Clostridium* (eg, *C difficile*) Cas and Cascade proteins, optionally *C difficile* resistant to one or more antibiotics selected from aminoglycosides, lincomycin, tetracyclines, erythromycin, clindamycin, penicillins, cephalosporins and fluoroquinolones;
 - (b) *Pseudomonas aeruginosa* Cas and Cascade proteins, optionally *P aeruginosa* resistant to one or more antibiotics selected from carbapenems, aminoglycosides, cefepime, ceftazidime, fluoroquinolones, piperacillin and tazobactam; or
 - (c) *Klebsiella pneumoniae* (eg, carbapenem-resistant *Klebsiella pneumoniae* or Extended-Spectrum Beta-Lactamase (ESBL)-producing *K pneumoniae*) Cas and Cascade proteins.
52. The vector of any preceding Clause, wherein the Cas and Cascade are *E coli*, *C difficile*, *P aeruginosa*, *K pneumoniae*, *P furiosus* or *B halodurans* Cas and Cascade proteins.
53. The vector of any preceding Clause, wherein the Cas3 is a Cas3 of a CRISPR/Cas locus of a first bacterial or archaeal species, wherein the distance between the Cas3-encoding sequence of the locus and its cognate

promoter is further than the distance between the Cas3-encoding sequence and the respective promoter comprised by the vector.

54. The vector of any preceding Clause, wherein the distance between the promoter and the Cas3-encoding sequence and/or Cascade protein-encoding sequence(s) is shorter than in a corresponding wild-type Type I locus.
55. The vector of any preceding Clause, wherein the vector comprises (i) a CRISPR array for producing crRNAs in the host cell and/or (ii) one or more nucleotide sequences encoding one or more guide RNAs (gRNAs or single gRNAs), wherein the crRNAs or gRNAs are cognate to the Cas3 (and optionally cognate to the Cascade proteins).
56. The vector of Clause 55 when dependent from Clause 2, wherein the array or gRNA-encoding sequence(s) are comprised by the operon and under the control of the promoter.
57. The vector of Clause 56, wherein the array or gRNA-encoding sequence(s) are under the control of a promoter that is different from the promoter that controls the expression of the Cas3.
58. The vector of Clause 56 or 57, wherein one or more of the crRNAs or gRNAs comprises a spacer sequence that is capable of hybridising to a target nucleotide sequence of the host cell, wherein the target sequence is adjacent a PAM, the PAM being cognate to the Cas3.
59. The vector of Clause 58, wherein the target sequence is a chromosomal sequence of the host cell.
60. The vector of Clause 58 or 59, wherein the Cas3 is operable to cut the target sequence.
61. The vector of any preceding Clause, wherein the vector is a plasmid or phagemid.
62. A delivery vehicle comprising the vector of any preceding Clause, wherein the delivery vehicle is capable of delivering the vector into the host cell.
63. The vehicle of Clause 62, wherein the delivery vehicle is a phage, non-replicative transduction particle, nanoparticle carrier, bacterium or liposome.
64. The vector or vehicle of any preceding Clause, wherein the host cell is a bacterial or archaeal cell, optionally, the host cell is a *C difficile*, *P aeruginosa*, *K pneumoniae* (eg, carbapenem-resistant *Klebsiella pneumoniae* or Extended-Spectrum Beta-Lactamase (ESBL)-producing *K pneumoniae*), *E coli* (eg, ESBL-producing *E. coli*, or *E. coli* ST131-O25b:H4), *H pylori*, *S pneumoniae* or *S aureus* cell.
65. The vector or vehicle of any preceding Clause for administration to a human or animal subject for treating or reducing the risk of a disease or condition in the subject.
66. The vector or vehicle of Clause 65, wherein the disease or condition is an infection of the subject with host cells (eg, bacterial cells), or wherein the disease or condition is mediated by host cells (eg, bacterial cells).
67. A pharmaceutical composition comprising the vector or vehicle of any preceding Clause and a pharmaceutically acceptable diluent, excipient or carrier.

It will be understood that particular embodiments described herein are shown by way of illustration and not as limitations of the invention. The principal features of this invention can be employed in various embodiments without departing from the scope of the invention. Those skilled in the art will recognize, or be able to ascertain using no more than routine study, numerous equivalents to the specific procedures described herein. Such equivalents are consid-

ered to be within the scope of this invention and are covered by the claims. All publications and patent applications mentioned in the specification are indicative of the level of skill of those skilled in the art to which this invention pertains. All publications and patent applications and all US equivalent patent applications and patents are herein incorporated by reference to the same extent as if each individual publication or patent application was specifically and individually indicated to be incorporated by reference. Reference is made to WO2017/118598, US20180140698, US20170246221, US20180273940, US20160115488, US20180179547, US20170175142, US20160024510, US20150064138, US20170022499, US20160345578, US20180155729, US20180200342, WO2017112620, WO2018081502, PCT/EP2018/066954, PCT/EP2018/066980, PCT/EP2018/071454 and U.S. Ser. No. 15/985,658 and equivalent publications by the US Patent and Trademark Office (USPTO) or WIPO, the disclosures of which are incorporated herein by reference for providing disclosure that may be used in the present invention and/or to provide one or more features (eg, of a vector) that may be included in one or more claims herein.

The use of the word “a” or “an” when used in conjunction with the term “comprising” in the claims and/or the specification may mean “one,” but it is also consistent with the meaning of “one or more,” “at least one,” and “one or more than one.” The use of the term “or” in the claims is used to mean “and/or” unless explicitly indicated to refer to alternatives only or the alternatives are mutually exclusive, although the disclosure supports a definition that refers to only alternatives and “and/or.” Throughout this application, the term “about” is used to indicate that a value includes the inherent variation of error for the device, the method being employed to determine the value, or the variation that exists among the study subjects.

As used in this specification and claim(s), the words “comprising” (and any form of comprising, such as “comprise” and “comprises”), “having” (and any form of having, such as “have” and “has”), “including” (and any form of including, such as “includes” and “include”) or “containing” (and any form of containing, such as “contains” and “contain”) are inclusive or open-ended and do not exclude additional, unrecited elements or method steps.

The term “or combinations thereof” or similar as used herein refers to all permutations and combinations of the listed items preceding the term. For example, “A, B, C, or combinations thereof” is intended to include at least one of: A, B, C, AB, AC, BC, or ABC, and if order is important in a particular context, also BA, CA, CB, CBA, BCA, ACB, BAC, or CAB. Continuing with this example, expressly included are combinations that contain repeats of one or more item or term, such as BB, AAA, MB, BBC, AAABCCCC, CBBAAA, CABABB, and so forth. The skilled artisan will understand that typically there is no limit on the number of items or terms in any combination, unless otherwise apparent from the context.

Any part of this disclosure may be read in combination with any other part of the disclosure, unless otherwise apparent from the context.

All of the compositions and/or methods disclosed and claimed herein can be made and executed without undue experimentation in light of the present disclosure. While the compositions and methods of this invention have been described in terms of preferred embodiments, it will be apparent to those of skill in the art that variations may be applied to the compositions and/or methods and in the steps or in the sequence of steps of the method described herein

without departing from the concept, spirit and scope of the invention. All such similar substitutes and modifications apparent to those skilled in the art are deemed to be within the spirit, scope and concept of the invention as defined by the appended claims.

The present invention is described in more detail in the following non-limiting Examples.

EXAMPLES

The examples illustrate fast and precision killing of *Escherichia coli* strains. As a model programmable nuclease system, we used a CRISPR guided vector (CGVTM) to specifically target *Escherichia coli* MG1655.

Example 1

Single-Vector Cas3 & Cascade: Type I CRISPR-Cas System Targeting *E. Coli*

A plasmid (which we call a CRISPR Guided VectorTM, CGVTM) was constructed comprising an operon with nucleotide sequences encoding a Type I Cas3 and Cascade proteins under the control of a common promoter. *C. difficile* Type IB Cas3 and Cascade was used. A cognate CRISPR array comprising *C. difficile* repeat sequences and spacer sequence for targeting an *E. coli* host cell chromosome was also introduced into target cells. An adaptation module containing Cas1, Cas2 and Cas4 was omitted in the vector (see FIG. 1A). In the wild-type *C. difficile* Type IB CRISPR/Cas locus, the cas3 gene is 3' of the Cascade genes (cas8b1, cas7 and cas5) and thus spaced away from the promoter upstream of the Cascade genes. When we tried this arrangement, we found killing of *E. coli* cells, but surprisingly when we changed to a synthetic operon arrangement (in 5' to 3' orientation) of promoter, cas3, cas8b1, cas7 and cas5 we saw significantly higher killing of the target *E. coli* cells.

Results using this synthetic operon arrangement are shown in FIGS. 1A-1C. In FIG. 1B there is shown a dilution series (10¹-10⁶) of drop spots (5 μl) of target *E. coli* MG1655 cells harboring the CGV on LB agar plates with and without inducers. CRISPR/Cas induction surprisingly killed 99.9% of the population (FIG. 1C, grey bar). Growth in absence of induction is shown in black. CGVTM refers to a CRISPR Guided VectorTM, which is a nucleic acid vector comprising nucleotide sequences encoding CRISPR/Cas components.

We also managed to achieve desirable targeted killing of *E. coli* cells using a similar set-up, except that *E. coli* Type IE Cas and Cascade were used, together with a cognate array targeting host cell *E. coli* chromosomal DNA (data not shown). In this case, a vector was used comprising (in 5' to 3' direction) a promoter controlling the expression of Cas3, Cas8e, Cas 11, Cas7, Cas5 and Cas6 in an operon.

Materials and Methods

E. coli MG1655 was grown in lysogeny broth (LB) with shaking (250 rpm) at 37° C. When necessary, cultures were supplemented with tetracycline (10 μg/mL), and spectinomycin (400 μg/mL).

To construct a plasmid containing *C. difficile* CRISPR system under arabinose inducible pBAD promoter, cas3, cas6, cas8b, cas7 and cas5 genes from *C. difficile* were amplified and cloned in a low copy number plasmid (pSC101 ori). cas3 was located in the beginning of the operon followed by cas6, cas8b, cas7 and cas5. The adaptation module (consisting of cas1, cas2, and cas4) was omitted in the vector (FIG. 1A). A second plasmid containing an IPTG inducible single-spacer array targeting a chro-

mosomal intergenic region in *E. coli* MG1655 was constructed (FIG. 1A). The spacer was cloned under control of the IPTG-inducible P_{trc} promoter, in a CloDF13 ori backbone. It contains 37 nucleotides from the genome of *E. coli* MG1655 (cttgccgcgcttcgtcacgtaattctcgtcgcaa) (SEQ ID NO: 26). Additionally, the 3'-CCT protospacer adjacent motif (PAM) is located adjacent to the selected target sequence in the genome of *E. coli* MG1655 (FIG. 1A).

To perform killing assays, both plasmids were transformed into *E. coli* MG1655 by electroporation. Transformants were grown in liquid LB with antibiotics to mid-log phase, and the killing efficiency was determined by serial dilution and spot plating onto LB, and LB+inducers (0.5 mM IPTG and 1% arabinose). Viability was calculated by counting colony forming units (CFUs) on the plates and data were calculated as viable cell concentration (CFU/ml).

Example 2

Single-Vector Cas3-Cascade & Array: Type I CRISPR-Cas System Targeting *E. Coli*

A plasmid (which we call a CRISPR Guided VectorTM, CGVTM, which is a nucleic acid vector comprising nucleotide sequences encoding CRISPR/Cas components) was constructed comprising an operon with nucleotide sequences encoding a Type I Cas3 and Cascade proteins under the control of a common promoter. *C. difficile* Type IB Cas3 and Cascade was used. Adaptation module containing Cas1, Cas2 and Cas4 was omitted in the vector. A cognate CRISPR array comprising *C. difficile* repeat sequences and spacer sequence for targeting an *E. coli* host cell chromosome was also cloned in the vector (see FIG. 2A). Similarly we also constructed a plasmid comprising of an operon with nucleotide sequences encoding *E. coli* Type IE Cas3 and Cascade proteins under control of a common promoter. The *E. coli* adaption module containing Cas1 and Cas2 was omitted, in the vector. A cognate CRISPR array comprising *E. coli* repeat sequences and spacer sequence for targeting an *E. coli* host cell chromosome was also cloned in the vector.

The CGV containing the *C. difficile* CRISPR-Cas system was transformed into *E. coli* MG1655 which contains a pks sequence incorporated into the genome. Results using this synthetic operon arrangement are shown in FIGS. 2A-2C. In FIG. 2B there is shown a dilution series (10¹-10⁵) of drop spots (5 μl) of target *E. coli* MG1655 cells harboring the CGV on synthetic medium (SM) agar plates with and without inducers. CRISPR/Cas induction resulted in more than 2-log₁₀ reductions in viable cells of *E. coli* MG1655 (FIG. 2C, grey bar). Growth in absence of induction is shown in black. CGVTM refers to a CRISPR Guided VectorTM.

The survival of *E. coli* MG1655 upon induction was followed over time by plating the cultures in serial dilutions every 60 minutes, for 2 h (FIG. 3A) Killing curves revealed that CRISPR/Cas induction mediated rapid killing of *E. coli* MG1655, generating a two-log₁₀ reduction in *E. coli* by the first 60 minutes. FIG. 3B shows a dilution series (10¹-10⁶) of drop spots (5 μl) of induced and non-induced cultures of target *E. coli* MG1655 on SM agar plates.

The CGV containing the *E. coli* CRISPR-Cas system was transformed into other *E. coli* MG1655 cells which contain a lambda sequence incorporated into the genome. Results using this synthetic operon arrangement are shown in FIGS. 6A-6B. In FIG. 6A there is shown a dilution series (10¹-10⁵) of drop spots (5 μl) of target *E. coli* MG1655 cells harboring the CGV on synthetic medium (SM) agar plates with and

without inducers. CRISPR/Cas induction resulted in more than 2-log₁₀ reductions in viable cells of *E. coli* MG1655 (FIG. 6B, grey bar). Growth in absence of induction is shown in black. In a repeat experiment (not shown) we saw a 3-log₁₀ reductions in viable cells of *E. coli* MG1655 with CRISPR/Cas induction.

Materials and Methods

E. coli MG1655 was grown in synthetic medium (SM) with shaking (250 rpm) at 37° C. Cultures were supplemented with 10 µg/mL tetracycline when required.

To construct a plasmid containing *C. difficile* CRISPR system under arabinose inducible pBAD promoter, cas3, cas6, cas8b, cas7 and cas5 genes from *C. difficile* were amplified and cloned in a low copy number plasmid (pSC101 ori). cas3 was located in the beginning of the operon followed by cas6, cas8b, cas7 and cas5. Additionally, an IPTG inducible single-spacer array targeting a chromosomal intergenic region in *E. coli* MG1655 was included in the vector under control of the IPTG-inducible P_{trc} promoter (FIG. 2A). It contains 37 nucleotides from the PKS gene (previously integrated into the genome of *E. coli* MG1655) (gtttggcgatggcgggtgtggtgtgcttcggcgt) (SEQ ID NO: 27). Additionally, the 3'-CCT protospacer adjacent motif (PAM) is located adjacent to the selected target sequence in the genome of *E. coli* MG1655 (FIG. 2A).

To construct a plasmid containing *E. coli* CRISPR system under arabinose inducible pBAD promoter, cas3, cse1, cse2, cas7, cas5 and cas6 genes from *E. coli* were amplified and cloned in a low copy number plasmid (pSC101 ori). The operon comprised (in 5' to 3' direction) cas3 followed by cse1 cse2, cas7, cas5 and cas6. Additionally, an IPTG inducible single-spacer array targeting a chromosomal intergenic region in *E. coli* MG1655 was included in the vector under control of the IPTG-inducible P_{trc} promoter. It contained 32 nucleotides from the lambda sequence (previously integrated into the genome of *E. coli* MG1655) (tgggatgcc-taccgcaagcagctggcctgaa) (SEQ ID NO: 28) and found to efficiently target in Brouns et al., 2008 (Science. 2008 Aug. 15; 321(5891):960-4. doi: 10.1126/science.1159689; "Small CRISPR RNAs guide antiviral defense in prokaryotes"). Additionally, the 3'-ATG protospacer adjacent motif (PAM) is located adjacent to the selected target sequence in the genome of *E. coli* MG1655.

The CGVs were transformed into *E. coli* MG1655 by electroporation. Transformants were grown in liquid SM with antibiotics to mid-log phase, and the killing efficiency was determined by serial dilution and spot plating onto LB, and LB+inducers (0.5 mM IPTG and 1% arabinose). Viability was calculated by counting colony forming units (CFUs) on the plates and data were calculated as viable cell concentration (CFU/ml).

To perform killing curves, *E. coli* MG1655 harboring the CGV was grown in liquid SM with antibiotics to mid-log phase. The culture was divided into two tubes and either inducers (0.5 mM IPTG and 1% arabinose) or PBS were added. Survival of the strain was followed over time by plating the cultures in serial dilutions (10¹-10⁶) of drop spots (5 µl) every 60 minutes, for 2 h, on SM plates with antibiotics. Survival frequency was calculated by counting colony forming units (CFUs) on the plates and data were calculated as viable cell concentration (CFU/ml).

Example 3

Precision Killing of Target Strain *E. Coli* MG1655 in a Microbiome

An artificial microbial consortium was constructed to study the efficiency of the CGV carrying the CRISPR-Cas

system of *C. difficile*, to specifically target *E. coli* MG1655 in the presence of other microbes, mimicking the human microbiome.

The synthetic consortium consisted of three strains (two different species) with differential antibiotic resistance profiles: a streptomycin-resistant *E. coli* MG1655 (target strain), an ampicillin-resistant *E. coli* Top10, and a chloramphenicol-resistant *Lactococcus lactis* NZ9000. To create the consortium, bacterial cultures were grown separately in Brain Heart Infusion broth (BHI, optimal growth medium for *L. lactis*) to mid-log phase and mixed in fresh BHI broth with and without inducers. After 1 h induction at 30° C., the composition of the consortium was determined by counting viable colonies on selective plates. Induction of the CRISPR system in the mixed community, resulted in >10-fold killing of target *E. coli* MG1655, while leaving *E. coli* Top10 and *L. lactis* NZ9000 cell populations unharmed (FIG. 4A). In FIG. 4B there is shown a dilution series (10¹-10⁵) of drop spots (5 µl) of the synthetic consortium after 1 h induction on BHI agar plates.

Additionally, CRISPR killing of target strain *E. coli* MG1655 in the synthetic microbial consortium was compared to a pure culture (ie, target strain *E. coli* MG1655 that is not mixed with another strain or species). Unexpectedly, in both conditions, killing of 3 logs was achieved when plated on BHI agar plates with inducers (FIG. 5A). Thus, surprisingly the killing in the microbiome setting was as efficient as the killing in pure culture. In FIG. 5B there is shown a dilution series (10¹-10⁵) of drop spots (5 µl) of the synthetic consortium and *E. coli* MG1655 in pure culture on BHI agar plates with and without inducers.

Materials and Methods

E. coli MG1655, *E. coli* Top10, and *Lactococcus lactis* NZ9000 were grown in BHI broth with shaking (250 rpm) at 30° C. Cultures were supplemented with 1000 µg/mL streptomycin, 100 µg/mL ampicillin, or 10 µg/mL chloramphenicol, respectively.

To create the consortium, bacterial cultures were grown in BHI with appropriate antibiotics to mid-log phase. Cultures were washed twice in PBS to remove the antibiotics and mixed in fresh BHI broth. The mixed culture was spotted onto BHI plates with streptomycin, ampicillin or chloramphenicol to quantify the initial concentration of *E. coli* MG1655, *E. coli* Top10 and *L. lactis* NZ9000, respectively. The mixed culture was divided into two tubes and either inducers (0.5 mM IPTG and 1% arabinose) or PBS were added. After 1 h induction at 30° C., the composition of the consortium was calculated by counting colony forming units (CFUs) on selective plates and data were calculated as viable cell concentration (CFU/ml).

Example 4

Use of Promoter Repression in Vector Amplification Strains

We engineered an *E. coli* Top10 production strain cell population comprising plasmid CGV DNA and an expressible sequence encoding a Tet repressor (TetR). The DNA comprised a Cas9-encoding nucleotide sequence under the control of a Tet promoter (pLtetO-1 promoter). The promoter is normally constitutively ON, but it was repressed by TetR in our cells. Thus, in this way we could successfully culture the cells and amplify the CGV without observing adverse toxicity due to Cas9 expression.

In an experiment in the absence of repression, we did not observe any colonies of production strain bacteria, and we

surmise that this was due to Cas9 toxicity. We believe, in addition to providing a way of increasing CGV yield (eg, for subsequent packaging into phage or non-self-replicative transduction particles), our method using repression can minimize selection for mutations in the DNA that would otherwise be forced by higher Cas9 expression and cutting (eg, due to CGV cutting).

REFERENCES

Mutalik et al, Nat Methods. 2013 April; 10(4):354-60. doi: 10.1038/nmeth. 2404. Epub 2013 Mar. 10, "Precise and reliable gene expression via standard transcription and translation initiation elements".

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TABLE 1

Example Bacteria		
Optionally, the target host cells are cells of a genus or species selected from this Table and/or the production strain cells are cells of a genus or species selected from this Table	Optionally, the target host cells are cells of a genus or species selected from this Table and/or the production strain cells are cells of a genus or species selected from this Table	Optionally, the target host cells are cells of a genus or species selected from this Table and/or the production strain cells are cells of a genus or species selected from this Table
<i>Abiotrophia</i>	<i>Acidocella</i>	<i>Alkallimnicola</i>
<i>Abiotrophia defectiva</i>	<i>Acidocella aminolytica</i>	<i>Alkallimnicola ehrlichii</i>
<i>Acaricomes</i>	<i>Acidocella facilis</i>	<i>Alkaliphilus</i>
<i>Acarticomes phytoseutili</i>	<i>Acidomonas</i>	<i>Alkaliphilus orenlandii</i>
<i>Acetitomaculum</i>	<i>Acidomonas methanolica</i>	<i>Alkaliphilus transvaalensis</i>
<i>Acetitomaculum ruminis</i>	<i>Acidothermus</i>	<i>Allochromatium</i>
<i>Acetivibrio</i>	<i>Acidothermus cellulolyticus</i>	<i>Allochromatium vinosum</i>
<i>Acetivibrio cellulolyticus</i>	<i>Acidovorax</i>	<i>Alloiooccus</i>
<i>Acetivibrio ethanoligenens</i>	<i>Acidovorax anthurii</i>	<i>Alloiooccus otitis</i>
<i>Acetivibrio multivorans</i>	<i>Acidovorax caeni</i>	<i>Allokatzeria</i>
<i>Acetoanaerobium</i>	<i>Acidovorax catleyae</i>	<i>Allokatzeria alba</i>
<i>Acetoanaerobium moterae</i>	<i>Acidovorax citrulli</i>	<i>Altererythrobacter</i>
<i>Acetobacter</i>	<i>Acidovorax defluvi</i>	<i>Altererythrobacter ishigakiensis</i>
<i>Acetobacter acetii</i>	<i>Acidovorax delafeldii</i>	<i>Altermonas</i>
<i>Acetobacter cerevisiae</i>	<i>Acidovorax facilis</i>	<i>Altermonas haloplanktis</i>
<i>Acetobacter cibinongensis</i>	<i>Acidovorax konjaci</i>	<i>Altermonas nucleodii</i>
<i>Acetobacter estunensis</i>	<i>Acidovorax temperans</i>	<i>Alysiella</i>
<i>Acetobacter fabarum</i>	<i>Acidovorax valerianellae</i>	<i>Alysiella crassa</i>
<i>Acetobacter ghanensis</i>	<i>Acinetobacter</i>	<i>Alysiella filiformis</i>
<i>Acetobacter indonesiensis</i>	<i>Acinetobacter baumannii</i>	<i>Aminobacter</i>
<i>Acetobacter lovanensis</i>	<i>Acinetobacter baylyi</i>	<i>Aminobacter aganoensis</i>
<i>Acetobacter malorum</i>	<i>Acinetobacter boveyi</i>	<i>Aminobacter aminovorans</i>
<i>Acetobacter nitroreducingens</i>	<i>Acinetobacter calcoaceticus</i>	<i>Aminobacter niigataensis</i>
<i>Acetobacter oeni</i>	<i>Acinetobacter generi</i>	<i>Aminobacterium</i>
<i>Acetobacter orientalis</i>	<i>Acinetobacter haemolyticus</i>	<i>Aminobacterium mobile</i>
<i>Acetobacter orleanensis</i>	<i>Acinetobacter johnsonii</i>	<i>Aminomonas</i>
<i>Acetobacter pasteurianus</i>	<i>Acinetobacter junii</i>	<i>Aminomonas paucivorans</i>
<i>Acetobacter pomorum</i>	<i>Acinetobacter lawfi</i>	<i>Ammophilus</i>
<i>Acetobacter senegalensis</i>	<i>Acinetobacter narvus</i>	<i>Ammophilus ovalaticus</i>
<i>Acetobacter xylinus</i>	<i>Acinetobacter radioreducens</i>	<i>Ammophilus oxalivorans</i>
<i>Acetobacterium</i>	<i>Acinetobacter schindleri</i>	<i>Amphibacillus</i>
<i>Acetobacterium bakii</i>	<i>Acinetobacter tandoi</i>	<i>Amphibacillus xylanus</i>
<i>Acetobacterium carbinolicum</i>	<i>Acinetobacter tjernbergiae</i>	<i>Amphritea</i>
<i>Acetobacterium delhalogenans</i>	<i>Acinetobacter townieri</i>	<i>Amphritea balenae</i>
<i>Acetobacterium fimetarium</i>	<i>Acinetobacter ursingii</i>	<i>Amphritea japonica</i>
<i>Acetobacterium malicum</i>	<i>Acinetobacter venetianus</i>	<i>Amycolatopsis</i>
<i>Acetobacterium pallidum</i>	<i>Acrocarpospora</i>	<i>Amycolatopsis alba</i>
<i>Acetobacterium landrae</i>	<i>Acrocarpospora corrugata</i>	<i>Amycolatopsis albidoflavus</i>
<i>Acetobacterium wieringae</i>	<i>Acrocarpospora</i>	<i>Amycolatopsis azurea</i>
<i>Acetofilamentum</i>	<i>Acrocarpospora macrocephala</i>	<i>Amycolatopsis coloradensis</i>
<i>Acetofilamentum rigidum</i>	<i>Acrocarpospora plectomorpha</i>	<i>Amycolatopsis lurida</i>
<i>Acetohalobium</i>	<i>Actibacter</i>	<i>Amycolatopsis mediterranei</i>
<i>Acetohalobium arabaticum</i>	<i>Actibacter sedimitis</i>	<i>Amycolatopsis rifamycinica</i>
<i>Acetomicrobium</i>	<i>Actinoallotrichus</i>	<i>Amycolatopsis rubida</i>
<i>Acetomicrobium faecale</i>	<i>Actinoallotrichus cyanogriseus</i>	<i>Amycolatopsis sulphurea</i>
<i>Acetomicrobium flavidum</i>	<i>Actinoallotrichus</i>	<i>Amycolatopsis tolypomycina</i>
<i>Acetomena</i>		<i>Anabaena</i>
		<i>Anabaena cylindrica</i>
		<i>Arthrospira</i>
		<i>Arthrospira carolinolicum</i>
		<i>Arthrospira delhalogenans</i>
		<i>Arthrospira fimetarium</i>
		<i>Arthrospira malicum</i>
		<i>Arthrospira pallidum</i>
		<i>Arthrospira landrae</i>
		<i>Arthrospira wieringae</i>
		<i>Arthrospira woodii</i>
		<i>Arthrospira rigidum</i>
		<i>Arthrospira arabaticum</i>
		<i>Arthrospira faecale</i>
		<i>Arthrospira flavidum</i>
		<i>Arthrospira protophormiae</i>
		<i>Aquaspirillum</i>
		<i>Aquaspirillum polymorphum</i>
		<i>Aquaspirillum putridiconchylum</i>
		<i>Aquaspirillum serpens</i>
		<i>Aquinarina</i>
		<i>Aquinarina latercula</i>
		<i>Arcanobacterium</i>
		<i>Arcanobacterium</i>
		<i>haemolyticum</i>
		<i>Arcanobacterium pyogenes</i>
		<i>Archangium</i>
		<i>Archangium gephyra</i>
		<i>Arcobacter</i>
		<i>Arcobacter butzleri</i>
		<i>Arcobacter cryaerophilus</i>
		<i>Arcobacter halophilus</i>
		<i>Arcobacter nitrofigilis</i>
		<i>Arcobacter skirrowii</i>
		<i>Arhodomonas</i>
		<i>Arhodomonas aquaeolei</i>
		<i>Arsenophonus</i>
		<i>Arsenophonus nasoniae</i>
		<i>Arthrospira</i>
		<i>Arthrospira agilis</i>
		<i>Arthrospira albus</i>
		<i>Arthrospira aurescens</i>
		<i>Arthrospira chlorophenolicus</i>
		<i>Arthrospira citreus</i>
		<i>Arthrospira crystallopoietes</i>
		<i>Arthrospira cumminsii</i>
		<i>Arthrospira globiformis</i>
		<i>Arthrospira</i>
		<i>hispidinolorans</i>
		<i>Arthrospira ilicis</i>
		<i>Arthrospira luteus</i>
		<i>Arthrospira methylotrophus</i>
		<i>Arthrospira nicotianae</i>
		<i>Arthrospira nicotianovans</i>
		<i>Arthrospira oxydans</i>
		<i>Arthrospira paszensis</i>
		<i>Arthrospira phenanthrenivorans</i>
		<i>Arthrospira</i>
		<i>polychromogenes</i>
		<i>Arthrospira</i>
		<i>protophormiae</i>

TABLE 1-continued

Optionally, the target host cells are cells of a genus or species selected from this Table		Example Bacteria	
<i>Acetonea longum</i>	<i>hymeniacidomus</i>	<i>Actinotalea</i>	<i>Anabaena flos-aquae</i>
<i>Acetothermus</i>	<i>Actinoallotetichus spitiensis</i>	<i>Actinotalea fermentans</i>	<i>Anabaena variabilis</i>
<i>Acetothermus paucivorans</i>	<i>Actinobacillus</i>	<i>Aerococcus</i>	<i>Anaerococcus</i>
<i>Acholeplasma</i>	<i>Actinobacillus capsulatus</i>	<i>Aerococcus sanguinicola</i>	<i>Anaerococcus burkinensis</i>
<i>Acholeplasma axanthum</i>	<i>Actinobacillus delphinicola</i>	<i>Aerococcus urinae</i>	<i>Anaerobaculum</i>
<i>Acholeplasma brassicae</i>	<i>Actinobacillus hominis</i>	<i>Aerococcus urinae-equi</i>	<i>Anaerobaculum mobile</i>
<i>Acholeplasma cavigentitulum</i>	<i>Actinobacillus indolicus</i>	<i>Aerococcus urinaehominis</i>	<i>Anaerobiospirillum</i>
<i>Acholeplasma equifetale</i>	<i>Actinobacillus ligidensis</i>	<i>Aerococcus viridans</i>	<i>Anaerobiospirillum</i>
<i>Acholeplasma granularum</i>	<i>Actinobacillus minor</i>	<i>Aeromicrobium</i>	<i>succiniciproducens</i>
<i>Acholeplasma hippikon</i>	<i>Actinobacillus muris</i>	<i>Aeromicrobium erythreum</i>	<i>Anaerobiospirillum thomastii</i>
<i>Acholeplasma laidlawii</i>	<i>Actinobacillus pleuropneumoniae</i>	<i>Aeromonas</i>	<i>Asaita bogorensis</i>
<i>Acholeplasma modicum</i>	<i>Actinobacillus porcini</i>	<i>allosaccharophila</i>	<i>Asanoa</i>
<i>Acholeplasma morum</i>	<i>Actinobacillus rossii</i>	<i>Aeromonas bestiarum</i>	<i>Asanoa ferruginea</i>
<i>Acholeplasma multilocale</i>	<i>Actinobacillus scotiae</i>	<i>Aeromonas caviae</i>	<i>Asiteccaulis</i>
<i>Acholeplasma oculi</i>	<i>Actinobacillus seminis</i>	<i>Aeromonas encheleia</i>	<i>Asiteccaulis biproshectum</i>
<i>Acholeplasma palmae</i>	<i>Actinobacillus succinogenes</i>	<i>Aeromonas entropelogenes</i>	<i>Asiteccaulis excentricus</i>
<i>Acholeplasma parvum</i>	<i>Actinobacillus ureae</i>	<i>Aeromonas eurenophila</i>	<i>Atropobacter</i>
<i>Acholeplasma plectiae</i>	<i>Actinobaculum</i>	<i>Aeromonas ichtiosmia</i>	<i>Atropobacter phocae</i>
<i>Acholeplasma vituli</i>	<i>Actinobaculum massiliense</i>	<i>Aeromonas jandaei</i>	<i>Atropobium</i>
<i>Achromobacter denitrificans</i>	<i>Actinobaculum schaalii</i>	<i>Aeromonas media</i>	<i>Atropobium fossor</i>
<i>Achromobacter insolitus</i>	<i>Actinobaculum rossii</i>	<i>Aeromonas popoffii</i>	<i>Atropobium minutum</i>
<i>Achromobacter piechandii</i>	<i>Actinomyces urinale</i>	<i>Aeromonas sobria</i>	<i>Atropobium parvulum</i>
<i>Achromobacter rublandii</i>	<i>Actinocatenispora</i>	<i>Aeromonas veronii</i>	<i>Atropobium rimae</i>
<i>Achromobacter spanius</i>	<i>Actinocatenispora rupis</i>	<i>Agrobacterium</i>	<i>Atropobium vaginae</i>
<i>Acidaminobacter</i>	<i>Actinocatenispora thailandica</i>	<i>gelatinovorum</i>	<i>Aureobacterium</i>
<i>Acidaminobacter</i>	<i>Actinocatenispora sera</i>	<i>Agrococcus citreus</i>	<i>Aureobacterium barkeri</i>
<i>hydrogeniformans</i>	<i>Actinocorallia</i>	<i>Agrococcus jenensis</i>	<i>Aureobacterium</i>
<i>Acidaminococcus fermentans</i>	<i>Actinocorallia aurantiaca</i>	<i>Agromonas</i>	<i>Aureobacterium liquefaciens</i>
<i>Acidicoccus</i>	<i>Actinocorallia aurea</i>	<i>Agromonas oligotrophica</i>	<i>Avibacterium</i>
<i>Acidicoccus organivorans</i>	<i>Actinocorallia cavernae</i>	<i>Agromyces</i>	<i>Avibacterium avium</i>
<i>Acidimicrobium</i>	<i>Actinocorallia glomerata</i>	<i>Agromyces fucosus</i>	<i>Avibacterium gallinarum</i>
<i>Acidimicrobium ferrooxidans</i>	<i>Actinocorallia herbida</i>	<i>Agromyces hippuratus</i>	<i>Avibacterium paragallinarum</i>
<i>Acidiphilium</i>	<i>Actinocorallia libanotica</i>	<i>Agromyces lateolus</i>	<i>Avibacterium volantium</i>
<i>Acidiphilium acidophilum</i>	<i>Actinocorallia longicatena</i>	<i>Agromyces mediolanus</i>	<i>Azoarcus</i>
<i>Acidiphilium angustum</i>	<i>Actinomadura</i>	<i>Agromyces ramosus</i>	<i>Azoarcus indigenus</i>
<i>Acidiphilium cryptum</i>	<i>Actinomadura atramentaria</i>	<i>Akkermansia</i>	<i>Azoarcus toluyticus</i>
<i>Acidiphilium multivorum</i>	<i>Actinomadura bangladeshensis</i>	<i>Albidiferax</i>	<i>Azoarcus toluvorans</i>
<i>Acidiphilium organovorum</i>	<i>Actinomadura catellatispora</i>	<i>Albidiferax ferritreducens</i>	<i>Azohydromonas</i>
<i>Acidiphilium rubrum</i>	<i>Actinomadura chibensis</i>	<i>Albidovulum</i>	<i>Azohydromonas australica</i>
<i>Acidisoma</i>	<i>Actinomadura chokotskensis</i>	<i>Albidovulum inexpectatum</i>	<i>Azohydromonas lata</i>
<i>Acidisoma sibiricum</i>	<i>Actinomadura citrea</i>	<i>Alcaligenes</i>	<i>Azomonas</i>
<i>Acidisoma tundrae</i>	<i>Actinomadura coerulea</i>	<i>Alcaligenes denitrificans</i>	<i>Azomonas agilis</i>
<i>Acidisphaera</i>	<i>Actinomadura echinospora</i>	<i>Alcaligenes faecalis</i>	<i>Azomonas insignis</i>
<i>Acidithiobacillus</i>	<i>Actinomadura fibrosa</i>	<i>Alcantivorax</i>	<i>Azomonas macrocytogenes</i>
<i>Acidithiobacillus albertensis</i>	<i>Actinomadura formosensis</i>		<i>Azorbaculum</i>
<i>Acidithiobacillus caldus</i>			<i>Azorbaculum caulinodans</i>
			<i>Azorbaculum pushtichinoensis</i>
			<i>Azorbaculum</i>
			<i>Azorbaculum paspali</i>
			<i>Azospirillum</i>
			<i>Azospirillum brasilense</i>

TABLE 1—continued

Optionally, the target host cells are cells of a genus or species selected from this Table and/or the production strain cells are cells of a genus or species selected from this Table	
Example Bacteria	
<i>Acidithiobacillus ferrooxidans</i>	<i>Alcamivorax borkuensis</i>
<i>Acidithiobacillus thiooxidans</i>	<i>Alcamivorax jadenis</i>
<i>Acidobacterium</i>	<i>Algicola</i>
<i>Acidobacterium capsulatum</i>	<i>Algicola bacteriolytica</i>
	<i>Alicyclobacillus</i>
	<i>Alicyclobacillus</i>
	<i>disulfidooxidans</i>
	<i>Alicyclobacillus vulcanalis</i>
	<i>sendaiensis</i>
	<i>Alishewanella</i>
	<i>Alishewanella fetalis</i>
	<i>Alkalibacillus</i>
	<i>haloalkaliphilus</i>
	<i>Bibersteinia</i>
	<i>Bibersteinia trehalosi</i>
	<i>Bifidobacterium</i>
	<i>Bifidobacterium adolescentis</i>
	<i>Bifidobacterium angulatum</i>
	<i>Bifidobacterium animalis</i>
	<i>Bifidobacterium asteroides</i>
	<i>Bifidobacterium bifidum</i>
	<i>Bifidobacterium boum</i>
	<i>Bifidobacterium breve</i>
	<i>Bifidobacterium catenulatum</i>
	<i>Bifidobacterium choerinum</i>
	<i>Bifidobacterium coryneforme</i>
	<i>Bifidobacterium cuniculi</i>
	<i>Bifidobacterium dentium</i>
	<i>Bifidobacterium gallinarum</i>
	<i>Bifidobacterium indicum</i>
	<i>Bifidobacterium longum</i>
	<i>Bifidobacterium</i>
	<i>magnumBifidobacterium</i>
	<i>merycicum</i>
	<i>Bifidobacterium minimum</i>
	<i>Bifidobacterium</i>
	<i>pseudocatenulatum</i>
	<i>Bifidobacterium</i>
	<i>pseudolongum</i>
	<i>Bifidobacterium pullorum</i>
	<i>Bifidobacterium ruminantium</i>
	<i>Bifidobacterium saeculare</i>
	<i>Bifidobacterium subtile</i>
<i>Bacillus</i>	<i>Borrelia</i>
[see below]	<i>Borrelia afzelii</i>
	<i>Borrelia americana</i>
	<i>Borrelia burgdorferi</i>
	<i>Borrelia carolinensis</i>
	<i>Borrelia coriaceae</i>
	<i>Borrelia garinii</i>
	<i>Borrelia japonica</i>
	<i>Bossea</i>
	<i>Bossea minatitlanensis</i>
	<i>Bossea thiooxidans</i>
	<i>Brachyobacterium</i>
	<i>alimentarium</i>
	<i>Brachyobacterium faecium</i>
	<i>Brachyobacterium</i>
	<i>paraconglomeratum</i>
	<i>Brachyobacterium rhammosum</i>
	<i>Brachyobacterium</i>
	<i>tyrofermentans</i>
	<i>Brachyspira</i>
	<i>Brachyspira alvinipulli</i>
	<i>Brachyspira hyodysenteriae</i>
	<i>Brachyspira imocens</i>
	<i>Brachyspira murdochii</i>
	<i>Brachyspira pilosicoli</i>
	<i>Bradyrhizobium</i>
	<i>Bradyrhizobium covenenans</i>
	<i>Bradyrhizobium canariense</i>
	<i>Bradyrhizobium elkanii</i>
	<i>Bradyrhizobium japonicum</i>
	<i>Brevinema</i>
	<i>Brevinema andersonii</i>
	<i>Brevundimonas</i>
	<i>Brevundimonas alba</i>
	<i>Brevundimonas aurantiaca</i>
	<i>Brevundimonas diminuta</i>
	<i>Brevundimonas intermedia</i>
	<i>Brevundimonas subvibrioides</i>
	<i>Brevundimonas vancouverii</i>
	<i>Brevundimonas variabilis</i>
	<i>Brevundimonas vesiculata</i>
	<i>Brochothrix</i>
	<i>Brochothrix campestris</i>
	<i>Brochothrix thermosphacta</i>
	<i>Brucella</i>
	<i>Brucella canis</i>
	<i>Brucella neotomae</i>
	<i>Bryobacter</i>
	<i>Bryobacter aggregatus</i>
	<i>Burkholderia</i>
	<i>Burkholderia ambifaria</i>
	<i>Burkholderia andropogonis</i>
	<i>Burkholderia anthina</i>
	<i>Burkholderia caledonica</i>
	<i>Burkholderia caryophylli</i>
	<i>Burkholderia cenocepacia</i>
	<i>Burkholderia cepacia</i>
	<i>Burkholderia cocovenenans</i>
	<i>Burkholderia dolosa</i>
	<i>Burkholderia fungorum</i>
	<i>Burkholderia glathei</i>

TABLE 1-continued

Example Bacteria		Optionally, the target host cells are cells of a genus or species selected from this Table and/or the production strain cells are cells of a genus or species selected from this Table	
<i>Bartonella elizabethae</i>	<i>Bifidobacterium</i>	<i>Bradyrhizobium liaoningense</i>	<i>Burkholderia glumae</i>
<i>Bartonella grahamii</i>	<i>thermophilum</i>	<i>Brenneria</i>	<i>Burkholderia graminis</i>
<i>Bartonella henselae</i>	<i>Bilophila</i>	<i>Brenneria albi</i>	<i>Burkholderia kururiensis</i>
<i>Bartonella rochalimae</i>	<i>Bilophila wadsworthia</i>	<i>Brenneria nigrifluens</i>	<i>Burkholderia multivorans</i>
<i>Bartonella vinsonii</i>	<i>Biostraticola</i>	<i>Brenneria quercina</i>	<i>Burkholderia phenazinium</i>
<i>Bovaricoccus</i>	<i>Biostraticola tofi</i>	<i>Brenneria quercina</i>	<i>Burkholderia plantarii</i>
<i>Bavaricoccus seileri</i>	<i>Bizionia</i>	<i>Brenneria salicis</i>	<i>Burkholderia pyrocinia</i>
<i>Bdellovibrio</i>	<i>Bizionia argentinensis</i>	<i>Brevibacillus</i>	<i>Burkholderia thailandensis</i>
<i>Bdellovibrio bacteriovorus</i>	<i>Blastobacter capsulatus</i>	<i>Brevibacillus agri</i>	<i>Burkholderia stabilis</i>
<i>Beggiatoa</i>	<i>Blastobacter denitrificans</i>	<i>Brevibacillus borstelensis</i>	<i>Burkholderia thailandensis</i>
<i>Beggiatoa alba</i>	<i>Blastococcus</i>	<i>Brevibacillus brevis</i>	<i>Burkholderia tropica</i>
<i>Beijerinckia</i>	<i>Blastococcus aggregatus</i>	<i>Brevibacillus centrosporus</i>	<i>Burkholderia unamae</i>
<i>Beijerinckia derxii</i>	<i>Blastococcus saxosidens</i>	<i>Brevibacillus choshimensis</i>	<i>Burkholderia vietnamiensis</i>
<i>Beijerinckia fluminensis</i>	<i>Blastochloris</i>	<i>Brevibacillus choshimensis</i>	<i>Buttiauxella</i>
<i>Beijerinckia indica</i>	<i>Blastochloris viridis</i>	<i>Brevibacillus invocatus</i>	<i>Buttiauxella agrestis</i>
<i>Beijerinckia mobilis</i>	<i>Blastomonas</i>	<i>Brevibacillus laterosporus</i>	<i>Buttiauxella bremeriae</i>
<i>Belliella</i>	<i>Blastomonas natatoria</i>	<i>Brevibacillus parabravis</i>	<i>Buttiauxella ferruginae</i>
<i>Belliella baltica</i>	<i>Blastopirellula</i>	<i>Brevibacterium</i>	<i>Buttiauxella goviniae</i>
<i>Bellilinea</i>	<i>Blastopirellula marina</i>	<i>Brevibacterium abidum</i>	<i>Buttiauxella tzardii</i>
<i>Bellilinea caldijstulae</i>	<i>Blautia</i>	<i>Brevibacterium album</i>	<i>Buttiauxella noackiae</i>
<i>Belnapia</i>	<i>Blautia coccoides</i>	<i>Brevibacterium aurantiacum</i>	<i>Buttiauxella warmboldiae</i>
<i>Belnapia moabensis</i>	<i>Blautia hanseni</i>	<i>Brevibacterium celere</i>	<i>Butyrvibrio</i>
<i>Bergierella</i>	<i>Blautia producta</i>	<i>Brevibacterium epidermidis</i>	<i>Butyrvibrio fibrisolvens</i>
<i>Bergierella denitrificans</i>	<i>Blautia westerae</i>	<i>Brevibacterium</i>	<i>Butyrvibrio hungatei</i>
<i>Beutenbergia</i>	<i>Bogoriella</i>	<i>frigoritolerans</i>	<i>Butyrvibrio proteoclasticus</i>
<i>Beutenbergia cavernae</i>	<i>Bogoriella caseilytica</i>	<i>Brevibacterium halotolerans</i>	
	<i>Bordetella</i>	<i>Brevibacterium iodinum</i>	
	<i>Bordetella avium</i>	<i>Brevibacterium linens</i>	
	<i>Bordetella bronchiseptica</i>	<i>Brevibacterium lyticum</i>	
	<i>Bordetella hinzii</i>	<i>Brevibacterium mcbrellneri</i>	
	<i>Bordetella holmesii</i>	<i>Brevibacterium otitidis</i>	
	<i>Bordetella parapertussis</i>	<i>Brevibacterium oxydans</i>	
	<i>Bordetella pertussis</i>	<i>Brevibacterium paucivorans</i>	
	<i>Bordetella petrii</i>	<i>Brevibacterium stationis</i>	
	<i>Bordetella trematum</i>		
	<i>B. glaucanolyticus</i>	<i>B. taeanensis</i>	<i>B. lautus</i>
<i>B. aminovorans</i>	<i>B. gordonae</i>	<i>B. tequilensis</i>	<i>B. lehensis</i>
<i>B. amylolyticus</i>	<i>B. gothelii</i>	<i>B. thermantarcticus</i>	<i>B. lentimorbus</i>
<i>B. andresenii</i>	<i>B. graminis</i>	<i>B. thermoaerophilus</i>	<i>B. lentus</i>
<i>B. aneurinilyticus</i>	<i>B. hadmapalus</i>	<i>B. thermoaerophilus</i>	<i>B. licheniformis</i>
<i>B. anthracis</i>	<i>B. haloalkaliphilus</i>	<i>B. thermoaerophilus</i>	<i>B. licheniformis</i>
<i>B. aquinaris</i>	<i>B. haloalkaliphilus</i>	<i>B. thermocatenulatus</i>	<i>B. ligniniphilus</i>
<i>B. arenosi</i>	<i>B. halocharis</i>	<i>B. thermocloacae</i>	<i>B. litoralis</i>
<i>B. arseniciselentatis</i>	<i>B. halodanitrificans</i>	<i>B. thermocloacae</i>	<i>B. locisalis</i>
<i>B. arsenicus</i>	<i>B. halodanitrificans</i>	<i>B. thermocloacae</i>	<i>B. luciferensis</i>
<i>B. aurantiacus</i>	<i>B. halodanitrificans</i>	<i>B. thermocloacae</i>	<i>B. luteolus</i>
<i>B. arvi</i>	<i>B. halophilus</i>	<i>B. thermocloacae</i>	<i>B. luteus</i>
<i>B. aryabhatai</i>	<i>B. halosaccharovorans</i>	<i>B. thermocloacae</i>	<i>B. macauensis</i>
<i>B. asahii</i>	<i>B. hemicyclotulolyticus</i>	<i>B. thermocloacae</i>	<i>B. macerans</i>
	<i>B. hemicyclotulolyticus</i>	<i>B. thermocloacae</i>	
<i>Bacillus</i>	<i>B. hemicyclotulolyticus</i>		
<i>B. acidoceler</i>	<i>B. hemicyclotulolyticus</i>		
<i>B. acidicola</i>	<i>B. hemicyclotulolyticus</i>		
<i>B. acidiproducens</i>	<i>B. hemicyclotulolyticus</i>		
<i>B. acidocaldarius</i>	<i>B. hemicyclotulolyticus</i>		
<i>B. acidoterrestris</i>	<i>B. hemicyclotulolyticus</i>		
<i>B. aeolius</i>	<i>B. hemicyclotulolyticus</i>		
<i>B. aerius</i>	<i>B. hemicyclotulolyticus</i>		
<i>B. aerophilus</i>	<i>B. hemicyclotulolyticus</i>		
<i>B. agaradhaerens</i>	<i>B. hemicyclotulolyticus</i>		
<i>B. agri</i>	<i>B. hemicyclotulolyticus</i>		
<i>B. aidungensis</i>	<i>B. hemicyclotulolyticus</i>		
<i>B. akibai</i>	<i>B. hemicyclotulolyticus</i>		
<i>B. alcalophilus</i>	<i>B. hemicyclotulolyticus</i>		

TABLE 1-continued

Example Bacteria		
Optionally, the target host cells are cells of a genus or species selected from this Table and/or the production strain cells are cells of a genus or species selected from this Table		
<i>B. algicola</i>	<i>B. atrophaeus</i>	<i>B. thermoruber</i>
<i>B. alginolyticus</i>	<i>B. axarquiensis</i>	<i>B. thermosphaericus</i>
<i>B. alkalicazotrophicus</i>	<i>B. azotoficans</i>	<i>B. thiaminolyticus</i>
<i>B. alkalinitritilicus</i>	<i>B. azotoformans</i>	<i>B. thioparans</i>
<i>B. alkalisediminis</i>	<i>B. badius</i>	<i>B. thuringiensis</i>
<i>B. alkalielluris</i>	<i>B. barbaricus</i>	<i>B. tianshenii</i>
<i>B. altitudinis</i>	<i>B. bataviensis</i>	<i>B. trypoxylicola</i>
<i>B. alveayensis</i>	<i>B. beijingensis</i>	<i>B. tuscatae</i>
<i>B. alvei</i>	<i>B. benzoovorans</i>	<i>B. validus</i>
<i>B. amyloliquefaciens</i>	<i>B. beringensis</i>	<i>B. vallismortis</i>
<i>B.</i>	<i>B. berkeleyi</i>	<i>B. vedderi</i>
a. subsp. <i>amyloliquefaciens</i>	<i>B. beveridgei</i>	<i>B. velezensis</i>
<i>B. a.</i> subsp. <i>planinarum</i>	<i>B. bogoriensis</i>	<i>B. vietnamensis</i>
	<i>B. boroniphilus</i>	<i>B. vulcani</i>
<i>B. diposauri</i>	<i>B. borstelensis</i>	<i>B. wakoensis</i>
<i>B. drentensis</i>	<i>B. brevis Migula</i>	<i>B. weihenstephanensis</i>
<i>B. edaphicus</i>	<i>B. butanolivorans</i>	<i>B. murimartini</i>
<i>B. ehimensis</i>	<i>B. canaverallus</i>	<i>B. mycoides</i>
<i>B. eiseniae</i>	<i>B. carboniphilus</i>	<i>B. naganoensis</i>
<i>B. enclensis</i>	<i>B. cecembensis</i>	<i>B. nanhaiensis</i>
<i>B. endophyticus</i>	<i>B. cellulolyticus</i>	<i>B. nanhaiisediminis</i>
<i>B. endoradicis</i>	<i>B. centrosporus</i>	<i>B. nealsonii</i>
<i>B. farraginis</i>	<i>B. cereus</i>	<i>B. neidei</i>
<i>B. fastidiosus</i>	<i>B. chagammorensis</i>	<i>B. neizhouensis</i>
<i>B. fengquensis</i>	<i>B. chitinolyticus</i>	<i>B. niabensis</i>
<i>B. firmus</i>	<i>B. chondroitinus</i>	<i>B. niacini</i>
<i>B. flexus</i>	<i>B. choshinensis</i>	<i>B. novalis</i>
<i>B. foraminis</i>	<i>B. chungangensis</i>	<i>B. oceanisediminis</i>
<i>B. fordii</i>	<i>B. citri</i>	<i>B. odysssei</i>
<i>B. formosus</i>	<i>B. circulans</i>	<i>B. okhensis</i>
<i>B. fortis</i>	<i>B. clarkii</i>	<i>B. okuhidensis</i>
<i>B. fumaroli</i>	<i>B. clausii</i>	<i>B. oleronius</i>
<i>B. funiculus</i>	<i>B. coagulans</i>	<i>B. oryzaecorticis</i>
<i>B. fusiformis</i>	<i>B. coahuilensis</i>	<i>B. oshimensis</i>
<i>B. galactophilus</i>	<i>B. cobiti</i>	<i>B. pabuli</i>
<i>B. galactosidilyticus</i>	<i>B. composti</i>	<i>B. pakistanensis</i>
<i>B. galliicensis</i>	<i>B. curdlanolyticus</i>	<i>B. pallidus</i>
<i>B. gelatini</i>	<i>B. cycloheptanicus</i>	<i>B. pallidus</i>
<i>B. gibsonii</i>	<i>B. cytotoxicus</i>	<i>B. panacisoli</i>
<i>B. ginsengi</i>	<i>B. daliensis</i>	<i>B. panaciterrae</i>
<i>B. ginsengihumi</i>	<i>B. decisiifrondis</i>	<i>B. pantothenicus</i>
<i>B. ginsengisoli</i>	<i>B. decolorationis</i>	<i>B. parabrevis</i>
<i>B. globisporus</i> (eg. <i>B.</i>	<i>B. deserti</i>	<i>B. pasteurii</i>
g. subsp. <i>Globisporus</i> ; or <i>B.</i>		<i>B. patagoniensis</i>
g. subsp. <i>Martinius</i>)		
	<i>B. herbersteinensis</i>	
	<i>B. horikoshii</i>	
	<i>B. horneckiae</i>	
	<i>B. horri</i>	
	<i>B. hutzhouensis</i>	
	<i>B. humi</i>	
	<i>B. huajinpoensis</i>	
	<i>B. idriensis</i>	
	<i>B. indicus</i>	
	<i>B. infantis</i>	
	<i>B. infernus</i>	
	<i>B. insolitus</i>	
	<i>B. invictae</i>	
	<i>B. iranensis</i>	
	<i>B. isabellae</i>	
	<i>B. isronensis</i>	
	<i>B. jeotgali</i>	
	<i>B. kaustophilus</i>	
	<i>B. kobensis</i>	
	<i>B. kochii</i>	
	<i>B. kokeshiformis</i>	
	<i>B. koreensis</i>	
	<i>B. korlensis</i>	
	<i>B. kribbensis</i>	
	<i>B. kratwichiae</i>	
	<i>B. laeulichiacus</i>	
	<i>B. larvae</i>	
	<i>B. laterosporus</i>	
	<i>B. salxivigens</i>	
	<i>B. saliphilus</i>	
	<i>B. schlegelii</i>	
	<i>B. sediminis</i>	
	<i>B. selenitarsenatis</i>	
	<i>B. selenitriducens</i>	
	<i>B. seohaeanensis</i>	
	<i>B. shachensis</i>	
	<i>B. shackletonii</i>	
	<i>B. siamensis</i>	
	<i>B. silvestris</i>	
	<i>B. simplex</i>	
	<i>B. stralis</i>	
	<i>B. smithii</i>	
	<i>B. soli</i>	
	<i>B. solimangrovi</i>	
	<i>B. solisalsi</i>	
	<i>B. songkdensis</i>	
	<i>B. sonorensis</i>	
	<i>B. sphaericus</i>	
	<i>B. sporothermophilus</i>	
	<i>B. steatothermophilus</i>	
	<i>B. macquariensis</i>	
	<i>B. macyae</i>	
	<i>B. malacitensis</i>	
	<i>B. mannilyticus</i>	
	<i>B. marisylavi</i>	
	<i>B. marismortui</i>	
	<i>B. marmarensis</i>	
	<i>B. massiliensis</i>	
	<i>B. megaterium</i>	
	<i>B. mesonae</i>	
	<i>B. methanolicus</i>	
	<i>B. methylotrophicus</i>	
	<i>B. migulamus</i>	
	<i>B. mojavensis</i>	
	<i>B. mucilaginosus</i>	
	<i>B. muralis</i>	
	<i>B. murimartini</i>	
	<i>B. mycoides</i>	
	<i>B. naganoensis</i>	
	<i>B. nanhaiensis</i>	
	<i>B. nanhaiisediminis</i>	
	<i>B. nealsonii</i>	
	<i>B. neidei</i>	
	<i>B. neizhouensis</i>	
	<i>B. niabensis</i>	
	<i>B. niacini</i>	
	<i>B. novalis</i>	
	<i>B. oceanisediminis</i>	
	<i>B. odysssei</i>	
	<i>B. okhensis</i>	
	<i>B. okuhidensis</i>	
	<i>B. oleronius</i>	
	<i>B. oryzaecorticis</i>	
	<i>B. oshimensis</i>	
	<i>B. pabuli</i>	
	<i>B. pakistanensis</i>	
	<i>B. pallidus</i>	
	<i>B. pallidus</i>	
	<i>B. panacisoli</i>	
	<i>B. panaciterrae</i>	
	<i>B. pantothenicus</i>	
	<i>B. parabrevis</i>	
	<i>B. pasteurii</i>	
	<i>B. patagoniensis</i>	

TABLE 1—continued

Optionally, the target host cells are cells of a genus or species selected from this Table and/or the production strain cells are cells of a genus or species selected from this Table

Example Bacteria

Caenimonas	Campylobacter	Catenuloplanes	Curtobacterium
Caenimonas koreensis	Campylobacter coli	Catenuloplanes arovinosus	Curtobacterium albidum
Caldakalibacillus	Campylobacter concisus	Catenuloplanes castaneus	Curtobacterium citreus
Caldakalibacillus uzonensis	Campylobacter curvus	Catenuloplanes crispus	
Caldanaerobacter	Campylobacter fetus	Catenuloplanes indicus	
Caldanaerobacter subterraneus	Campylobacter gracilis	Catenuloplanes japonicus	
Caldanaerobius	Campylobacter helveticus	Catenuloplanes nepalensis	
Caldanaerobius ifjiensis	Campylobacter hominis	Catenuloplanes niger	
Caldanaerobius	Campylobacter hyointestinalis	Chryseobacterium	
polysaccharolyticus	Campylobacter jejuni	Chryseobacterium	
Caldanaerobius zeae	Campylobacter lari	balustinum	
Caldanaerovirga	Campylobacter mucosalis	Citrobacter	
Caldanaerovirga acetisignens	Campylobacter reclus	C. analonaticus	
Caldicellulosiruptor	Campylobacter showae	C. braaki	
Caldicellulosiruptor bescii	Campylobacter sputorum	C. diversus	
Caldicellulosiruptor krisjanssonii	Campylobacter upsaliensis	C. farmeri	
Caldicellulosiruptor owensensis	Capnocytophaga	C. freundi	
	Capnocytophaga canimorsus	C. gillenii	
	Capnocytophaga cynodegmi	C. koxeri	
	Capnocytophaga gingivialis	C. murliniae	
	Capnocytophaga granulosa	C. pasteurii ¹¹	
	Capnocytophaga haemolytica	C. rodentium	
	Capnocytophaga ochracea	C. sedlakii	
	Capnocytophaga sputigena	C. werkananii	
		C. youngae	
		Clostridium	
		(see below)	
		Coccolhoris	
		Coccolhoris elabens	
		Corynebacterium	
		Corynebacterium flavescens	
		Corynebacterium variabile	

Clostridium	Clostridium aciditolerans	Clostridium acidurici	Clostridium aerotolerans	Clostridium
Clostridium absonum	Clostridium acetobutylicum	Clostridium acidisoli	Clostridium albidum	Clostridium
Clostridium aescuarri	Clostridium algidicarni	Clostridium algidicarni	Clostridium alginolyticum	Clostridium
Clostridium akagi	Clostridium aldrichi	Clostridium algidicarni	Clostridium alginolyticum	Clostridium
Clostridium aminophilum	Clostridium aminovalericum	Clostridium amygdalinum	Clostridium amylolyticum	Clostridium
Clostridium aurantibutyricum	Clostridium autoethanogenum	Clostridium baratii	Clostridium bartlettii	Clostridium
Clostridium botulinum	Clostridium bowmanii	Clostridium bayricum	Clostridium cadaveris	Clostridium
Clostridium carnis	Clostridium cavendishii	Clostridium celerecescens	Clostridium cellulosum	Clostridium
Clostridium cellulovorans	Clostridium charatapidum	Clostridium chauvoei	Clostridium chromiireducens	Clostridium
Clostridium cochlearium	Clostridium collettii	Clostridium coliticum	Clostridium collagenovorans	Clostridium
Clostridium durum	Clostridium colletii	Clostridium coliticum	Clostridium collagenovorans	Clostridium
Clostridium drakei	Clostridium estertheticum	Clostridium coliticum	Clostridium collagenovorans	Clostridium
Clostridium formicaceticum	Clostridium estertheticum	Clostridium coliticum	Clostridium collagenovorans	Clostridium
Clostridium fimetarium	Clostridium estertheticum	Clostridium coliticum	Clostridium collagenovorans	Clostridium
Clostridium frigidarium	Clostridium estertheticum	Clostridium coliticum	Clostridium collagenovorans	Clostridium
Clostridium gonghswense	Clostridium estertheticum	Clostridium coliticum	Clostridium collagenovorans	Clostridium
Clostridium granatii	Clostridium estertheticum	Clostridium coliticum	Clostridium collagenovorans	Clostridium
Clostridium haemolyticum	Clostridium estertheticum	Clostridium coliticum	Clostridium collagenovorans	Clostridium
Clostridium hathewayi	Clostridium estertheticum	Clostridium coliticum	Clostridium collagenovorans	Clostridium
Clostridium hastiforme	Clostridium estertheticum	Clostridium coliticum	Clostridium collagenovorans	Clostridium
Clostridium hiranonis	Clostridium estertheticum	Clostridium coliticum	Clostridium collagenovorans	Clostridium

TABLE 1-continued

Example Bacteria	
Optionally, the target host cells are cells of a genus or species selected from this Table and/or the production strain cells are cells of a genus or species selected from this Table	Optionally, the target host cells are cells of a genus or species selected from this Table and/or the production strain cells are cells of a genus or species selected from this Table
<i>Clostridium histolyticum</i> , <i>Clostridium homopropionicum</i> , <i>Clostridium huakuii</i> , <i>Clostridium hungatei</i> , <i>Clostridium hydrogeniformans</i> , <i>Clostridium hydroxybenzoicum</i> , <i>Clostridium hylemonae</i> , <i>Clostridium jejuniense</i> , <i>Clostridium indolis</i> , <i>Clostridium innocuum</i> , <i>Clostridium inestinale</i> , <i>Clostridium irregulare</i> , <i>Clostridium isaitidis</i> , <i>Clostridium josui</i> , <i>Clostridium klayveri</i> , <i>Clostridium lactatifermentans</i> , <i>Clostridium lacusphysellense</i> , <i>Clostridium larantiense</i> , <i>Clostridium lavadense</i> , <i>Clostridium lentocellum</i> , <i>Clostridium leptum</i> , <i>Clostridium limosum</i> , <i>Clostridium litorale</i> , <i>Clostridium lituseburense</i> , <i>Clostridium ljungdahlii</i> , <i>Clostridium lortetii</i> , <i>Clostridium lundense</i> , <i>Clostridium madenominatum</i> , <i>Clostridium mangenotii</i> , <i>Clostridium mayombei</i> , <i>Clostridium methoxybenzovorans</i> , <i>Clostridium methylophenosum</i> , <i>Clostridium neopropionicum</i> , <i>Clostridium nexile</i> , <i>Clostridium nitrophenolicum</i> , <i>Clostridium novyi</i> , <i>Clostridium oceanicum</i> , <i>Clostridium orbiscidens</i> , <i>Clostridium oroticum</i> , <i>Clostridium oxalicum</i> , <i>Clostridium papayrosolvens</i> , <i>Clostridium paradoxum</i> , <i>Clostridium paraperfringens</i> (Alias: C. welchii), <i>Clostridium paraputrificum</i> , <i>Clostridium pascui</i> , <i>Clostridium pasteurianum</i> , <i>Clostridium peptidivorans</i> , <i>Clostridium perenne</i> , <i>Clostridium peffringens</i> , <i>Clostridium plennigii</i> , <i>Clostridium phytofermentans</i> , <i>Clostridium piliforme</i> , <i>Clostridium polysaccharolyticum</i> , <i>Clostridium populeri</i> , <i>Clostridium propionicum</i> , <i>Clostridium proteoclasticum</i> , <i>Clostridium proteolyticum</i> , <i>Clostridium psychrophilum</i> , <i>Clostridium punicium</i> , <i>Clostridium purinilyticum</i> , <i>Clostridium putrefaciens</i> , <i>Clostridium purificans</i> , <i>Clostridium quercicolum</i> , <i>Clostridium quini</i> , <i>Clostridium ramosum</i> , <i>Clostridium rectum</i> , <i>Clostridium roseum</i> , <i>Clostridium saccharobutylicum</i> , <i>Clostridium saccharoguttatum</i> , <i>Clostridium saccharoperbutylacetonicum</i> , <i>Clostridium saratinense</i> , <i>Clostridium sartagoforme</i> , <i>Clostridium scatogenes</i> , <i>Clostridium schimacherense</i> , <i>Clostridium scindens</i> , <i>Clostridium seidelii</i> , <i>Clostridium sphenoides</i> , <i>Clostridium spiriforme</i> , <i>Clostridium sporogenes</i> , <i>Clostridium sporosphaeroides</i> , <i>Clostridium stercorarium</i> , <i>Clostridium stercorarium leptosporium</i> , <i>Clostridium stercorarium stercorarium</i> , <i>Clostridium stercorarium thermolacticum</i> , <i>Clostridium sticklandii</i> , <i>Clostridium straminisolvens</i> , <i>Clostridium subterminalis</i> , <i>Clostridium sufflavum</i> , <i>Clostridium sulfidigenes</i> , <i>Clostridium symbiosum</i> , <i>Clostridium tagluense</i> , <i>Clostridium tepidiprofundii</i> , <i>Clostridium termitidis</i> , <i>Clostridium tertium</i> , <i>Clostridium tetani</i> , <i>Clostridium tetanomorphum</i> , <i>Clostridium thermacetivum</i> , <i>Clostridium thermoautotrophicum</i> , <i>Clostridium thermoacetaliphilum</i> , <i>Clostridium thermobutylicum</i> , <i>Clostridium thermocellum</i> , <i>Clostridium thermocopriace</i> , <i>Clostridium thermohydrosulfuricum</i> , <i>Clostridium thermolacticum</i> , <i>Clostridium thermopalmarium</i> , <i>Clostridium thermopapyrolyticum</i> , <i>Clostridium thermosaccharolyticum</i> , <i>Clostridium thermosuccinogenes</i> , <i>Clostridium thermosulfurigenes</i> , <i>Clostridium thiosulfatireducens</i> , <i>Clostridium tyrobutyricum</i> , <i>Clostridium uliginosum</i> , <i>Clostridium villosum</i> , <i>Clostridium villosum</i> , <i>Clostridium vincentii</i> , <i>Clostridium viride</i> , <i>Clostridium xylanolyticum</i> , <i>Clostridium xylanovorans</i>	<i>Deinococcus</i> <i>Deinococcus aerius</i> <i>Deinococcus apachensis</i> <i>Deinococcus aquaticus</i> <i>Deinococcus aquatilis</i> <i>Deinococcus caeni</i> <i>Deinococcus radiodurans</i> <i>Deinococcus radiophilus</i> <i>Enterobacter kobei</i> <i>E. ludwigii</i> <i>E. mori</i> <i>E. nimipressuralis</i> <i>E. arachidis</i> <i>E. asburiae</i> <i>E. cancerogenus</i> <i>E. cloacae</i> <i>E. cowanii</i> <i>E. dissolvens</i> <i>E. gergoviae</i> <i>E. helveticus</i> <i>E. hormaechei</i> <i>E. intermedius</i>
<i>Dactylosporangium</i> <i>Dactylosporangium aurantiacum</i> <i>Dactylosporangium fulvum</i> <i>Dactylosporangium matsuzakiense</i> <i>Dactylosporangium roseum</i> <i>Dactylosporangium thailandense</i> <i>Dactylosporangium vinaceum</i>	<i>Faecalibacterium</i> <i>Faecalibacterium prausnitzii</i> <i>Fangia</i> <i>Fangia hongkongensis</i> <i>Fastidiosipila</i> <i>Fastidiosipila sanguinis</i> <i>Fusobacterium</i> <i>Fusobacterium nucleatum</i>
<i>Enterobacter</i> <i>E. aerogenes</i> <i>E. amnigenis</i> <i>E. agglomerans</i> <i>E. arachidis</i> <i>E. asburiae</i> <i>E. cancerogenus</i> <i>E. cloacae</i> <i>E. cowanii</i> <i>E. dissolvens</i> <i>E. gergoviae</i> <i>E. helveticus</i> <i>E. hormaechei</i> <i>E. intermedius</i>	<i>Faecalibacterium</i> <i>Faecalibacterium prausnitzii</i> <i>Fangia</i> <i>Fangia hongkongensis</i> <i>Fastidiosipila</i> <i>Fastidiosipila sanguinis</i> <i>Fusobacterium</i> <i>Fusobacterium nucleatum</i>
<i>Gaebulibacter</i> <i>Gaebulibacter saenamkumensis</i> <i>Gallibacterium</i> <i>Gallibacterium anatis</i> <i>Gallicola</i> <i>Gallicola barnesae</i> <i>Garcitella</i>	<i>Flavobacterium</i> <i>Flavobacterium antarcticum</i> <i>Flavobacterium aquatile</i> <i>Flavobacterium aquidulense</i> <i>Flavobacterium balustinum</i> <i>Flavobacterium croceum</i> <i>Flavobacterium cucumis</i> <i>Flavobacterium daejeonense</i> <i>Flavobacterium defluvi</i> <i>Flavobacterium degerlachei</i> <i>Flavobacterium denitrificans</i> <i>Flavobacterium flum</i> <i>Flavobacterium flevense</i> <i>Flavobacterium frigidarium</i> <i>Flavobacterium mizutai</i> <i>Flavobacterium okeanokoites</i> <i>Janibacter</i> <i>Janibacter anophelis</i> <i>Janibacter corallicola</i> <i>Janibacter limosus</i> <i>Janibacter melonis</i> <i>Janibacter terrae</i> <i>Jannaschia</i>
<i>Gaebulibacter</i> <i>Gaebulibacter saenamkumensis</i> <i>Gallibacterium</i> <i>Gallibacterium anatis</i> <i>Gallicola</i> <i>Gallicola barnesae</i> <i>Garcitella</i>	<i>Ideonella</i> <i>Ideonella azotifigens</i> <i>Idiomarina</i> <i>Idiomarina abyssalis</i> <i>Idiomarina ballica</i> <i>Idiomarina fontislapidosi</i> <i>Idiomarina loihiensis</i>

TABLE 1—continued

Optionally, the target host cells are cells of a genus or species selected from this Table and/or the production strain cells are cells of a genus or species selected from this Table		Example Bacteria
<i>Garcilla nitratireducens</i>	<i>Haemophilus paracuniculus</i>	<i>Idiomarina ramibicola</i>
<i>Geobacillus</i>	<i>Haemophilus parahaemolyticus</i>	<i>Idiomarina seosinensis</i>
<i>Geobacillus thermoglucosidarius</i>	<i>Haemophilus parainfluenzae</i>	<i>Idiomarina zobellii</i>
<i>Geobacillus stearothermophilus</i>	<i>Haemophilus paraprohaemolyticus</i>	<i>Ignatzschineria</i>
<i>Geobacter</i>	<i>Haemophilus parasuis</i>	<i>Ignatzschineria larvae</i>
<i>Geobacter bemiidensis</i>	<i>Haemophilus pitmaniae</i>	<i>Ignavigranum</i>
<i>Geobacter bremensis</i>	<i>Hafnia</i>	<i>Ignavigranum ruoffiae</i>
<i>Geobacter chapellet</i>	<i>Hafnia alvei</i>	<i>Ilumatobacter</i>
<i>Geobacter grbicata</i>	<i>Hahella</i>	<i>Ilumatobacter fluminis</i>
<i>Geobacter hydrogenophilus</i>	<i>Hahella ganghwensis</i>	<i>Ilyobacter</i>
<i>Geobacter lovleyi</i>	<i>Halalkalibacillus</i>	<i>Ilyobacter delafeldii</i>
<i>Geobacter metallireducens</i>	<i>Halalkalibacillus halophilus</i>	<i>Ilyobacter insuetus</i>
<i>Geobacter pelophilus</i>	<i>Helicobacter</i>	<i>Ilyobacter polytropus</i>
<i>Geobacter pickeringii</i>	<i>Helicobacter pylori</i>	<i>Ilyobacter tartaricus</i>
<i>Geobacter sulfurreducens</i>		
<i>Geodermatophilus</i>		
<i>Geodermatophilus obscurus</i>		
<i>Gluconacetobacter</i>		
<i>Gluconacetobacter xylinus</i>		
<i>Gordonia</i>		
<i>Gordonia rubripertincta</i>		
<i>Kaistia</i>	<i>Labedella</i>	<i>Listeria ivanovii</i>
<i>Kaistia adipata</i>	<i>Labedella gwalgjensis</i>	<i>L. marthii</i>
<i>Kaistia soli</i>	<i>Labrenzia</i>	<i>L. monocytogenes</i>
<i>Kangella</i>	<i>Labrenzia aggregata</i>	<i>L. newyorkensis</i>
<i>Kangella aquimarina</i>	<i>Labrenzia alba</i>	<i>L. riparia</i>
<i>Kangella koreensis</i>	<i>Labrenzia alexandrii</i>	<i>L. rocourtae</i>
	<i>Labrenzia marina</i>	<i>L. seeligeri</i>
<i>Kerstersia</i>	<i>Labrys</i>	<i>L. weihenstephanensis</i>
<i>Kerstersia gytonum</i>	<i>Labrys methylaminiphilus</i>	<i>L. welschimeri</i>
<i>Kiloniella</i>	<i>Labrys miyogiensis</i>	<i>Listonella</i>
<i>Kiloniella laminariae</i>	<i>Labrys monachus</i>	<i>Listonella anguillarum</i>
<i>Klebsiella</i>	<i>Labrys okinawensis</i>	<i>Macrococcus</i>
<i>K. granitiformis</i>	<i>Labrys portucalensis</i>	<i>Macrococcus bovicus</i>
<i>K. oxytoca</i>		<i>Marinobacter</i>
<i>K. pneumoniae</i>	<i>Lactobacillus</i>	<i>Marinobacter algicola</i>
<i>K. terrigena</i>	[see below]	<i>Marinobacter bryozorum</i>
<i>Klavera</i>	<i>Laceyella</i>	<i>Marinobacter flavimaris</i>
	<i>Laceyella putida</i>	<i>Meiothermus</i>
<i>Kluyvera</i>	<i>Lechevalieria</i>	<i>Meiothermus ruber</i>
<i>Kluyvera ascorbata</i>	<i>Lechevalieria aerocolonigenes</i>	<i>Methylophilus</i>
<i>Kocuria</i>	<i>Legionella</i>	<i>Methylophilus methylotrophus</i>
<i>Kocuria rosea</i>	[see below]	<i>Microbacterium</i>
<i>Kocuria varians</i>	<i>Listeria</i>	<i>Microbacterium ammoniaphilum</i>
<i>Kurthia</i>	<i>L. aquatica</i>	<i>Microbacterium arborescens</i>
<i>Kurthia zopfii</i>	<i>L. boorica</i>	<i>Microbacterium liquefaciens</i>
	<i>L. cornellensis</i>	<i>Microbacterium oxidans</i>
	<i>L. fleischmannii</i>	
	<i>L. floridensis</i>	
	<i>L. grandensis</i>	
		<i>Micrococcus</i>
		<i>Micrococcus luteus</i>
		<i>Micrococcus lysae</i>
		<i>Moraxella</i>
		<i>Moraxella bovis</i>
		<i>Moraxella nonliquefaciens</i>
		<i>Moraxella osloensis</i>
		<i>Nakamuraella</i>
		<i>Nakamuraella multipartita</i>
		<i>Nannocystis</i>
		<i>Nannocystis pusilla</i>
		<i>Natranaerobius</i>
		<i>Natranaerobius</i>
		<i>thermophilus</i>
		<i>Natranaerobius trueperi</i>
		<i>Naxibacter</i>
		<i>Naxibacter alkalicolerans</i>
		<i>Neisseria</i>
		<i>Neisseria cinerea</i>
		<i>Neisseria dentrificans</i>
		<i>Neisseria gonorrhoeae</i>
		<i>Neisseria lactamica</i>
		<i>Neisseria mucosa</i>
		<i>Neisseria sicca</i>
		<i>Neisseria subflava</i>
		<i>Neptunomonas</i>
		<i>Neptunomonas japonica</i>
		<i>Nesterenkonia</i>
		<i>Nesterenkonia holobia</i>
		<i>Nocardia</i>
		<i>Nocardia argentinensis</i>
		<i>Nocardia coralina</i>
		<i>Nocardia</i>
		<i>otitidiscaivarium</i>
		<i>Jannaschia cystaugens</i>
		<i>Jannaschia helgolandensis</i>
		<i>Jannaschia pohangensis</i>
		<i>Jannaschia rubra</i>
		<i>Janthinobacterium</i>
		<i>Janthinobacterium</i>
		<i>agaricidamnosum</i>
		<i>Janthinobacterium lividum</i>
		<i>Jejuia</i>
		<i>Jejuia pallidilutea</i>
		<i>Jeotgalibacillus</i>
		<i>Jeotgalibacillus</i>
		<i>alimentarius</i>
		<i>Jeotgalicoccus</i>
		<i>Jeotgalicoccus halotolerans</i>

TABLE 1-continued

Optionally, the target host cells are cells of a genus or species selected from this Table and/or the production strain cells are cells of a genus or species selected from this Table	
Example Bacteria	
<i>Lactobacillus</i>	
<i>L. acetolerans</i>	
<i>L. acidifarinae</i>	
<i>L. acidipiscis</i>	
<i>L. acidophilus</i>	
<i>Lactobacillus agilis</i>	
<i>L. algidus</i>	
<i>L. alimentarius</i>	
<i>L. amylolyticus</i>	
<i>L. amylophilus</i>	
<i>L. amylophilicus</i>	
<i>L. amylovorus</i>	
<i>L. animalis</i>	
<i>L. antri</i>	
<i>L. apodemi</i>	
<i>L. aviaris</i>	
<i>L. bifementans</i>	
<i>L. brevis</i>	
<i>L. buchneri</i>	
<i>L. camelinae</i>	
<i>L. casei</i>	
<i>L. kitasatonis</i>	
<i>L. kumkei</i>	
<i>L. leichmannii</i>	
<i>L. lindneri</i>	
<i>L. malefermentans</i>	
<i>Legionella</i>	
<i>Legionella adelaidensis</i>	
<i>Legionella anisa</i>	
<i>Legionella bellardensis</i>	
<i>Legionella birminghamensis</i>	
<i>Legionella bozemanae</i>	
<i>Legionella brunensis</i>	
<i>Legionella busanensis</i>	
<i>Legionella cardiaca</i>	
<i>Legionella cherrii</i>	
<i>Legionella cincinnatiensis</i>	
<i>Legionella clemsonensis</i>	
<i>Legionella donaldsonii</i>	
<i>Oceanibulbus</i>	
<i>Oceanibulbus indotifex</i>	
<i>L. grayi</i>	
<i>L. innocua</i>	
<i>L. cateniformis</i>	
<i>L. ceti</i>	
<i>L. coleohominis</i>	
<i>L. collinoides</i>	
<i>L. composti</i>	
<i>L. concavus</i>	
<i>L. coryniformis</i>	
<i>L. crispatus</i>	
<i>L. crustorum</i>	
<i>L. curvatus</i>	
<i>L. delbrueckii</i> subsp. <i>bulgaricus</i>	
<i>L. delbrueckii</i> subsp.	
<i>delbrueckii</i>	
<i>L. delbrueckii</i> subsp. <i>lactis</i>	
<i>L. dextrinicus</i>	
<i>L. dthivorans</i>	
<i>L. equi</i>	
<i>L. equigenerosi</i>	
<i>L. farraginis</i>	
<i>L. farcininis</i>	
<i>L. fermentum</i>	
<i>L. fornicalis</i>	
<i>L. fructivorans</i>	
<i>L. frumenti</i>	
<i>Candidatus Legionella jeonii</i>	
<i>Legionella jordanis</i>	
<i>Legionella lansingensis</i>	
<i>Legionella londinensis</i>	
<i>Legionella longbeachae</i>	
<i>Legionella lyica</i>	
<i>Legionella macacahernii</i>	
<i>Legionella massiliensis</i>	
<i>Legionella micdadei</i>	
<i>Legionella monrovia</i>	
<i>Legionella moravica</i>	
<i>Legionella nagasakiensis</i>	
<i>Legionella nautarum</i>	
<i>Legionella noritandica</i>	
<i>Legionella oakridgensis</i>	
<i>Legionella parisiensis</i>	
<i>Legionella pitsburghensis</i>	
<i>Legionella pneumophila</i>	
<i>Legionella quateirensis</i>	
<i>Prevotella</i>	
<i>Prevotella albensis</i>	
<i>Paenibacillus</i>	
<i>Paenibacillus thiaminolyticus</i>	
<i>L. mali</i>	
<i>L. manihotivivans</i>	
<i>L. mindensis</i>	
<i>L. mucosae</i>	
<i>L. murinus</i>	
<i>L. nagelii</i>	
<i>L. nanurensis</i>	
<i>L. nantensis</i>	
<i>L. oligofermentans</i>	
<i>L. oris</i>	
<i>L. panis</i>	
<i>L. pantheris</i>	
<i>L. parabrevis</i>	
<i>L. parabuchneri</i>	
<i>L. paracasei</i>	
<i>L. paracollinoides</i>	
<i>L. parafarraginis</i>	
<i>L. homohiochii</i>	
<i>L. iners</i>	
<i>L. ingluviei</i>	
<i>L. intestinalis</i>	
<i>L. fuchsensis</i>	
<i>L. gallinarum</i>	
<i>L. gasserii</i>	
<i>Legionella quinlivanii</i>	
<i>Legionella rowbothamii</i>	
<i>Legionella rubrilucens</i>	
<i>Legionella sainthelensii</i>	
<i>Legionella sanicrucis</i>	
<i>Legionella shakespearei</i>	
<i>Legionella spiritensis</i>	
<i>Legionella steelei</i>	
<i>Legionella steigerwaltii</i>	
<i>Legionella taurinensis</i>	
<i>Legionella tussonensis</i>	
<i>Legionella tunisensis</i>	
<i>Legionella wadsworthii</i>	
<i>Legionella waltersii</i>	
<i>Legionella worstelensis</i>	
<i>Legionella yabuuchiiae</i>	
<i>Quadriflustra</i>	
<i>Quadriflustra granulorum</i>	

TABLE 1-continued

Optionally, the target host cells are cells of a genus or species selected from this Table and/or the production strain cells are cells of a genus or species selected from this Table	
<i>Oceanicaulis</i>	<i>Prevotella amnii</i>
<i>Oceanicaulis alexandrii</i>	<i>Prevotella bergensis</i>
<i>Oceanicola</i>	<i>Prevotella bivia</i>
<i>Oceanicola batsensis</i>	<i>Prevotella brevis</i>
<i>Oceanicola granulosis</i>	<i>Prevotella bryantii</i>
<i>Oceanicola nanhatensis</i>	<i>Prevotella buccae</i>
<i>Oceanimonas</i>	<i>Prevotella buccalis</i>
<i>Oceanimonas baumannii</i>	<i>Prevotella copri</i>
<i>Oceaniserpentilla</i>	<i>Prevotella dentalis</i>
<i>Oceaniserpentilla halitosis</i>	<i>Prevotella denticola</i>
<i>Oceanisphaera</i>	<i>Prevotella disiens</i>
<i>Oceanisphaera donghaensis</i>	<i>Prevotella hisitcola</i>
<i>Oceanisphaera litoralis</i>	<i>Prevotella intermedia</i>
<i>Oceanithermus</i>	<i>Prevotella maculosa</i>
<i>Oceanithermus desulfurans</i>	<i>Prevotella marshallii</i>
<i>Oceanithermus profundus</i>	<i>Prevotella melaninogenica</i>
<i>Oceanobacillus</i>	<i>Prevotella micans</i>
<i>Oceanobacillus caeni</i>	<i>Prevotella multififormis</i>
<i>Oceanospirillum</i>	<i>Prevotella nigrescens</i>
<i>Oceanospirillum linum</i>	<i>Prevotella oralis</i>
	<i>Prevotella oris</i>
	<i>Prevotella oulorum</i>
	<i>Prevotella pallens</i>
	<i>Prevotella salivae</i>
	<i>Prevotella stercora</i>
	<i>Prevotella tanneriae</i>
	<i>Prevotella timonensis</i>
	<i>Prevotella veroralis</i>
	<i>Providencia</i>
	<i>Providencia stuartii</i>
	<i>Pseudomonas</i>
	<i>Pseudomonas aeruginosa</i>
	<i>Pseudomonas alcaligenes</i>
	<i>Pseudomonas anguilliseptica</i>
	<i>Pseudomonas fluorescens</i>
	<i>Pseudomonas fluorescens</i>
	<i>haloplanktis</i>
	<i>Pseudomonas mendocina</i>
	<i>Pseudomonas</i>
	<i>pseudocaligenes</i>
	<i>Pseudomonas putida</i>
	<i>Pseudomonas tutzeri</i>
	<i>Pseudomonas syringae</i>
	<i>Psychrobacter</i>
	<i>Psychrobacter faecalis</i>
	<i>Psychrobacter</i>
	<i>phenylpyruvicus</i>
	<i>Sanguibacter</i>
	<i>Sanguibacter keddiei</i>
	<i>Sanguibacter suarezi</i>
<i>Saccharococcus</i>	<i>Stenotrophomonas</i>
<i>Saccharococcus thermophilus</i>	<i>Stenotrophomonas</i>
<i>Saccharomonospora</i>	<i>mallophila</i>
	<i>Tatlockia</i>
	<i>Tatlockia maceachernii</i>
	<i>Tatlockia micdadei</i>

TABLE 1-continued

Optionally, the target host cells are cells of a genus or species selected from this Table and/or the production strain cells are cells of a genus or species selected from this Table		Example Bacteria	
<i>Saccharomonospora azurea</i>	<i>Salegmitibacter salegens</i>	<i>Saprospira</i>	<i>Streptococcus</i>
<i>Saccharomonospora cyanea</i>	<i>Salimicrobium</i>	<i>Saprospira grandis</i>	[also see below]
<i>Saccharomonospora viridis</i>	<i>Salimicrobium album</i>	<i>Sarcina</i>	<i>Streptomyces</i>
<i>Saccharophagus</i>	<i>Salinibacter</i>	<i>Sarcina maxima</i>	<i>Streptomyces</i>
<i>Saccharophagus degradans</i>	<i>Salinibacter ruber</i>	<i>Sarcina ventriculi</i>	<i>achromogenes</i>
<i>Saccharopolyspora</i>	<i>Salinicoccus</i>	<i>Sebadella</i>	<i>Streptomyces cesalbus</i>
<i>Saccharopolyspora erythraea</i>	<i>Salinicoccus alkaliophilus</i>	<i>Sebadella termitidis</i>	<i>Streptomyces cesaetiposus</i>
<i>Saccharopolyspora gregori</i>	<i>Salinicoccus hispanicus</i>		<i>Streptomyces cesdiastaticus</i>
<i>Saccharopolyspora hirsuta</i>	<i>Salinicoccus roseus</i>		<i>Streptomyces cesxifolius</i>
<i>Saccharopolyspora hordei</i>	<i>Salinispora</i>	<i>Serratia</i>	<i>Streptomyces fimbriatus</i>
<i>Saccharopolyspora recitvirgula</i>	<i>Salinispora arenicola</i>	<i>Serratia fonticola</i>	<i>Streptomyces fradiae</i>
<i>Saccharopolyspora spinosa</i>	<i>Salinispora tropica</i>	<i>Serratia marcescens</i>	<i>Streptomyces fulvissimus</i>
<i>Saccharopolyspora taberi</i>	<i>Salinivibrio</i>	<i>Sphaerotilus</i>	<i>Streptomyces griseoruber</i>
<i>Saccharothrix</i>	<i>Salinivibrio costicola</i>	<i>Sphaerotilus natans</i>	<i>Streptomyces griseus</i>
<i>Saccharothrix australiensis</i>	<i>Salmonella</i>	<i>Sphingobacterium</i>	<i>Streptomyces lavendulae</i>
<i>Saccharothrix coeruleofusca</i>	<i>Salmonella bongori</i>	<i>Sphingobacterium multivorum</i>	<i>Streptomyces</i>
<i>Saccharothrix espanaensis</i>	<i>Salmonella enterica</i>	<i>Staphylococcus</i>	<i>phaeochromogenes</i>
<i>Saccharothrix longispora</i>	<i>Salmonella subterranea</i>	[see below]	<i>thermodiastaticus</i>
<i>Saccharothrix mutabilis</i>	<i>Salmonella typhi</i>		<i>Thermus</i>
<i>Saccharothrix syringae</i>			<i>Thermus aquaticus</i>
<i>Saccharothrix tangerinus</i>			<i>Thermus filiformis</i>
<i>Saccharothrix texasensis</i>			<i>Thermus thermophilus</i>
<i>Staphylococcus</i>			
<i>S. arlettae</i>	<i>S. equorum</i>	<i>S. microti</i>	<i>S. schleiferi</i>
<i>S. agnetis</i>	<i>S. felis</i>	<i>S. muscae</i>	<i>S. scituri</i>
<i>S. aureus</i>	<i>S. fleuretii</i>	<i>S. nepalensis</i>	<i>S. simiae</i>
<i>S. auricularis</i>	<i>S. gallinarum</i>	<i>S. pasteurii</i>	<i>S. simulans</i>
<i>S. capitis</i>	<i>S. haemolyticus</i>	<i>S. petrasii</i>	<i>S. stepanovicii</i>
<i>S. caprae</i>	<i>S. hominis</i>	<i>S. pettenkoferi</i>	<i>S. succinus</i>
<i>S. carnosus</i>	<i>S. lycicus</i>	<i>S. piscifermentans</i>	<i>S. vitulinus</i>
<i>S. caseolyticus</i>	<i>S. intermedius</i>	<i>S. pseudointermedius</i>	<i>S. warneri</i>
<i>S. chromogenes</i>	<i>S. kloosii</i>	<i>S. pseudolugdunensis</i>	<i>S. xylosus</i>
<i>S. cohnii</i>	<i>S. leei</i>	<i>S. pulvereri</i>	
<i>S. condimenti</i>	<i>S. lentus</i>	<i>S. rostri</i>	
<i>S. delphini</i>	<i>S. lugdunensis</i>	<i>S. saaccharolyticus</i>	
<i>S. devriesei</i>	<i>S. luriae</i>	<i>S. saprophyticus</i>	
<i>S. epidermidis</i>	<i>S. lyticans</i>		
	<i>S. massiliensis</i>		
<i>Streptococcus</i>	<i>Streptococcus infantarius</i>	<i>Streptococcus orisratti</i>	<i>Streptococcus thermophilus</i>
<i>Streptococcus agalactiae</i>	<i>Streptococcus iniae</i>	<i>Streptococcus parasanguinis</i>	<i>Streptococcus sanguinis</i>
<i>Streptococcus anginosus</i>	<i>Streptococcus intermedius</i>	<i>Streptococcus peroris</i>	<i>Streptococcus sobrinus</i>
<i>Streptococcus bovis</i>	<i>Streptococcus lactarius</i>	<i>Streptococcus pneumoniae</i>	<i>Streptococcus suis</i>
<i>Streptococcus canis</i>	<i>Streptococcus milleri</i>	<i>pseudopneumoniae</i>	<i>Streptococcus uberis</i>
<i>Streptococcus constellatus</i>	<i>Streptococcus mitis</i>	<i>Streptococcus pyogenes</i>	<i>Streptococcus vestibularis</i>
<i>Streptococcus downei</i>	<i>Streptococcus mutans</i>	<i>Streptococcus rattii</i>	<i>Streptococcus viridans</i>
<i>Streptococcus dysgalactiae</i>	<i>Streptococcus oralis</i>	<i>Streptococcus salivarius</i>	<i>Streptococcus</i>
<i>Streptococcus equinus</i>	<i>Streptococcus tigurinus</i>		<i>zooepidemicus</i>
<i>Streptococcus faecalis</i>			
<i>Streptococcus ferus</i>			

TABLE 1-continued

Optionally, the target host cells are cells of a genus or species selected from this Table and/or the production strain cells are cells of a genus or species selected from this Table	
Example Bacteria	
<i>Xenorhabdus griffithiae</i>	<i>Yersinia</i>
<i>Xenorhabdus hominickii</i>	<i>Yersinia aldovae</i>
<i>Xenorhabdus koppenhoeferi</i>	<i>Yersinia bercovieri</i>
<i>Xenorhabdus nematophila</i>	<i>Yersinia enterocolitica</i>
<i>Xenorhabdus poinarii</i>	<i>Yersinia entomophaga</i>
<i>Xylanibacter</i>	<i>Yersinia frederiksenii</i>
<i>Xylanibacter oryzae</i>	<i>Yersinia intermedia</i>
	<i>Yersinia kristensenii</i>
	<i>Yokenella regensburgi</i>
	<i>Yonghaparkia</i>
	<i>Yonghaparkia alkaliphila</i>
	<i>Zavarzinia</i>
	<i>Zavarzinia compransoris</i>
	<i>Zymomonas mobilis</i>
	<i>Zymophilus</i>
	<i>Zymophilus paucivorans</i>
	<i>Zymophilus raffinosivorans</i>
	<i>Zhihengliuella</i>
	<i>Zhihengliuella</i>
	<i>halotolerans</i>
	<i>Xylanibacterium</i>
	<i>Xylanibacterium ulmi</i>

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TABLE 2

Sequences
Nucleic acid sequences herein are written in 5' to 3' direction; amino acid sequences are written in N- to C-terminal direction. SEQ ID NO: 1 (P10) TTTCAATTTAATCATCCGGCTCGTATAATGTGTGGA
SEQ ID NO: 2 (BCD14) GGGCCCAAGTTCACCTTAAAAAGGAGATCAACAATGAAAGCAATTTTCGTA CTGAAACATCTTAATCATCGGTGGAGGTTTCTAATG
SEQ ID NO: 3 (gfp) ATGAGCAAAGGAGAAGAACTTTTCTACTGGAGTTGTC
SEQ IDs NO: 4 & 29 (example Expression Operating Unit, EOU) The EOU is (in 5' to 3' direction):- [SEQ ID NO: 4]-[promoter]-[TIS]-[GFP-encoding nucleotide sequence]-[SEQ ID NO: 29]
Where SEQ ID NO: 4 is GAATTCAAAAGATCTTAAGTAAGTAAGAGTATACGTATATCGGCTAATAA CGTATGAAGGCGCTTCGGCGCCTTTTTTTATGGGGGTATTTTCATCCCAA

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TABLE 2-continued

Sequences
5 TCCACACGTCCAACGCACAGCAAACACCACGTCGACCCTATCAGCTGCGT GCTTTCTATGAGTCGTTGCTGCATAAAGTACAAATTAATCATCCGGCTCG TATAATGTGTGGA SEQ ID NO: 29 is GGATCCAAACTCGAGTAAGGATCTCCAGGCATCAATAAAACGAAAGGCT CAGTCGAAAGACTGGGCCTTTTCGTTTTATCTGTTGTTGTCGGTGAACGC 10 TCTCTACTAGAGTCACACTGGCTCACCTTCGGGTGGCCCTTTCTGCGTTT ATA SEQ ID NO: 5 (Example Shine Dalgarno Sequence) AAAGAGGAGAAA 15 SEQ ID NO: 26 (Spacer sequence) CTTTGCCGCGCGCTTCGTCACGTAATTCCTCGTCGCAA SEQ ID NO: 27 (Spacer sequence) GTTTGGCGATGGCGCGGTGTGTTGCTTCGGCGT 20 SEQ ID NO: 28 (Spacer sequence) TGGGATGCCTACCGCAAGCAGCTTGGCCGTAA

TABLE 3

Anderson Promoter Collection			
SEQ ID NO:	Identifier	Sequence ^a	Measured Strength ^b
6	BBa J23119	TTGACAGCTAGCTCAGTCCTAGGTATAATGCTAGC	n/a
7	BBa J23100	TTGACGGCTAGCTCAGTCCTAGGTACAGTGCTAGC	1
8	BBa J23101	TTTACAGCTAGCTCAGTCCTAGGTATTATGCTAGC	0.7
9	BBa J23102	TTGACAGCTAGCTCAGTCCTAGGTACTGTGCTAGC	0.86
10	BBa J23103	CTGATAGCTAGCTCAGTCCTAGGGATTATGCTAGC	0.01
11	BBa J23104	TTGACAGCTAGCTCAGTCCTAGGTATTGTGCTAGC	0.72
12	BBa J23105	TTTACGGCTAGCTCAGTCCTAGGTACTATGCTAGC	0.24
13	BBa J23106	TTTACGGCTAGCTCAGTCCTAGGTATAGTGTAGC	0.47
14	BBa J23107	TTTACGGCTAGCTCAGCCCTAGGTATTATGCTAGC	0.36
15	BBa J23108	CTGACAGCTAGCTCAGTCCTAGGTATAATGCTAGC	0.51
16	BBa J23109	TTTACAGCTAGCTCAGTCCTAGGGACTGTGCTAGC	0.04
17	BBa J23110	TTTACGGCTAGCTCAGTCCTAGGTACAATGCTAGC	0.33
18	BBa J23111	TTGACGGCTAGCTCAGTCCTAGGTATAGTGTAGC	0.58
19	BBa J23112	CTGATAGCTAGCTCAGTCCTAGGGATTATGCTAGC	0
20	BBa J23113	CTGATGGCTAGCTCAGTCCTAGGGATTATGCTAGC	0.01
21	BBa J23114	TTTATGGCTAGCTCAGTCCTAGGTACAATGCTAGC	0.1
22	BBa J23115	TTTATAGCTAGCTCAGCCCTTGGTACAATGCTAGC	0.15
23	BBa J23116	TTGACAGCTAGCTCAGTCCTAGGGACTATGCTAGC	0.16

TABLE 3-continued

Anderson Promoter Collection			
SEQ ID NO:	Identifier	Sequence ^a	Measured Strength ^b
24	BBa J23117	TGACAGCTAGCTCAGTCCTAGGGATTGTGCTAGC	0.06
25	BBa J23118	TGACGGCTAGCTCAGTCCTAGGTATTGTGCTAGC	0.56

aalso shown in the Anderson Catalog, see parts.igem.org/Promoters/Catalog/Anderson

^bStrength is the Anderson Score (AS), e.g., a strength of 1 is a AS of 1. Reported activities of the promoters are given as the relative fluorescence of plasmids in strain TGl grown in LB media to saturation. A suitable plasmid is EX-Ptet-S-rbsRFP-P "RFP reporter" as described at parts.igem.org/Part:BBa_J61002; insertion of a promoter element between XbaI and SpeI sites results in a RFP reporter.

SEQUENCE LISTING

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ttaatcatgc ggtggagggt ttctaattg 88

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acaattaatc atccggctcg tataatgtgt ggaa 214

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gctcaccttc gggtaggcct ttctgcgttt ata          153

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The invention claimed is:

1. A production strain bacterial cell comprising a nucleic acid vector for introduction into a target bacterial host cell for expression of Type I Cas3 and Cascade proteins in the target bacterial host cell, the vector comprising a first nucleotide sequence encoding a Type I Cas3 and a second nucleotide sequence encoding one or more cognate Cascade proteins, wherein the first nucleotide sequence is under the control of a promoter for controlling the expression of Type

I Cas3 in the target bacterial host cell, wherein the promoter has a strength that is weaker than the Anderson Score strength of promoter BBa_J23108,

wherein the target bacterial host cell is selected from the group consisting of Fusobacteria, *Bacteroides*, *Staphylococcus*, *Clostridium*, *Lactobacillus*, *Bacillus*, *Escherichia*, *Streptococcus*, *Streptomyces*, *Pseudomonas*, and *Klebsiella*,

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wherein the nucleic acid vector further comprises: (i) a CRISPR array for producing crRNAs in the target bacterial host cell; or (ii) one or more nucleotide sequences encoding one or more guide RNAs (gRNA), wherein the crRNAs or gRNAs each comprise a spacer sequence complementary to a target sequence of the target bacterial host cell, and

wherein the production strain bacterial cell does not comprise a crRNA or gRNA operable with the Cas3 to target and cut a chromosomal sequence of the production strain cell.

2. The production strain bacterial cell of claim 1, wherein the nucleic acid vector comprises an operon for expression of the Type I Cas3 and Cascade proteins, and:

(a) the first nucleotide sequence is between the promoter and the second nucleotide sequence in the operon;

(b) the operon comprises no Cas-encoding nucleotide sequences between the promoter and the first nucleotide sequence; or

(c) the operon comprises, in 5' to 3' direction, the promoter, the first nucleotide sequence, and the second nucleotide sequence.

3. The production strain bacterial cell of claim 1, wherein the promoter is a constitutive promoter.

4. The production strain bacterial cell of claim 1, wherein the promoter is repressible.

5. The production strain bacterial cell of claim 1, wherein the promoter has a strength that is greater than the Anderson Score strength of promoter BBa_J23114.

6. The production strain bacterial cell of claim 1, further comprising an origin of replication that is operable in the target bacterial host cell.

7. The production strain bacterial cell of claim 1, wherein the nucleic acid vector is devoid of a Cas adaption module.

8. The production strain bacterial cell of claim 1, wherein the nucleic acid vector is devoid of a nucleotide sequence encoding one or more of a Cas1, Cas2, Cas4, Cas6, Cas7, and Cas8.

9. The production strain bacterial cell of claim 1, wherein the second nucleotide sequence encodes one or more of (a)-(g):

(a) Cas11, Cas7, and Cas8a1;

(b) Cas8b1, Cas7, and Cas5;

(c) Cas5, Cas8c, and Cas7;

(d) Cas8U2, Cas7, Cas5, and Cas6;

(e) Cas10d, Cas7, and Cas5;

(f) Cas8e, Cas11, Cas7, Cas5, and Cas6; and

(g) Cas8f, Cas5, Cas7, and Cas6f.

10. The production strain bacterial cell of claim 9, wherein the Type I Cas3 is a Cas3' or Cas3".

11. The production strain bacterial cell of claim 9, wherein the Type I Cas3 is a Cas3, Cas3' or Cas3", and wherein the Type I Cas3 is between the promoter and the second nucleotide sequence.

12. The production strain bacterial cell of claim 11, wherein the nucleic acid vector is devoid of a nucleotide sequence encoding a further Cas between the promoter and the Type I Cas3.

13. The production strain bacterial cell of claim 9, wherein the vector comprises the CRISPR array, the CRISPR array is cognate with the Type I Cas3, and wherein:

(a) the CRISPR array is a Type IA array and the nucleic acid vector comprises Cas11, Cas7, and Cas8a1;

(b) the CRISPR array is a Type IB array and the nucleic acid vector comprises Cas8b1, Cas7, and Cas5;

(c) the CRISPR array is a Type IC array and the nucleic acid vector comprises Cas5, Cas8c, and Cas7;

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(d) the CRISPR array is a Type IU array and the nucleic acid vector comprises Cas8U2, Cas7, Cas5, and Cas6;

(e) the CRISPR array is a Type ID array and the nucleic acid vector comprises Cas10d, Cas7, and Cas5;

5 (f) the CRISPR array is a Type IE array and the nucleic acid vector comprises Cas8e, Cas11, Cas7, Cas5, and Cas6; or

(g) the CRISPR array is a Type IF array and the nucleic acid vector comprises Cas8f, Cas5, Cas7, and Cas6f.

14. The production strain bacterial cell of claim 1, wherein the Type I Cas3 and Cascade are:

(a) Type IA Cas and Cascade proteins;

(b) Type IB Cas and Cascade proteins;

(c) Type IC Cas and Cascade proteins;

(d) Type ID Cas and Cascade proteins;

(e) Type IE Cas and Cascade proteins;

(f) Type IF Cas and Cascade proteins; or

(g) Type IU Cas and Cascade proteins.

15. The production strain bacterial cell of claim 1, wherein the Type I Cas3 and Cascade are *E. coli* Cas and Cascade proteins.

16. The production strain bacterial cell of claim 1, wherein the promoter is operable in a target host cell selected from: an ESBL-producing *E. coli* or *E. coli* ST131-O25b:H4; *C. difficile* resistant to one or more antibiotics selected from aminoglycosides, lincomycin, tetracyclines, erythromycin, clindamycin, penicillins, cephalosporins and fluoroquinolones; *P. aeruginosa* resistant to one or more antibiotics selected from carbapenems, aminoglycosides, cefepime, ceftazidime, fluoroquinolones, piperacillin and tazobactam; carbapenem-resistant *Klebsiella pneumoniae*; and an Extended-Spectrum Beta-Lactamase (ESBL)-producing *K. pneumoniae* cell.

17. The production strain bacterial cell of claim 16, wherein the Type I Cas3 and Cascade are *E. coli*, *C. difficile*, *P. aeruginosa*, *K. pneumoniae*, *P. furiosus*, or *B. halodurans* Cas and Cascade proteins.

18. The production strain bacterial cell of claim 1, wherein the Type I Cas3 and Cascade are *E. coli*, *C. difficile*, *P. aeruginosa*, *K. pneumoniae*, *P. furiosus*, or *B. halodurans* Cas and Cascade proteins.

19. The production strain bacterial cell of claim 1, wherein the Type I Cas3 is a Cas3 of a CRISPR/Cas locus of *E. coli*, and wherein the distance between the Cas3-encoding sequence of the locus and its cognate promoter in *E. coli* is further than the distance between the Cas3-encoding sequence and the promoter for controlling the expression of Type I Cas3 in the nucleic acid vector.

20. The production strain bacterial cell of claim 1, wherein the CRISPR array or the gRNA-encoding sequence(s) are under the control of a second promoter that is different from the promoter that controls the expression of the Type I Cas3.

21. The production strain bacterial cell of claim 1, wherein the nucleic acid vector is a plasmid or phagemid.

22. The production strain bacterial cell of claim 1, wherein the production strain bacterial cell comprises a nucleotide sequence whose expression is inducible to produce phage coat proteins in the cell of the production strain, wherein the production strain bacterial cell comprises amplified copies of the nucleic acid vector,

wherein the production strain bacterial cell is capable of packaging the amplified copies of the nucleic acid vector into phage particles or non-self-replicative transduction particles for introducing the amplified copies of the nucleic acid vector into the target host cell.

23. The production strain bacterial cell of claim 22, wherein the nucleic acid vector is a plasmid or phagemid and the delivery vehicle is a non-replicative transduction particle.

24. The production strain bacterial cell of claim 1, 5 wherein the second nucleotide sequence is under the control of the same promoter as the first nucleotide sequence.

25. The production strain bacterial cell of claim 1, wherein the target sequence of the target bacterial host cell is a chromosomal sequence of the target bacterial host cell. 10

26. The production strain bacterial cell of claim 1, wherein the production strain bacterial cell is an *Escherichia coli* (*E. coli*) cell.

* * * * *